



The Evolving Landscape of the Colorectal Cancer Vaccines: From Biological Mechanisms to Translational Therapeutics

Sabrina Brigitta Valerie Setiono ¹, Grace Lendawati Amelia Turalaki ², and Trina Ekawati Tallei ^{2,3,*}

¹ Faculty of Medicine, Sam Ratulangi University, Manado 95115, Indonesia; sabrinasetiono011@student.unsrat.ac.id (S.B.V.S.)

² Department of Biology, Faculty of Medicine, Sam Ratulangi University, Manado 95115, Indonesia; trina_tallei@unsrat.ac.id (T.E.T.); gracelat@unsrat.ac.id (G.L.A.T.)

³ Department of Biology, Faculty of Mathematics and Natural Sciences, Sam Ratulangi University, Manado 95115, Indonesia

* Correspondence: trina_tallei@unsrat.ac.id

Article History

Received 13 December 2025
Revised 7 February 2026
Accepted 19 February 2026
Available Online 27 February 2026

Keywords:

Colorectal carcinoma
Immunotherapy
Therapeutic vaccination
Tumor antigens
Immunotherapy advancements

Abstract

Colorectal cancer (CRC) remains a major global health burden, and despite substantial advances in cancer immunotherapy, the clinical efficacy of therapeutic cancer vaccines in CRC has been limited. This review critically examines the biological, immunological, and translational factors that shape CRC vaccine development, with a particular focus on tumor immunopathology, antigen selection, vaccine platforms, and emerging combination strategies. We summarize current knowledge on CRC-associated tumor antigens, including selected tumor-associated antigens and neoantigen-based approaches, alongside major vaccine modalities evaluated in preclinical and early-phase clinical studies. Across the literature, vaccine-induced immunogenicity frequently exceeds demonstrated clinical benefit, highlighting a persistent translational gap. Synthesis of available evidence suggests that this gap is driven primarily by CRC-specific immune constraints, including immune exclusion, dominance of immunologically cold MSS/pMMR tumors, and tolerogenic pressures within metastatic niches, particularly the liver. We further discuss how rational combination strategies, especially those integrating cancer vaccines with immune checkpoint inhibitors (ICIs), may partially overcome these barriers. In addition, the review outlines the conceptual role of bioinformatics and immunoinformatics in supporting antigen prioritization, neoantigen discovery, and patient stratification in CRC vaccine research. Overall, this review emphasizes that future progress will depend on CRC-tailored antigen selection, mechanistically informed vaccine design, rational combination regimens, and rigorous clinical evaluation to define the realistic clinical role of therapeutic cancer vaccines in CRC.



Copyright: © 2026 by the authors. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License. (<https://creativecommons.org/licenses/by-nc/4.0/>)

1. Introduction

Colorectal cancer (CRC) is one of the most common malignancies worldwide and remains a major cause of cancer-related mortality [1–3]. According to global cancer statistics published by the International Agency for Research on Cancer (IARC) in 2022, CRC ranks as the second leading cause of cancer-related deaths and the third most frequently diagnosed cancer globally. The

global burden of CRC is projected to rise by nearly 60% by 2030, reaching more than 2.2 million new cases and approximately 1.1 million deaths annually [4, 5]. This rising incidence, particularly in developing countries, is strongly associated with lifestyle transitions toward Westernized behavioral patterns, including higher rates of obesity, physical inactivity, high consumption of red meat, alcohol, and tobacco, as well as high-calorie and low-fiber diets [6–12].

Despite notable progress in therapeutic modalities, clinical outcomes for CRC remain unsatisfactory, especially among patients with metastatic disease [3]. Although surgical resection, chemotherapy, radiotherapy, biological agents, immunotherapy, and various combination regimens are widely used and have received regulatory approval, their clinical performance remains constrained by therapeutic resistance, significant adverse effects, and limited long-term effectiveness. These shortcomings highlight the urgent need for more effective and durable therapeutic strategies [1, 13].

Immunotherapy has gained increasing prominence as a promising alternative to conventional cytotoxic chemotherapy due to its ability to restore and augment immune competence, which is often severely impaired in cancer patients undergoing standard treatments [14]. Immune checkpoint inhibitors (ICIs), adoptive cell therapy (ACT), and cancer vaccines represent some of the most widely investigated and impactful immunotherapeutic modalities to date [15]. However, the efficacy of ICIs and ACT remains confined to a subset of patients, with durable responses observed in only a minority [16]. Even though, monoclonal antibodies targeting ICIs have transformed the therapeutic landscape across a range of solid tumors and hematologic malignancies, their therapeutic activity is limited to specific patient subsets [17]. ICIs generally show limited efficacy in tumors lacking sufficient tumor-infiltrating lymphocytes (TILs) and are frequently associated with immune-related adverse events that may compromise patient quality of life and adherence to therapy [18]. Collectively, these limitations highlight the importance of therapeutic approaches that can facilitate the initiation or amplification of tumor-specific immune responses beyond reliance on pre-existing immunity alone.

Within this context, vaccine-based immunotherapy has emerged as a compelling therapeutic avenue because of their unique ability to prime and expand antigen-specific immune responses *de novo*. Vaccination remains one of the most successful medical interventions in human history, including within oncology [14]. Cancer vaccines offer unique advantages, particularly for patients who are refractory to other therapies, by inducing broad, antigen-specific, and durable T-cell responses targeting a diverse repertoire of tumor antigens [18, 19]. Unlike other treatment modalities, vaccines are designed to train the host immune system to recognize and eliminate tumor cells with heightened specificity [20–22].

Nevertheless, the success of immunotherapy continues to be hindered by the profoundly immunosuppressive nature of the tumor microenvironment (TME), which

suppresses endogenous immunity and limits the efficacy of multiple immunotherapeutic platforms [23]. Immune responses elicited by therapeutic vaccines can be attenuated by immunoregulatory mechanisms within the TME [18]. In CRC, accumulating evidence demonstrates that key TME characteristics, most notably the density and functional competence of cytotoxic T lymphocyte (CTL) infiltration, serve as critical determinants of therapeutic outcomes. These observations underscore the need for innovative vaccine-based strategies capable of eliciting robust and sustained anti-tumor immune responses [24, 25]. Effective anti-tumor immunity plays a critical role in suppressing tumor initiation, growth, progression, and metastatic dissemination, thereby facilitating meaningful clinical responses [26].

Building upon the substantial advances achieved with ICIs, integrating these agents with therapeutic cancer vaccines offers a compelling scientific rationale for amplifying anti-tumor immunity [27, 28]. This rationale stems from the fundamentally distinct, yet highly synergistic, mechanisms through which cancer vaccines and ICIs modulate the cancer-immunity cycle. Therapeutic cancer vaccines function by priming and expanding antigen-specific T cells, thereby enhancing the activation and intratumoral infiltration of cytotoxic immune cells. In contrast, ICIs operate principally by overcoming inhibitory signals that suppress effector function, thus preventing or reversing the dysfunction and exhaustion of these effector T-cells within the TME. Although both modalities have demonstrated clinical activity as monotherapies, each faces substantial limitations, as previously discussed. The convergence of these complementary mechanisms provides a strong mechanistic foundation for therapeutic synergy. By driving potent antigen-specific immune priming while simultaneously relieving inhibitory signaling pathways, vaccine-ICI combinations have the potential to generate more durable, comprehensive, and clinically meaningful anti-tumor responses than either approach alone [18].

The review by Gallio et al. provides a valuable overview of CRC vaccine platforms, including recent updates in vaccine technologies, and highlights the emerging potential of combining cancer vaccines with ICIs. The authors appropriately recognize that such combination strategies may enhance therapeutic efficacy compared with vaccine or ICI monotherapy. However, the review does not provide a comprehensive mechanistic explanation of why vaccine-ICI combinations are biologically synergistic and how these combinations mechanistically overcome the immunological barriers that limit vaccine efficacy in CRC [29].

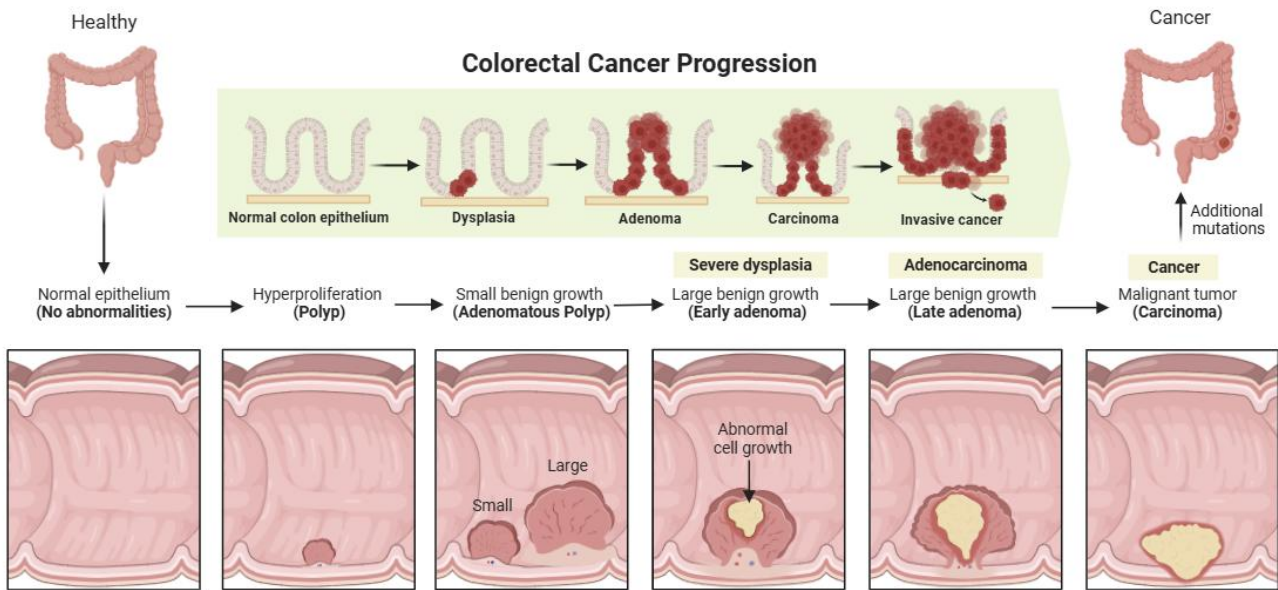


Figure 1. CRC progression and metastasis. CRC, Colorectal cancer.

In contrast, the present review is specifically designed to address these unresolved mechanistic gaps. This review aims to provide an overview of the evolution and current status of CRC vaccine development, highlighting recent advances in vaccine engineering, ongoing clinical challenges, and emerging strategies aimed at improving vaccine efficacy and accelerating translational potential. We also provide an in-depth analysis of the fundamental immunological mechanisms that underpin vaccine-ICI synergy, linking vaccine-induced antigen priming and T-cell expansion with checkpoint blockade-mediated reinvigoration of effector function within the TME. By integrating CRC-specific immunopathology with current knowledge of immune checkpoint biology, this review establishes a clearer biological rationale for rational vaccine-ICI combination strategies and offers a more mechanistically grounded framework to guide future translational and clinical development.

To clearly define its scope, this review focuses on therapeutic (rather than prophylactic) cancer vaccines for CRC, drawing primarily on preclinical and clinical (if available) studies without any restriction to the year of publication. We cover major vaccine platforms currently under investigation in CRC, including peptide-based, dendritic cell-based, nucleic acid-based, viral vector-based, and cancer cell-based vaccine.

2. Immunopathology of CRC

The development of CRC typically follows the classical adenoma-carcinoma-metastasis sequence. Aberrant crypt foci gradually evolve into benign adenomatous polyps, which may subsequently progress into sporadic CRC (Figure 1) [30]. As a multifactorial disease, CRC arises

through a continuum of histopathological changes in the colonic epithelium, including hyperplasia, varying degrees of dysplasia (mild, moderate, or severe), and adenoma formation. These transitions are driven by the stepwise accumulation of structural DNA alterations induced by carcinogenic stimuli, ultimately culminating in malignant transformation [31]. Malignant transformation is generally characterized by three major stages: (1) acquisition of driver mutations within epithelial or stem cells, yielding genetic alterations that give rise to mutant cancer-initiating cells; (2) clonal fixation, during which mutant cells outcompete and replace their wild-type counterparts within the crypt; and (3) clonal expansion via crypt fission, enabling the spread and establishment of the mutant clone [30, 32]. At the molecular level, CRC arises through three principals, and often overlapping, genetic pathways: chromosomal instability (CIN), DNA mismatch repair (MMR) deficiency, and the CpG island methylator phenotype (CIMP) [31, 33–38].

As the second leading cause of cancer-related mortality worldwide, metastasis remains the predominant determinant of CRC lethality [39, 40]. Accumulating evidence demonstrates that signaling pathways within the TME critically shape both the progression and dissemination of CRC [41]. The TME constitutes a dynamic and highly interactive milieu that influences virtually all stages of tumor evolution, from initial tumor cell adhesion and early growth to mechanisms enabling immune evasion. It comprises a heterogeneous network of malignant cells and non-malignant constituents, including immune cells, fibroblasts, endothelial cells, and a complex extracellular matrix. Although each component possesses distinct biological functions, their

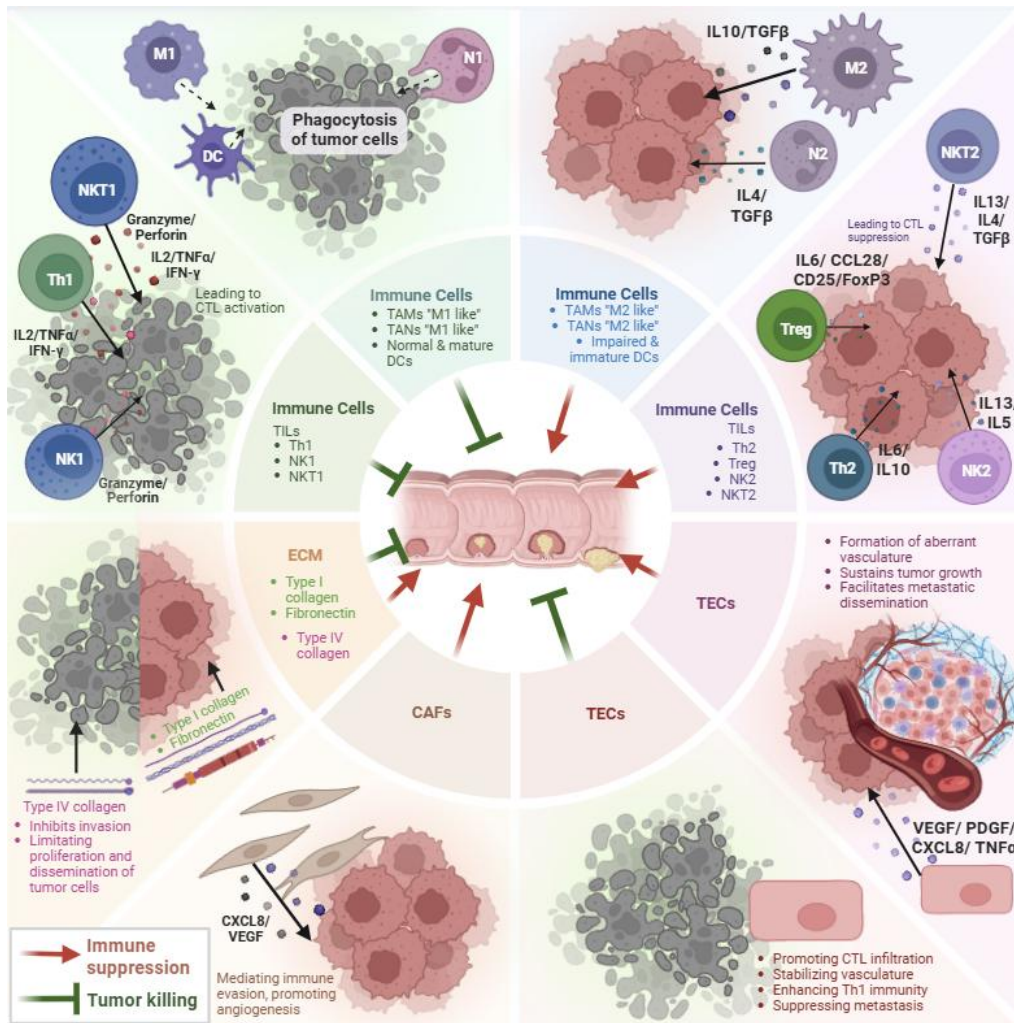


Figure 2. TME components and CRC. Both cellular and non-cellular elements of the TME may act as either promoters or suppressors of tumor progression, depending on the biological context. CAFs, Cancer-associated fibroblasts; CRC, Colorectal cancer; CTL, Cytotoxic T cell; DC, Dendritic cell; ECM, extracellular matrix; M, Macrophage; N, Neutrophil; NK, Natural killer; NKT, Natural killer T; TAMs, Tumor-associated macrophages; TANs, Tumor-associated neutrophils; TECs, Tumor-associated endothelial cells; Th, T-helper; TILs, Tumor-infiltrating lymphocytes; TME, Tumor microenvironment; Treg, T-regulator.

reciprocal interactions collectively orchestrate tumor progression, metastatic competence, and resistance to immune-mediated elimination (Figure 2). This intricate crosstalk underscores the profound complexity of CRC immunopathology and highlights the central role of the TME in dictating disease trajectory [40, 42].

Importantly, cancer progression is not solely dictated by tumor-intrinsic characteristics but is profoundly shaped by continuous interactions between malignant cells and host immune responses within the TME [43, 44]. Notably, the CRC TME exhibits pronounced immune spatial heterogeneity, whereby immune cell composition, density, and functional states vary across distinct tumor regions, including the tumor core (main concentration of tumor cells), tumor stroma, invasive margin (transition zone between tumor cells and normal cells), and tertiary lymphoid structures (TLS), thereby critically determines prognosis and therapeutic responsiveness. The tumor

core contains the bulk of malignant cells, so that immune cells residing within tumor core display more direct and efficient interaction with tumor cells [45]. The tumor core tends to establish an immunosuppressive microenvironment characterized by hypoxia, dense extracellular matrix (ECM), and enrichment of regulatory immune populations, limit effective infiltration and function of CTLs. Surrounding this region is the invasive tumor margin represents a critical immunological interface in antitumor defense, thereby characterized by a markedly higher density of infiltrating immune cells, including T cells, B cells, natural killer (NK) cells, and dendritic cells (DCs), compared with other tumor regions [46]. In contrast, immune cells localized within the tumor stroma, located peripheral to the tumor core, are strongly associated with stromal remodeling and angiogenic processes, composed of abundant stromal elements that support tumor growth by providing structural support and metabolic resources, thereby

exerting a substantial impact on tumor growth, invasion, and metastatic potential [47, 48].

TLS are ectopic aggregates of immune cells that arise in non-lymphoid tissues, most prominently within the TME. Structurally analogous to secondary lymphoid organs, TLS displays organizational and functional features analogous to those of fully developed lymphoid organs, largely orchestrated by the coordinated interactions among T-/B-cells, DCs, germinal centers, and high endothelial venules (HEVs) within its architecture. TLS have been identified across multiple solid malignancies, including CRC, where their presence is frequently associated with favorable clinical outcomes [49–51]. Pathological investigations have demonstrated that tertiary lymphoid structures located within the mucosa of CRC patients display a high degree of structural organization, with well-defined T-cell and B-cell compartments, suggesting their capacity to sustain effective local immune responses [52]. The development is driven by chronic inflammation via cytokine- and chemokine-mediated signaling, particularly involving molecules such as CXCL13 and CCL21, along with the formation of high endothelial venules (HEVs). Through these coordinated processes, TLS facilitate localized aggregation and interaction of immune cells within the TME, enabling more efficient activation of adaptive immune responses compared with unstructured or diffuse immune cell infiltration [53, 54]. Lv et al. demonstrated that increased TLS density and a higher degree of structural maturation correlate with reduced recurrence rates and prolonged patient survival. Moreover, emerging data suggest a functional interplay between TLS and immune checkpoint inhibition, underscoring the therapeutic potential of targeting or leveraging TLS to enhance antitumor immunity in CRC [54].

Although multiple innate and adaptive immune populations actively participate in tumor-immune regulation, current American Joint Committee on Cancer (AJCC)/Union for International Cancer Control (UICC)-tumour, node and metastasis (TNM) staging follows a predominantly tumor-centric paradigm, in which prognostic assessment is based almost exclusively on anatomical and pathological features such as depth of invasion and lymph node involvement [55, 56]. TNM classification offers limited prognostic resolution, as clinical outcomes can differ markedly among patients classified within the same histopathological tumor stage, despite its considerable utility [57]. In contrast, immune effector cells, including CTLs, B cells, and macrophages, can mediate tumor control and elimination, whereas immunosuppressive populations such as Treg cells

attenuate antitumor immunity and facilitate tumor progression and tissue invasion [58]. Studies demonstrated that the type, density, and spatial distribution of immune cells within CRC tumors provide superior prognostic value compared with conventional TNM staging. This immune contexture concept has been translated into a clinically applicable immune-based classification system, known as the Immunoscore [43, 59–61]. The Immunoscore quantitatively assesses intratumoral (tumor core) and invasive-margin T-cell infiltration and has been validated as a robust prognostic marker in CRC [43, 59, 62–64]. Importantly, the Immunoscore not only represents one of the strongest predictors of patient survival, but also offers a biological framework that is directly relevant to immunotherapeutic intervention [25]. The presence of naturally infiltrating T cells suggests a pre-existing immune response that may be further amplified by novel immunotherapy approaches.

Growing insights into the TME, including baseline antitumor immunity, and its critical role in CRC progression underscore substantial opportunities for TME-targeted therapeutic strategies, particularly in the management of metastatic CRC (mCRC). ICIs such as pembrolizumab and nivolumab, which block the PD-1 pathway, have been approved for use in patients with deficient mismatch repair (dMMR) or high microsatellite instability (MSI-H) [65, 66]. Despite their clinical success, only a minority of CRC patients derive meaningful benefit from these agents because MSI-H tumors account for approximately 15% of all CRC cases, whereas the remaining ~85% exhibit proficient mismatch repair (pMMR) or microsatellite-stable (MSS) phenotypes, which are characteristically unresponsive to current ICIs [67, 68]. This disparity in therapeutic responsiveness is largely attributed to distinct immunogenic profiles. MSI-H/dMMR tumors possess a high tumor mutational burden (TMB), generating abundant neoantigens that promote robust TIL recruitment and enhance intrinsic immunogenicity. Conversely, pMMR/MSS tumors exhibit substantially lower TMB, resulting in reduced neoantigen availability, weaker TIL infiltration, and consequently an attenuated antitumor immune response [69].

2.1. Immune Cells: Macrophages

Macrophages, a major myeloid cell subset, exert context-dependent effects in CRC. While they can suppress tumor cell proliferation, migration, invasion, and metastasis, macrophages may also promote tumor development and drive therapeutic resistance [70]. Tumor-associated macrophages (TAMs) are broadly categorized into classically activated “M1-like” and alternatively activated “M2-like” phenotypes [71]. Under physiological immune

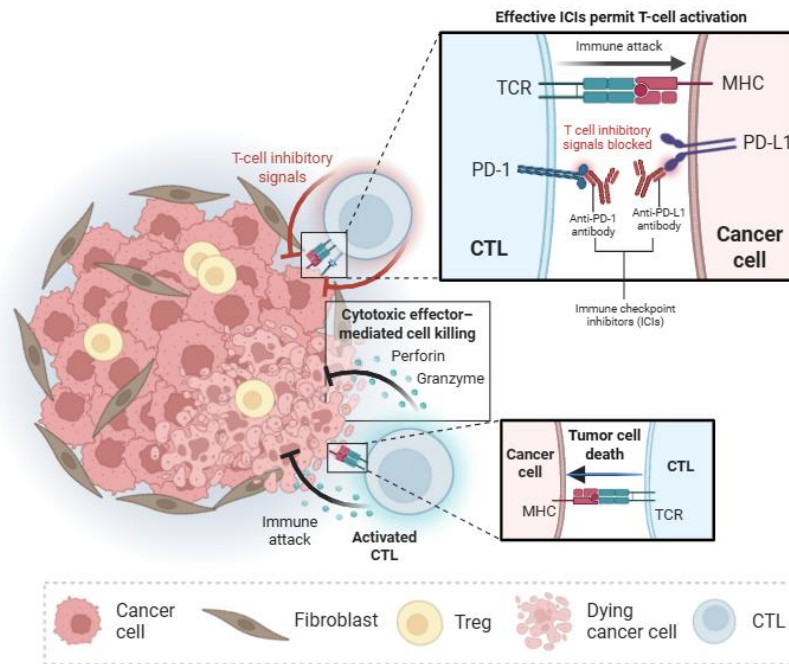


Figure 3. Immune evasion orchestrated by the PD-1/PD-L1 pathway in tumors. CRC upregulate PD-L1 to inhibit cytotoxic T-cell activity, but immune checkpoint inhibitors targeting PD-1/PD-L1 disrupt this suppressive interaction. Once released from inhibition, T cells regain effector function and discharge cytotoxic mediators such as perforin and granzymes, leading to targeted tumor cell elimination. CRC, Colorectal cancer; CTL, Cytotoxic T cell; ICIs, Immune checkpoint inhibitors; MHC, Major histocompatibility complex; PD; Programmed death; TCR, T-cell receptor; Treg, T-regulator.

conditions, macrophages predominantly adopt an M1 phenotype characterized by proinflammatory and immunostimulatory activity, which supports Th1 cytokine responses and inhibits CRC progression [72]. However, the anti-tumor role of M1 macrophages is not absolute. Chronic M1-driven inflammation, such as in colitis, has been linked to increased CRC risk, illustrating the dualistic nature of inflammation in colorectal tumorigenesis [73].

Sustained inflammation and/or infection contributes to an immunosuppressive TME by recruiting regulatory T cells (Tregs) and myeloid-derived suppressor cells or by skewing immune populations toward pro-tumorigenic states [74, 75]. Chronic inflammation further exacerbates genomic instability and induces the upregulation of immune checkpoints such as programmed death-ligand 1 (PD-L1), collectively dampening immune surveillance [76–80]. PD-L1 plays a pivotal role in restraining anti-tumor immunity by engaging the inhibitory receptor PD-1 on activated T cells, resulting in diminished proliferation, attenuated cytokine production, and impaired cytolytic activity, thereby enabling malignant cells to evade immune surveillance and defend themselves against immunological attack (Figure 3) [81–83]. These insights underscore the intricate interplay between inflammatory signaling and immunosuppression in CRC and reinforce the need for vaccines capable of generating robust immune responses while overcoming regulatory constraints.

Conversely, M2 macrophages promote malignant progression by: (1) secreting epidermal growth factor (EGF) and fibroblast growth factor-1 (FGF-1) to stimulate tumor cell proliferation; (2) releasing vascular endothelial growth factor A (VEGF-A) to enhance angiogenesis and suppress apoptosis; and (3) producing matrix metalloproteinases (MMPs) that facilitate invasion and metastasis [84–94]. M2 macrophages also induce epithelial-mesenchymal transition (EMT) through the IL-6/JAK2/STAT3 and PI3K/AKT pathways, increasing CRC cell motility and invasiveness [93, 95]. Through secretion of IL-10 and transforming growth factor- β (TGF- β), M2 macrophages suppress T-cell function, thereby accelerating tumor growth and contributing to aggressive CRC phenotypes [96, 97]. Increased immunosuppressive myeloid populations, including M2 macrophages, are a prominent feature of pMMR/MSS tumors and play a key role in driving immune exclusion and resistance to ICIs. In contrast, MSI-H/dMMR tumors are characterized by a high TMB, resulting in abundant neoantigen generation and a more “inflamed” TME enriched with infiltrating immune cells such as CTLs and M1-like macrophages, which are associated with improved anti-tumor immunity and better responses to ICIs [98–100].

Collectively, these immunopathological features highlight TAMs as critical regulators of immune suppression and therapeutic resistance in CRC, with important implications for cancer vaccine design. The

predominance of M2-like macrophages and their immunosuppressive mediators underscore the need for vaccines that elicit strong cytotoxic T-cell responses capable of overcoming macrophage-driven inhibition. Moreover, the central role of PD-L1 expression and macrophage-mediated T-cell suppression provides a strong mechanistic rationale for combining therapeutic cancer vaccines with ICIs or macrophage-modulating agents, either through the incorporation of immunostimulatory adjuvants or nanoparticle-based delivery systems. Another potential is to combine the vaccine with colony-stimulating factor-1 receptor (CSF-1R) inhibitors, which have been shown in preclinical CRC models to deplete M2 macrophages, enhance anti-tumor immunity by increasing CTLs, and enhance response to immunotherapy. The blockade of the CSF-1/CSF-1R pathway reprograms macrophages from M2 to a more immunostimulatory phenotype and enhances anti-PD-1 therapy efficacy in CRC mouse models [101–103]. Such combination strategies have the potential to simultaneously enhance antigen-specific T-cell priming while alleviating immunosuppressive barriers within the TME.

2.2. Immune Cells: Lymphocytes

Tumor-infiltrating lymphocytes (TILs), comprising helper T lymphocytes (HTLs), CTLs, B cells, and NK cells, play pivotal roles in tumor recognition, immune surveillance, and immune escape [104]. Ropponen et al. demonstrated an inverse correlation between TIL density and tumor stage, noting significantly fewer TILs in late-stage (Dukes C–D) CRC compared to early-stage disease (Dukes A–B) [105]. Kuwahara et al. reported that elevated HTL infiltration correlates with improved survival in CRC, although HTL subsets exert divergent effects [106, 107]. Th1-polarized HTLs promote cytotoxic immunity and are associated with favorable prognosis, whereas Th2 responses tend to support tumorigenesis [108]. Elevated Th17 cell levels are linked to poorer outcomes, with Th17-derived cytokines (IL-17, IL-21, IL-22) driving CRC growth and progression [109, 110].

High intratumoral CTL density, representing the central effector population of the adaptive immune system and critical mediators of anti-tumor immunity, correlates strongly with decreased recurrence and enhanced survival [61, 111–113]. Nevertheless, CRC cells employ multiple mechanisms to evade CTL-mediated cytotoxicity. One prominent pathway involves TNF- α -driven upregulation of PD-L1, which suppresses CTL activation and diminishes the cytolytic capacity of granzyme-producing CTLs [114]. Elevated PD-L1 expression is consistently associated with advanced tumor stage, lymphatic involvement, distant metastasis,

and poorer overall survival [115–117]. Moreover, additional TME-derived immunosuppressive factors, particularly TGF- β , further compromise CTL effector function by downregulating perforin, granzymes, and Fas ligand (FasL), collectively attenuating CTL-mediated tumor cell killing [118].

The accumulation of Tregs, central mediators of immunosuppression, are strongly associated with heightened metastasis and poorer clinical outcomes [119–121]. Tregs are characterized by high expression of CD25 and the transcription factor forkhead box P3 (FoxP3), both of which are essential for their suppressive activity on effector T cells [122–125]. Under physiological conditions, Tregs maintain immune homeostasis and prevent autoimmunity by suppressing IL-2 production, releasing adenosine, and secreting immunoregulatory cytokines such as TGF- β , IL-10, and IL-35 [126]. However, within the CRC TME, Treg-mediated effects are context-dependent, reflecting a functional and phenotypic heterogeneity that contributes to both pro- and anti-tumor outcomes [127–129]. During malignant transformation, TGF- β signaling undergoes changes that resulting in tumor promotor function rather than a suppressor [130]. Then, these aberrant TGF- β activation and signaling promote tumor progression through the induction of EMT, angiogenesis, cancer-associated fibroblasts (CAFs) activation, and immunosuppression within the TME [131, 132]. Not only EMT plays a central role in cancer cell migration and invasion, EMT has also been associated with resistance to PD-L1 blockade, targeted therapies, and conventional chemotherapy [133–135]. In addition, dysregulated TGF- β signaling promotes tumor angiogenesis through upregulation of pro-angiogenic mediators, leading to increased tumor vascularization and growth [136]. TGF- β further drives the differentiation of fibroblasts into CAFs, which contribute to therapeutic resistance and metastasis by remodeling the ECM, restricting T-cell infiltration, and secreting immunosuppressive and pro-tumorigenic factors. In the TME [135, 137], TGF- β drives phenotypic reprogramming of multiple immune cell populations, including DCs, macrophages, neutrophils, NK cells, Tregs, and CTLs, to a tolerogenic phenotype [131]. The ECM and TME populations, including CAFs, DCs, neutrophils, will be further explained in the next subsections.

Saito et al. delineated three Treg subsets in CRC: naive Tregs (FoxP3^{low}/CD45RA⁺), terminally suppressive Tregs (FoxP3^{high}/CD45RA⁻), and a FoxP3^{low}/CD45RA⁻ population exhibiting pro-inflammatory characteristics. Notably, CRC tissues demonstrated a disproportionately higher frequency of FoxP3^{low}/CD45RA⁻ inflammatory Tregs compared

with other malignancies, such as melanoma. Based on these immunological profiles, CRC was further categorized into two immunological subtypes: “type A,” characterized by low levels of inflammatory Tregs, and “type B,” distinguished by abundant inflammatory Tregs. Type B tumors displayed marked upregulation of inflammation- and immunity-associated genes, including IL-12, TNF- β , and TGF- β , suggesting a more immunologically active microenvironment. Conversely, in type A CRC, high FOXP3 expression correlated with inferior clinical outcomes, underscoring the prognostic relevance of Treg phenotypic diversity in shaping disease behavior [138].

In addition to the cellular complexity described above, the molecular classification of CRC into MSI-H/dMMR and MSS/pMMR subtypes profoundly influences lymphocyte infiltration patterns, TME composition, and clinical responsiveness to immunotherapy as explained previously. Tumors with dMMR/MSI-H characterized by abundant TILs, including activated CTLs and Th1-polarized HTLs, making these tumors sensitive to treatment with ICIs while pMMR/MSS tumors are characterized by reduced immune cell infiltration. These distinct immune landscapes underscore the need for CRC vaccine strategies tailored to the molecular and immunological context of each subtype. This includes enhancing TIL recruitment and functional competence, amplifying intratumoral immune activation, and converting immunologically “cold” pMMR/MSS tumors into immune-reactive states, while leveraging the existing inflamed milieu in dMMR/MSI-H tumors to sustain effective antitumor immunity [139–142].

These immunopathological insights indicate that successful CRC vaccine strategies, including optimization of delivery platforms and synergistic combination regimens, should be guided by the functional composition and phenotypic quality of TILs, rather than by immune cell abundance alone. Preclinical evidences support dual targeting of TGF- β and immune checkpoint (PD-L1) as a strategy to overcome complementary immunosuppressive mechanisms in the TME, where simultaneous co-inhibition of these pathways has been shown to significantly enhanced antitumor immune responses by relieving TGF- β -mediated immune suppression and enhancing CTLs access and effector function beyond what is achieved with single agent alone [143, 144].

2.3 Immune Cells: APCs and Dendritic Cells

Antigen-presenting cells (APCs) are indispensable for initiating adaptive immunity by processing and presenting tumor antigens to T cells [145, 146]. Mature

DCs, the most potent APC subset whose development is driven by Flt3L, are associated with favorable clinical outcomes when present in high numbers due to its central role to the initiation, coordination, and amplification of antitumor immune responses [146–150]. DCs promote the infiltration cytotoxic activity of Th1 cells, NK cells, and CTLs by facilitating their migration through CCR7 expression, upregulating costimulatory molecules such as CD80, CD83, and CD86, and secreting pro-inflammatory cytokines including IL-12, IL-6, TNF- α , and IL-1 β [151–153].

DCs are broadly classified into two primary subsets: plasmacytoid DCs (pDCs) and classical DCs (cDCs) [154]. pDCs are characterized by their capacity to secrete large quantities of type I interferons (IFNs) upon viral stimulation and are critically involved in the regulation of immune tolerance, while cDCs are particularly efficient in antigen presentation and are further subdivided into CD141⁺ cDC1 and CD1c⁺ cDC2. CD141⁺ cDC1 specialize in cross-presenting tumor antigens via major histocompatibility complex (MHC) class I to prime CTLs, whereas CD1c⁺ cDC2 predominantly present antigens through MHC class II to activate HTLs [155, 156]. cDCs are pivotal to antitumor immune responses through their capacity to process and present TAAs to naïve T cells, but also function as central coordinators of antitumor immunity by engaging multiple immune cell populations, including T cells, NK cells, macrophages, and B cells [157]. Therefore, their capacity to prime T cells and facilitate the recruitment of additional immune effectors into the TME is critical for effective anticancer immune response [158]. Consequently, dynamic crosstalk between cDCs and other immune components within the TME plays a decisive role in shaping the efficacy of cancer vaccines, as effective antigen presentation and T-cell priming are prerequisites for translating vaccine-induced immunogenicity into durable antitumor responses [159].

cDCs counteract tumor immune evasion by processing and presenting tumor antigens and by priming CTLs that selectively eliminate malignant cells. In addition, cDCs regulate the equilibrium between immune tolerance and immune activation within the TME, a balance that is critical for effective antitumor immunity [158, 160]. Owing to their widespread distribution across peripheral blood, lymphoid tissues, and peripheral organs, cDCs are highly efficient at capturing TAAs, cross-presenting exogenous antigens via MHC class I, and migrating to draining lymph nodes to initiate interactions with naïve T cells and activate CTLs responses. These intrinsic properties enable cDCs to restore immune surveillance and overcome immunological tolerance within the TME [161, 162]. However, CRC cells can severely impair DC

differentiation, maturation, and function through various immunosuppressive mechanisms, including the production of TGF- β , thus leading to immune tolerance [145]. Within a TGF- β -enriched TME, DCs adopt a tolerogenic state characterized by impaired antigen presentation and diminished capacity to activate T cells [163]. Consistent with this immunosuppressive reprogramming, DCs in CRC patients have been reported to express CD85k, an inhibitory receptor associated with immune suppression and tolerance, indicating direct tumor-mediated modulation of the host immune landscape [164]. Collectively, these alterations underscore that reversing the tolerogenic programming of cDCs represents a critical strategy for restoring effective antitumor immune responses. Accordingly, cDC-based immunotherapeutic approaches in CRC hold considerable promise for enhancing vaccine-induced immunity, overcoming tumor immune evasion, and addressing tumor heterogeneity, while maintaining a favorable safety profile [165].

DC dysfunction in CRC arises from impaired precursor differentiation that reduces the overall DC population, phenotypic shifts toward tolerogenic profiles that promote immune suppression, and impaired maturation that diminishes their antigen-presenting and costimulatory capacities. As a consequence, dysfunctional DCs fail to effectively recognize antigens or deliver adequate activation signals required for robust T-cell priming [41]. Orsini et al. demonstrated that DCs from CRC patients exhibit reduced antigen-presenting capacity and lower expression of costimulatory molecules, accompanied by elevated immunosuppressive cytokine IL-10 and reduced IL-12/TNF- α production, cytokines essential for Th1 polarization and effective anti-tumor immunity [166]. Additionally, tumor-derived factor, such as Ese-3 downregulation in cancer cells, further impairs DC maturation and consequently facilitates tumor proliferation [167, 168]. Nagorsen et al. reported paradoxical findings in which S100+ DC infiltration predicts improved survival yet coincides with greater Treg accumulation, thereby supporting local immunosuppression and tumor growth [169]. Furthermore, Hsu et al. showed that CRC-associated DCs express high levels of CXCL1, a chemokine that promotes tumor cell migration, drives EMT, and enhances stemness features [170]. Together, these observations reveal that CRC-induced reprogramming transforms potent immunostimulatory DCs into dysfunctional, tolerogenic, and pro-invasive phenotypes resembling immature DCs. This phenotypic shift represents a major barrier to effective anti-tumor immunity and contributes significantly to immune evasion in CRC [164]. Consequently, DCs emerge as indispensable

determinants of cancer vaccine immunogenicity and clinical efficacy.

Importantly, the extent and functional consequences of DC impairment are further shaped by CRC molecular subtypes. The distribution of pDCs population does not differ markedly between MSI-H/dMMR and MSS/pMMR CRC [171]. However, Ho et al. shows that the enrichment score for activated and immature DCs, particularly in the context of pMMR/MSS CRC liver metastases, is lower than MSI-H/dMMR primary CRCs, indicating a profound deficiency in antigen presentation capacity and early immune priming within these tumors [172]. These findings suggest that effective vaccine strategies for pMMR/MSS CRC cannot rely solely on antigen delivery but must incorporate approaches that actively restore DC number and function, as explained previously. Rational vaccine design should therefore relieve immunosuppressive signaling within the TME and emphasize DC-targeted delivery systems, incorporation of potent innate immune adjuvants (e.g., Toll-like receptor (TLR) agonists), and combination strategies that promote DC expansion and maturation (such as granulocyte-macrophage colony-stimulating factor (GM-CSF)).

In order to create durable antitumor responses, therapeutic use of cDCs in CRC immunotherapy involves diverse strategies, such as ex vivo manipulation, DC-based vaccine platforms, and TME modulation aimed at improving DC activity. In ex vivo approaches, patient-derived cDCs are isolated, activated under controlled conditions, and loaded with tumor antigens before reinfusion, enabling personalized antigen presentation and robust T-cell priming while minimizing off-target toxicity. Controlled conditions during the activation of DCs maximizes their antigen-presenting capacity and ability to induce robust immune responses [173–176]. Also, loading cDCs with a diverse array of tumor antigens further helps address intratumoral heterogeneity and broadens immune recognition across distinct tumor cell subsets [177, 178]. Moreover, successful DC-based vaccines can promote durable immune protection through the induction of memory T cells while exhibiting a favorable safety profile compared with conventional anticancer therapies [165].

However, the efficacy of DC-based interventions is frequently constrained by the profoundly immunosuppressive CRC TME, which collectively impair DC maturation and T-cell activation [179]. Consequently, combination strategies aimed at reprogramming the TME have emerged as critical adjuncts to DC-based therapies. ICIs can alleviate T-cell exhaustion, enhance intratumoral T-cell infiltration, and synergize with DC-mediated

antigen presentations [180, 181]. In parallel, cytokine-based interventions such as GM-CSF promote DC maturation through activation of JAK/STAT5, MAPK, and NF- κ B signaling pathways, leading to upregulation of MHC molecules, costimulatory ligands (CD80/CD86), and pro-inflammatory cytokines that collectively boost antigen presentation and amplify T-cell priming [182–184]. Additional approaches targeting suppressive immune populations, such as Tregs and TAMs, within the TME further improve DC efficacy by promoting anti-tumor immune responses [185, 186]. In preclinical models, the combination of cryoablation or radiofrequency ablation with immune checkpoint blockade and DC loading has been shown to restore immune equilibrium within the TME, confer protection against tumor recurrence, and enhance tumor-specific T-cell responses. The immunological effects of these approaches arise from the localized physical destruction of tumor tissue, via cold- or heat-induced ablation, which facilitates antigen release, precise tumor targeting, and subsequent activation of systemic antitumor immunity [187]. Collectively, these integrated strategies highlight the central role of cDCs as both antigen-presenting cells and immune orchestrators, underscoring the importance of combining DC-based approaches with TME-modulating interventions to maximize the therapeutic potential of CRC vaccines and immunotherapies.

2.4 Immune Cells: Neutrophils

Neutrophils, another major subset of the myeloid lineage, constitute the first line of defense against infection and acute inflammation. They exert antimicrobial activity through the release of lytic enzymes, phagocytosis, the formation of neutrophil extracellular traps (NETs), and the secretion of cytokines that sustain inflammatory responses and recruit additional immune cells [188, 189]. Within the context of cancer, tumor-associated neutrophils (TANs) have emerged as critical regulators of tumor biology. In CRC, TANs are strongly associated with patient prognosis and display both anti-tumor and pro-tumor activities depending on their phenotypic state and microenvironmental cues. Importantly, accumulating evidence indicates that neutrophils within the CRC TME can undergo phenotypic polarization toward pro-tumorigenic functions, thereby contributing to disease progression and immune evasion [189].

Triner et al. demonstrated that neutrophils suppress CRC progression and metastasis in mouse models by controlling tumor-associated microbiota. Conversely, neutrophil depletion disrupts intestinal flora, increases IL-17-producing bacteria, and accelerates CRC growth [190]. Other studies have shown that elevated neutrophil

infiltration accelerates CRC progression via CXCL1/CXCR2 signaling, increased immunosuppressive gene expression, and matrix metalloproteinase-9 (MMP9)-mediated T-cell suppression via TGF- β activation [191, 192]. As mentioned before, TGF- β also suppresses IFN- γ production, a key cytokine required for macrophage activation and for sustaining the effector functions of NK cells and neutrophils, thereby reinforcing a broadly immunosuppressive tumor milieu [193, 194]. Consistent with these findings, Fridlender et al. reported that TGF- β within the CRC TME can polarize TANs from an anti-tumor “N1” phenotype to a tumor-promoting “N2” phenotype. N2 TANs enhance tumor growth by fostering angiogenesis and conferring resistance to anti-VEGF therapy, while simultaneously promoting a more invasive tumor phenotype [195]. Beyond their role in immune evasion, TANs further promote metastasis by degrading basement membrane components, thereby enabling tumor cell migration and extravasation. Collectively, these observations underscore the dual and context-dependent roles of TANs in CRC and highlight their substantial contribution to tumor progression, immune suppression, and metastatic spread [196].

dMMR/MSI-H tumors are characterized by elevated TMB and dense infiltration of CTLs, which limits the dominance of immunosuppressive neutrophil populations and contributes to improved responsiveness to ICIs. In contrast, pMMR/MSS tumors generally exhibit an effector T-cell infiltration and enrichment of immunosuppressive myeloid populations, including pro-tumorigenic N2-like TANs [99]. These subtype-specific immune landscapes underscore why TAN-driven immune suppression is particularly relevant in MSS CRC and highlight the need for vaccine and combination strategies that actively modulate neutrophil recruitment and polarization to remodel the TME and enable effective antitumor immunity in immunologically cold CRC.

Collectively, these findings highlight the functional plasticity of TANs and their substantial impact on CRC progression and immune suppression, specifically into pro-tumor N2 phenotypes under the influence of TGF- β and other tumor-derived signals. Anti-TGF- β exerts its suppressive effects on CRC through dual and complementary mechanisms, involving inhibition of PI3K/AKT signaling in TANs and blockade of TGF- β /Smad signaling within tumor cells. Consistent with these effects, blockade of TGF- β signaling has been shown to repolarize TANs toward an anti-tumor N1 phenotype, thereby enhancing their cytotoxic activity against CRC cells while reducing the production of metastatic chemoattractants. Blockade of TGF- β signaling also enhances the expression of GM-CSF and IFN- γ , key

cytokines that regulate neutrophil differentiation within the TME [197]. These findings provide a strong mechanistic rationale for combination strategies aimed at alleviating TAN-mediated immune suppression, promoting T-cell infiltration, and ultimately improving overall therapeutic efficacy.

2.5 Cancer-Associated Fibroblasts

CAFs, the most abundant stromal component of the TME, play central roles in ECM maintenance and remodeling, desmoplasia, angiogenesis, tumor proliferation, immunosuppression, invasion, metastasis, and therapy resistance [198–200]. Within the TME, fibroblasts commonly acquire a chronically activated phenotype and differentiate into CAFs in response to tumor-derived growth factors, including TGF- β , epidermal growth factor, and bone morphogenetic proteins. TGF- β plays a crucial role in the transformation of fibroblasts into CAFs. CAFs are characterized by an elongated, spindle-like morphology, lack canonical markers of epithelial cells, endothelial cells, and leukocytes, and, importantly, do not harbor the oncogenic mutations observed in malignant cells [201]. Through ECM organization, CAFs regulate cancer cell behavior and increase tissue stiffness, thereby driving EMT and metastatic dissemination [202, 203]. Activin A secreted by CAFs has been shown to increase ECM stiffness and promote CRC progression [204].

CAFs also promote tumor growth by secreting a broad spectrum of chemokines and cytokines that engage in reciprocal crosstalk with cancer cells [200, 205]. Although the majority of CAFs arise from resident stromal fibroblasts, a significant subset originates from mesenchymal stem cells (MSCs) and mesothelial cells [198, 206]. MSCs, which possess pluripotent capacity, can be recruited to tumor sites, where tumor-derived signals drive their differentiation toward a CAF-like phenotype [207, 208]. Direct CAF–tumor cell interactions enhance tumor progression, invasiveness, and metastasis potential [209–211]. Mesothelial cells can similarly undergo mesenchymal transition, especially in highly invasive tumors, further contributing to CAF heterogeneity [212].

CAFs also mediate immune evasion [198, 213–215]. Zhang et al. reported that CXCL8 secreted by CAFs recruit monocytes into the CRC TME and induces M2 polarization through ICAM-1/VCAM-1 signaling or IL-8/CXCR2 pathways. The resulting M2-polarized macrophages synergize with CAFs to suppress immunosurveillance, amplifying TME immunosuppression [216, 217]. Consistent with these observations, high VCAM1 expression in CRC correlates with more aggressive tumor behavior and poor clinical outcomes, partly due to its

ability to promote pseudopodia formation and enhance CRC cell transmigration across endothelial layers in vitro [218]. In addition to modulating immune escape, CAFs stimulate angiogenic processes by inducing endothelial cells to secrete VEGF. This increase in VEGF production is driven, at least in part, by CRC-mediated potentiation of IL-6 secretion by CAFs [219]. Collectively, these findings underscore the multifaceted pro-tumorigenic roles of CAFs in CRC biology.

Collectively, transforming CAFs into anti-tumor fibroblasts is essential for preserving ECM mechanical homeostasis. Current therapeutic strategies include therapeutic cancer vaccines targeting fibroblast activation protein (FAP), inhibition of TGF- β secretion, and targeting key signaling pathways [220].

2.6 Endothelial Cells

Tumor vascularization is essential for CRC invasion and metastasis. Tumor vasculature is primarily composed of endothelial cells, supported by pericytes, smooth muscle cells, and endothelial progenitor cells [221]. Endothelial cells, typically quiescent in healthy tissues, undergo rapid proliferation and structural remodeling in response to tumor-derived pro-angiogenic signals, culminating in the formation of new blood vessels (angiogenesis) [222]. Angiogenesis is driven by paracrine cues, including VEGF, platelet-derived growth factor (PDGF), CXCL8, and TNF- α , secreted by various components of the TME. These signals drive the formation of aberrant vasculature that sustains tumor expansion by supplying oxygen and nutrients, removing metabolic waste, and facilitating metastatic dissemination [223–225].

Tumor-associated endothelial cells (TECs) further potentiate CRC progression by upregulating multiple growth factor receptors, including VEGFR and EGFR, thereby amplifying angiogenic signaling [226]. Beyond their angiogenic role, TECs contribute to tumor aggressiveness through additional mechanisms, such as secreting soluble Jagged-1, which activates Notch signaling and promotes stem-like phenotypes in CRC cells [227]. TECs also express adhesion molecules such as E-selectin, which facilitates tumor cell adhesion, invasion, and metastasis, while simultaneously recruiting neutrophils that reinforce an immunosuppressive TME [228].

Analogous to CAFs, TECs also modulate immune responses within the TME. For example, TEC-expressed FasL can induce apoptosis in CTLs, thereby promoting immune evasion [229]. Transcriptomic analyses further reveal that TEC expression of SPARC, COL1A1, COL1A2, and IGFBP3 is strongly associated with immunoinhibitory cell infiltration and correlates with T-cell exhaustion

[230]. Nevertheless, a subset of TECs may exert antitumor effects by enhancing CTL infiltration, maintaining vascular normalization, fostering a Th1-dominant immune milieu, and attenuating tumor growth, migration, and neovascularization [231, 232].

Anti-angiogenic therapies, particularly those targeting VEGF/VEGFR signaling, can transiently normalize tumor vasculature, improve T-cell access to the tumor bed, and synergize with ICIs [233]. Together, these observations provide a strong rationale for rational combination strategies to overcome immune exclusion, optimize antigen delivery, and improve the magnitude and durability of antitumor immune responses in CRC.

2.7 Extracellular Matrix

Beyond serving as a structural scaffold, the ECM, composed of diverse cellular constituents and structural proteins, is a dynamic regulator of cell–cell interactions, paracrine signaling, proliferation, immune evasion, and metastasis, thereby representing as a critical component of the TME [234]. Accumulation and remodeling of ECM proteins increase matrix stiffness, a hallmark of EMT and a key driver of CRC progression and metastasis. Substantial evidence indicates that ECM components, including collagen and fibronectin, play essential roles in these processes [204, 235–237]. Collagen promotes stem-like traits and metastatic potential through activation of the integrin/PI3K/AKT/Snail signaling axis; notably, elevated COL5A2 expression has been linked to poorer clinical outcomes [237, 238]. Li et al. similarly demonstrated a positive correlation between type I collagen density and tumor stage [234]. Similarly, fibronectin contributes to CRC aggressiveness and portends unfavorable prognosis [239, 240].

Enhanced ECM stiffness creates a substantial physical barrier that restricts immune cell infiltration and limits the effective intratumoral delivery of immunotherapeutic agents. In addition, ECM stiffening can further promote immune escape by modulating the expression of immune checkpoint molecules, thereby diminishing the therapeutic benefit of immune checkpoint blockade. For example, increased collagen fiber density and the resulting matrix stiffening can upregulate PD-L1 expression, thereby facilitating tumor immune evasion [220]. ECM stiffness is also a critical determinant of tumor responsiveness to chemotherapy. Increased matrix rigidity has been shown to upregulate the activity of multidrug resistance protein 1 at the cell membrane, thereby enhancing drug efflux and promoting chemoresistance [241]. In addition, elevated tumor stiffness impairs intratumoral drug penetration, further reducing therapeutic efficacy [242].

Despite these pro-tumorigenic functions, certain ECM constituents exhibit antitumor properties. Type IV collagen, for instance, can inhibit cancer cell invasion, and the positioning of the ECM between the basement membrane and interstitial space functions as a physical barrier that restricts tumor proliferation, differentiation, and dissemination [234, 243, 244]. However, ECM remodeling, characterized by upregulated MMP-2 and MMP-9, disrupts this barrier by degrading type IV collagen, thereby facilitating tumor cell motility, invasion, lymphovascular infiltration, and progression to advanced stages. Such remodeling is a defining feature of the CRC TME and contributes significantly to poor clinical outcomes [245–249].

Therapeutic modulation of ECM architecture therefore represents a biologically plausible strategy to enhance vaccine delivery, remodel the TME, and improve the efficacy of therapeutic cancer vaccines. ECM-targeted vaccine approaches primarily aim to reduce matrix stiffness and density in order to facilitate T-cell infiltration and augment cytotoxic immune responses. Cancer vaccines encoding the *Spam1* gene express hyaluronidase, leading to ECM degradation, improved tumor perfusion, reduced hypoxia, and reprogramming of the immune milieu, ultimately enhancing antitumor activity and lowering recurrence rates [250]. Similarly, vaccines directed against the extra-domain A (EDA) of fibronectin (FN) have demonstrated efficacy in limiting metastatic dissemination [251], whereas those targeting the extra-domain B (EDB) of FN effectively reduce solid tumor burden [252]. In addition, early clinical studies of FAP-targeted cancer vaccines have reported favorable safety profiles and evidence of tumor engagement, accompanied by enhanced CTLs-mediated antitumor immunity [253]. Moreover, vaccines expressing immature laminin receptor proteins have been associated with durable CTLs-dependent protective immune responses [254]. Additionally, reducing ECM stiffness by targeting collagen crosslinking can enhance T cell migration, thereby improving the efficacy of ICIs [255]. Similarly, small-molecule inhibitors of MMP-2/9 have been shown to enhance antitumor immune responses, decrease tumor burden, and improve survival, while simultaneously downregulating PD-L1 expression and potentiating the efficacy of ICIs-based immunotherapies [256]. Collectively, these findings highlight ECM stiffness as a critical modulator of immunotherapy responsiveness and underscore the therapeutic value of ECM remodeling in optimizing immunotherapeutic efficacy.

Importantly, the immunosuppressive features of the CRC TME have direct implications for therapeutic vaccine design and rational combination strategies. The dominance of Tregs, M2-like macrophages, and so on limits effective antigen presentation and attenuates vaccine-induced T-cell priming, highlighting the need for vaccine platforms that incorporate potent adjuvants or DC-targeting strategies to enhance cross-presentation. Likewise, T-cell exhaustion driven by chronic antigen exposure and inhibitory signaling pathways (e.g., PD-1/PD-L1, CTLA-4) provides a strong mechanistic rationale for combining CRC vaccines with ICIs to restore effector function rather than relying on vaccination alone. In addition, stromal desmoplasia, aberrant vasculature, and metabolic constraints within the CRC TME impair T-cell trafficking and survival, supporting the integration of vaccines with agents that remodel the ECM, normalize tumor vasculature, or modulate tumor metabolism. Together, these observations underscore that effective CRC vaccine strategies must be explicitly designed to counteract dominant immunosuppressive mechanisms within the TME.

3. Immunological Basis of CRC Vaccines

Besides being one of the most prevalent forms of cancer and cancer-related deaths, CRC poses unique immunological challenges compared with many other solid tumors, such as melanoma, due to relatively low TMB and limited tumor antigens diversity for most of CRC cases, particularly MSS/pMMR, resulting in weak baseline immunogenicity and poor spontaneous T-cell priming [257]. Lower expression of tumor antigens hinders effective cross-priming and accelerates T cell dysfunction [258]. Moreover, CRC tumors are characterized by a highly immunosuppressive TME enriched in Tregs, M2-polarized macrophages, and pro-tumorigenic neutrophils, alongside dominant inhibitory signaling pathways such as PD-1/PD-L1 and TGF- β [259]. Collectively, these features promote immune exclusion, inappropriate antigen presentation, T-cell exhaustion, and resistance to immune-mediated tumor eradication, distinguishing CRC as a malignancy in which effective immunotherapy, and particularly vaccine-based approaches, must overcome multiple, layered barriers to achieve durable clinical benefit.

Because CRC represents a chronic pathological condition, effective and potentially curative antitumor immunity, unlike immune responses to acute infections, relies on the establishment of durable, long-lived T-cell immunity. Compared with short-lived effector T cells, memory T cells possess the capacity for long-term survival and functional activity across peripheral tissues and tumor

sites, thereby representing a central benchmark of effective antitumor immune protection [260]. Effective antitumor immunity is sustained by antigen-specific memory T cells with strong proliferative potential [261]. In this context, vaccination represents a highly theoretically attractive therapeutic strategy due to its capacity to elicit highly specific anti-tumor immune responses with minimal adverse effects. Beyond acute immune activation, therapeutic CRC vaccines aim to establish durable immunological memory through the generation of long-lived memory T-cell subsets, including central memory T cells (T_{CM}), effector memory T cells (T_{EM}), and tissue-resident memory T cells (T_{RM}) [262]. Mizukoshi et al. showed that peptide vaccines can induce tumor-specific CTLs that persist for years and exhibit characteristics of long-lived memory, including self-renewal and sustained immune function without repeated vaccination [261]. In experimental models, cancer vaccines have also been shown to augment memory T-cell responses, and their combination with other immunotherapeutic modalities, such as bispecific antibodies, further enhances the durability and protective capacity of antitumor immune memory [263]. Collectively, these memory populations enable sustained immune surveillance and rapid recall responses upon antigen re-encounter, which are critical for long-term tumor control and prevention of disease recurrence [264]. By inducing robust antigen-specific priming and memory differentiation, cancer vaccines can enhance TIL density, support persistent effector function, and facilitate durable cytotoxic elimination of antigen-expressing CRC cells [262]. Compared with conventional therapeutic modalities, vaccines exhibit superior tolerability and are less likely to cause dose-dependent toxicities [265].

3.1 CRC-Antigen Processing and Presentation

Tumor antigens are broadly categorized into two groups: tumor-associated antigens (TAAs) and tumor-specific antigens (TSAs), also referred to as neoantigens. TAAs are proteins markedly overexpressed on malignant cells relative to normal tissues, whereas TSAs arise exclusively from tumor cells and are absent in healthy tissues. Both antigen types, however, are capable of binding to human leukocyte antigen (HLA) molecules and being presented by MHC molecules to T cells, thereby initiating antitumor immune responses [178].

As illustrated in Figure 4, antigen processing begins with transcription of genomic loci (for TAAs) or mutated loci (for TSAs), followed by translation, protein degradation, and MHC loading, culminating in surface presentation of TAA/TSA peptides on cancer cells. Tumor recognition by

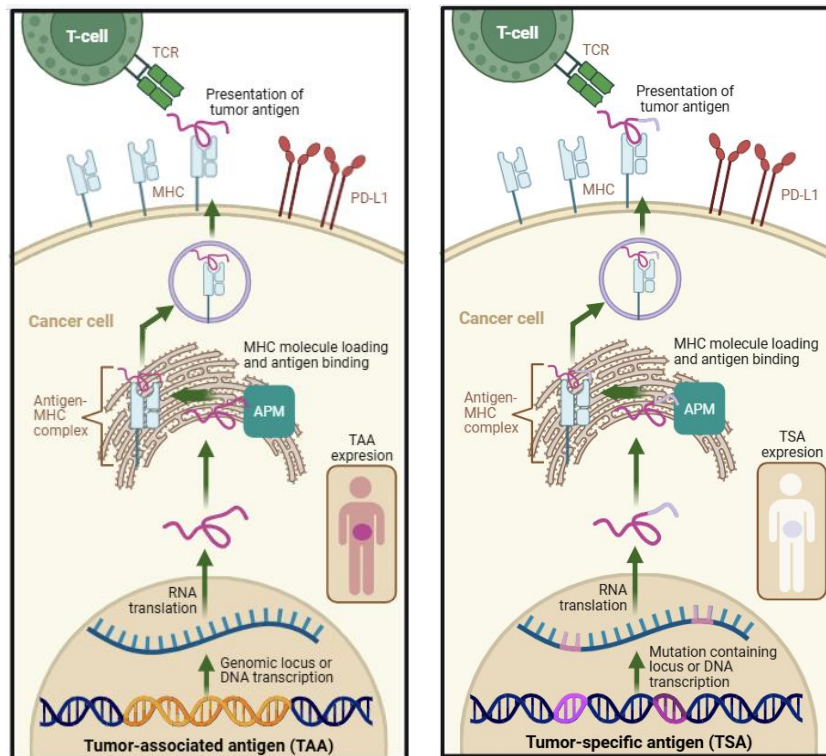


Figure 4. Key characteristics and processing pathways of tumor antigens. Antigen generation follows several steps: transcription of either the corresponding genomic locus (TAA; left) or the mutation-containing locus (TSA; right), followed by RNA translation, proteasomal degradation, loading of peptide fragments onto MHC molecules, and eventual presentation of the processed antigen-MHC complex on the tumor cell surface. TAA-derived proteins are highly expressed in tumors but remain detectable at low levels in normal tissues (pink patient; left), whereas the neoantigenic components of TSAs arise exclusively within tumor cells (cream patient; right). Tumor recognition by CTLs occurs through engagement of the TCR (green). Tumors may suppress this immune recognition by upregulating immune checkpoint molecules, including PD-L1 (red). APM, Antigen-presenting machinery; CTL, cytotoxic T lymphocyte; MHC, Major histocompatibility complex; TAA, Tumor-associated antigen; TCR, T-cell receptor; TSA, Tumor-specific antigen.

T cells occurs through the T-cell receptor (TCR). However, malignant cells frequently evade immune surveillance by upregulating immune checkpoint molecules such as PD-L1 [178]. Accordingly, immune checkpoint proteins have also emerged as auxiliary targets in vaccine-based immunotherapy, as their inhibition may enhance T-cell activation and mitigate immunosuppressive signaling [28]. Supporting this, Qi et al. demonstrated that checkpoint inhibition improves survival outcomes and yields early therapeutic signals therapeutic effects when combined with additional immunomodulatory strategies [30].

3.2 Mechanistic Basis of CRC Vaccine-Induced Immune Responses and Their Efficacy

The immune system comprises an intricate network of defense mechanisms that collectively protect the host from pathogens and foreign objects. It operates through two cooperative arms: innate and adaptive immunity [266]. Innate immunity responds rapidly, within hours, to pathogenic stimuli in a uniform and nonspecific manner, serving as the first line of defense. Conversely, adaptive immunity is highly specific and occurs when the innate

immune system fails to destroy a pathogen or foreign object. However, its specific target requires the adaptive immune system to recognize the pathogen or foreign object invading the body first, resulting in a slower response. Although slower to develop, it is more precise and capable of generating immunological memory, thereby enabling accelerated and amplified responses upon re-exposure [266, 267].

Adaptive immunity encompasses humoral and cellular responses. Humoral immunity, the antibody-mediated arm of adaptive immunity, operates primarily within the extracellular space. It is initiated when plasma B cells, with assistance from HTLs (HTL-dependent activation), produce antigen-specific antibodies following engagement with MHC molecules or macrophage-derived signaling. Cellular immunity, on the other hand, is mediated by antigen-specific T lymphocytes that develop in the bone marrow, mature in the thymus, and become activated upon recognition of peptide antigens presented by APCs through MHC molecules. CTLs eliminate infected or malignant cells via apoptosis, whereas HTLs enhance B-cell antibody production, augment cytotoxic cell activity, and activate

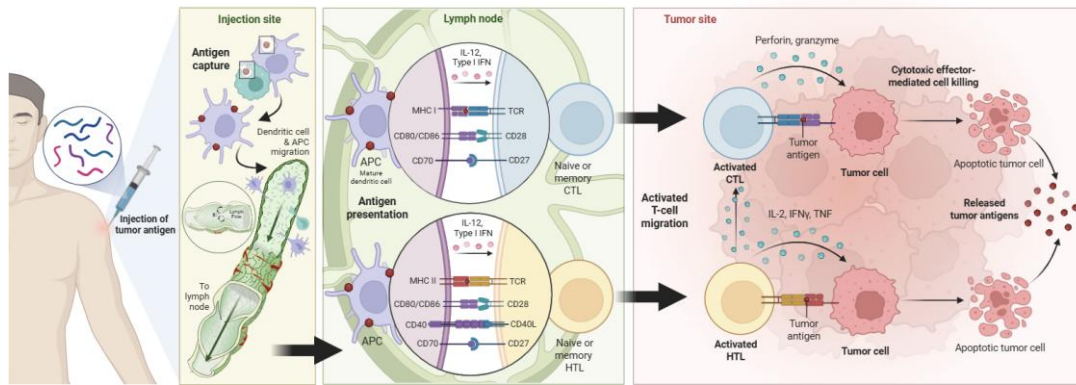


Figure 5. CRC vaccine immunological principle. APC, Antigen presenting cell; CTL, Cytotoxic T lymphocyte; HTL, Helper T lymphocyte; MHC, Major histocompatibility complex.

macrophages. Thus, HTLs function primarily as orchestrators of immune responses rather than direct cytotoxic effectors [268–270].

Like natural pathogens, vaccines are designed to initiate the immune activation cascade by engaging innate immunity. APCs, including DCs and macrophages, detect pathogen-associated molecular patterns (PAMPs) through pattern-recognition receptors (PRRs) such as TLRs [271–273]. Immature DCs possess high antigen-capturing capacity. Following antigen uptake, APCs upregulate MHC class I/II and costimulatory molecules in response to cytokines such as IL-12 and chemokine signaling (Figure 5). Antigen-loaded APCs then migrate to draining lymph nodes, where T-cell priming occurs [274].

Within the lymph nodes, captured antigens are processed into peptide epitopes, which are the essential immunogenic determinants recognized by T cells [271–273, 275, 276]. MHC molecules present only processed peptide fragments, not intact proteins [269, 277, 278]. The resulting epitopes are presented via MHC-I to activate CTLs or via MHC-II to activate Th1-type HTLs (Figure 5) [275, 279]. MHC-I-restricted presentation drives CTL proliferation and trafficking to the TME, where CTLs mediate cancer cell killing through perforin- and granzyme-dependent cytotoxicity and through cytokine release (e.g., IFN- γ , TNF- α) [271–273, 280, 281]. MHC-II presentation activates Th1-polarized HTLs, which further shape antitumor immunity by promoting CTL activation, supporting antibody production via B cells (including antibody-dependent cellular cytotoxicity, ADCC), and maintaining a pro-inflammatory milieu [271–273, 282].

Th1 cytokines generally potentiate antitumor immunity by activating both cellular and humoral immunity directed toward intracellular pathogens, whereas Th2-associated responses are primarily anti-inflammatory and less effective at controlling malignant cells. Th2 cells predominantly induce humoral immune responses

against extracellular pathogens [283, 284]. Within the tumor setting, Th1-polarized HTLs play a pivotal role in strengthening antitumor immunity by activating APCs through CD40–CD40L costimulatory interactions, enhancing MHC class I expression on tumor cells via IFN- γ secretion, and coordinating multiple immune effector populations to establish a robust immunostimulatory environment [280, 281].

Damaged or stressed tumor cells release additional tumor antigens and damage-associated molecular patterns (DAMPs). These signals enhance antigen cross-presentation by APCs, broaden the spectrum of T-cell responses (polyclonality), and further amplify vaccine-induced antitumor immunity [285]. Collectively, CRC vaccines function by shifting the immune landscape from tolerance toward active antitumor immunity, thereby enabling more efficient progression of the cancer-immunity cycle [286].

3.3 Memory T-Cell Formation in CRC Vaccination

As explained previously, APCs secrete immunostimulatory cytokines, which enhance the expression of costimulatory molecules and promote effective T-cell activation [287]. T-cell responses to acute antigenic stimulation typically progress through three sequential phases: an initial phase of rapid clonal expansion, followed by contraction of the effector population, and culminating in the establishment of long-lived immunological memory [288]. CRC-specific T cells are activated and clonally expand through coordinated engagement of the TCR with peptide–MHC complexes and requisite costimulatory signals, subsequently differentiating into effector and long-lived memory T-cell populations [289]. After the elimination of vaccine antigens, effector CTLs enter a rapid contraction phase in which most antigen-specific cells undergo apoptosis, while a small fraction (approximately 5–10%) persist and differentiate into long-lived memory cells. Importantly,

this transition is not stochastic, as memory potential is unequally distributed among effector cells rather than being uniformly inherited. These memory cells have a distinguish feature to rapidly acquire effector functions and generate a robust secondary expansion of CTLs capable of swiftly controlling recurrences [264, 288]. The maintenance and differentiation of memory T-cell subsets are regulated by a combination of extrinsic cues and intrinsic transcriptional programs. During priming, cytokines such as IL-2 drive T-cell proliferation and are critical for effective CTL memory recall responses, whereas IL-7 and IL-15 support the survival, differentiation, and long-term maintenance of memory T cells [290, 291].

Clinical studies in CRC have demonstrated that vaccine-induced adaptive responses can persist over extended periods and are associated with sustained antigen-specific T-cell activity, retained resistance to subsequent tumor rechallenge, and improved long-term outcomes. This durable antitumor protection was shown to be dependent on CTL memory cells, as depletion of this population abolished the protective immune response [292, 293]. For example, vector-based VRP-CEA vaccination was associated with long-term survival and durable T-cell responses in stage III colon cancer patients [294], and GUCY2C-PADRE vaccines induced CTL memory responses in early-stage CRC patients that provided durable protection against metastases [295].

CTL memory cells persist as a heterogeneous compartment comprising both circulating and non-circulating subsets. Circulating memory populations include T_{CM} , T_{EM} , and stem cell-like memory T cells (T_{SCM}), whereas non-circulating cells are classified as tissue-resident memory T cells (T_{RM}), which reside within tissues and tumors and are largely excluded from the blood. T_{CM} and T_{SCM} cells express lymphoid homing receptors such as CD62L and CCR7, enabling their trafficking to secondary lymphoid organs and the bone marrow, while T_{EM} cells lack these receptors and preferentially circulate through the blood and peripheral tissues [296–298]. In contrast, T_{RM} cells are defined by the absence of circulating markers and the expression of tissue retention molecules, including CD103 and CD69 [299].

The diversification of memory T-cell subsets likely optimizes protective immunity through functional specialization. Upon antigen re-exposure, T_{EM} and T_{RM} cells provide rapid effector functions and frontline defense at sites of antigen encounter, while T_{CM} mount robust recall responses by undergoing self-renewal and rapidly differentiating into effector CTL, T_{EM} , and T_{RM} populations [260, 288]. The presence of TCM and TEM cells has been correlated with improved clinical

outcomes in cancer patients. Consistent with this observation, ICIs immunotherapy has been shown in murine models of infection to enhance the generation and persistence of CTL memory cells, suggesting that similar mechanisms may contribute to the promotion of durable antitumor immune memory [300]. T_{SCM} similarly exhibit self-renewal capacity and highest degree of multipotency. Emerging evidence also indicates that locally reactivated T_{SCM} cells can transition into T_{CM} , T_{EM} , and effector CTL subsets within lymphoid tissues [260, 301]. In addition, locally reactivated T_{RM} cells can further contribute to immune recall by transitioning into T_{CM} and T_{EM} phenotypes, thereby reinforcing systemic and tissue-level antitumor immunity [260]. Because TRM are strategically positioned within peripheral tissues, they enable rapid local immune responses and represent attractive targets for vaccines aimed at enhancing protection at barrier sites and other organs [302].

3.4 Impact of the CRC TME on Vaccine-Induced Immunity and Rationale for CRC Vaccine-Based Combination Strategies

As detailed in Section 2, the CRC TME evolves in parallel with tumor progression and progressively dampens antitumor immune responses elicited during tumor development, thereby facilitating immune evasion [303]. This immunosuppressive milieu exerts profound suppressive effects on vaccine-induced immune responses. Immunosuppressive populations actively inhibit CTL infiltration, survival, and effector function [257, 304, 305]. In parallel, dominant inhibitory pathways such as PD-1/PD-L1 and TGF- β signaling promote T-cell exhaustion, exclusion, and functional impairment within the tumor bed. Consequently, although cancer vaccines can effectively prime antigen-specific T cells in peripheral lymphoid organs, these cells often fail to exert cytotoxic activity once they encounter the hostile CRC TME [135, 215]. This disparity between effective immune priming and intratumoral effector function represents a central obstacle to CRC vaccine efficacy and underscores the critical importance of elucidating CRC-specific immunosuppressive mechanisms for the rational design of effective immunotherapeutic strategies [303, 306].

Within this CRC-specific immunological framework, therapeutic CRC vaccines should be viewed as immune initiators rather than standalone effectors. In CRC, combination immunotherapy is designed to overcome the inherent limitations of single-agent immunotherapy and achieve improved therapeutic outcomes [257]. Accordingly, synergistic therapeutic benefits can be achieved through combination immunotherapy, in which complementary treatment modalities cooperatively enhance antitumor efficacy [307]. Vaccines provide the critical first step of expanding tumor-specific T-cell

repertoires and increasing tumor immunogenicity, while combination strategies, particularly ICIs, TGF- β blockade, and myeloid-modulating agents, are required to sustain and unleash these responses within the TME [308, 309]. ICIs or other immunomodulatory agents are administered in combination with cancer vaccines to potentiate vaccine-induced immune responses and enhance tumor cell eradication [310]. Such approaches modulate the TME by reprogramming immune-resistant T-cell populations, alleviating immunosuppressive signals, and promoting an inflamed, immunologically active milieu, thereby strengthening immune recognition and elimination of tumor cells [307, 311]. By enhancing antigen-specific priming while simultaneously relieving immune suppression and exclusion, rational vaccine-based combination approaches offer a biologically grounded strategy to convert immunologically “cold” CRC tumors into responsive, inflamed lesions capable of supporting durable antitumor immunity.

3.5 Determinants of Vaccine-Induced Memory T-Cell Quality in CRC Patients

The quality and durability of vaccine-induced immunological memory in CRC are influenced by host-related factors, particularly immunosenescence, which is prevalent in elderly CRC patients. Immunosenescence refers to the progressive decline in immune competence that occurs with advancing age, resulting in diminished responsiveness to new antigens and impaired vaccine efficacy in older adults. The immune system undergoes substantial age-related remodeling, resulting in immune responses that are strongly influenced by chronological age [312]. This age-related immune remodeling is characterized by a reduction in naïve T-cell output due to thymic involution [313], contraction of T-cell receptor diversity [314], and compromised generation of effective primary immune responses as illustrated by the poor vaccine response to hepatitis B [312, 315], while low-grade chronic inflammation and declines in regenerative capacity of hematopoietic lineages further exacerbate immune dysfunction [312, 316, 317]. Concurrent changes in hematopoietic stem cell (HSC) differentiation bias toward myeloid lineages at the expense of lymphoid precursor and declines in regenerative potential also contribute to decreased lymphoid cell production and repertoire diversity, which collectively undermine both innate and adaptive responses to vaccination [316].

Recent evidence suggests that age-related skewing of HSC differentiation toward the myeloid lineage is further intensified by telomeric stress [318]. Under conditions of telomerase insufficiency, the transcription factor basic leucine zipper transcription factor ATF-like (BATF) constrains the self-renewal capacity of lymphoid-biased

HSCs, promoting obligatory lymphoid differentiation coupled with reduced progenitor maintenance. This dual effect is proposed to drive loss of HSC pluripotency and preferential expansion of myeloid-committed HSCs, a mechanism particularly relevant in humans where telomere erosion accompanies aging [319, 320]. Consistently, human HSC aging is characterized by increased DNA damage and elevated BATF expression, ultimately resulting in an age-dependent decline in early and committed B-cell progenitors expressing CD45RA, CD38, CD10, and CD19 [321].

A central functional hallmark of T-cell aging is the reduced production of IL-2 following T-cell activation, which compromises clonal expansion and memory differentiation; notably, supplementation with exogenous IL-2 has been shown to partially restore age-associated defects in T-cell activation [322]. In addition, age-related T-cell dysfunction can be ameliorated by pro-inflammatory cytokines or TLR ligands, providing a strong mechanistic rationale for the use of potent adjuvants in vaccine design to enhance immunogenicity in aged hosts [323]. Understanding these mechanisms is therefore critical for optimizing therapeutic cancer vaccine strategies, particularly in elderly CRC patients who represent the majority of the affected population.

Beyond age-related reductions in T-cell numbers and repertoire diversity, intrinsic functional impairments also arise within memory T cells during aging. Many of these defects stem from altered cellular resource allocation, in which enhanced investment in metabolic maintenance compromises the capacity of T cells to mount effective activation and effector responses to antigenic stimulation. Clinically, this deterioration of T-cell memory is exemplified by the reactivation of latent varicella-zoster virus (VZV) as herpes zoster and by reduced responsiveness to vaccines such as influenza in elderly individuals [324]. Furthermore, aging is associated with a loss of polyfunctional antigen-specific T cells, with a shift toward monofunctional populations, thereby further compromising effective T-cell-mediated immunity [325].

4. Tumor Antigens in CRC Vaccine Development

Vaccine efficacy is fundamentally determined by the precision with which target antigens are selected. Consequently, the identification of tumor antigens represents a critical first step in vaccine development. To identify CRC-associated antigens, Corulli et al. evaluated two principal characteristics: (1) overexpression of candidate proteins in malignant versus normal tissues, and (2) biological relevance to CRC pathogenesis and progression, including their association with adverse prognosis [326]. Aberrantly high expression under

malignant conditions is necessary, as elevated antigen levels enhance immunogenicity [327]. Additional criteria proposed by Boris-Minev for prostate cancer, also relevant to CRC, include: (1) high tumor-specific expression, (2) absent or minimal expression in normal tissues, and (3) exposure of extracellular amino acid domains accessible to immunologic targeting [328].

4.1 Candidate Tumor Antigens for CRC Vaccines

4.1.1 Carcinoembryonic Antigen

One of the earliest TAAs to be identified is carcinoembryonic antigen (CEA), a 200-kDa glycoprotein that is physiologically expressed only during embryonic development but aberrantly re-expressed at high levels in CRC [178, 329–331]. In addition to its association with several malignancies, CEA is strongly linked to gastrointestinal cancers and is implicated in anti-apoptotic signaling, tumor invasion, and metastatic dissemination [329, 331–335]. Multiple studies have demonstrated that CEA-targeted vaccines are capable of eliciting antigen-specific humoral responses, activating HTLs, and inducing CTL-mediated cytotoxicity in human subjects [3, 336, 337]. Early in vitro studies also demonstrated that DCs pulsed with CEA-derived peptides are capable of priming CEA-specific CTL responses [338]. However, as a TAA, CEA lacks strict tumor specificity and is also expressed at low levels by normal epithelial tissues, resulting in immune tolerance. To overcome this limitation, several strategies have been developed to enhance the immunogenicity of CEA-based vaccines. One approach involves the use of altered peptide ligands with increased HLA-binding affinity, which has been shown to more effectively activate CEA-specific CTLs in vitro [339]. Alternatively, DNA vaccines encoding CEA-derived epitopes in combination with immunostimulatory cytokines, adjuvants, or helper T-cell epitopes have been designed to augment antigen-specific T-cell responses. In murine models, these multicomponent vaccine platforms induced stronger T-cell activation compared with peptide-only vaccines [340, 341].

4.1.2 Cell Surface-Anchored CEA-Related Cell Adhesion Molecule 6

Cell surface-anchored CEA-related cell adhesion molecule 6 (CEACAM6) is a glycosylphosphatidylinositol (GPI)-anchored surface glycoprotein that is markedly overexpressed in CRC and functions as both a tumor marker and an independent prognostic factor [342–345]. Its elevated expression in hyperplastic polyps and adenomas suggests a role in early tumorigenesis. CEACAM6 contributes to CRC initiation, progression, and metastasis, positioning it as a biologically relevant therapeutic target [343–348].

4.1.3 Mucin-1

Mucin-1 (MUC-1) is a transmembrane protein with a molecular weight of 120–225 kDa, which can increase to approximately 250–500 kDa upon glycosylation [349]. MUC-1 is normally expressed on epithelial cells, where it is polarized to the apical surface and characterized by extensive glycosylation [350]. In contrast, neoplasia-associated MUC1 is markedly overexpressed and exhibits pronounced hypoglycosylation relative to its normal counterpart. This aberrant expression is upregulated in CRC, as well as in precursor colorectal adenomas, and is associated with its immunogenic properties, involvement in TCR and BCR epitopes, mediation of metastasis and chemoresistance, and correlation with poor prognosis and advanced disease stage in CRC patients [349–353]. Notably, elevated MUC-1 expression has been shown to correlate with increasing degrees of dysplasia [352]. These effects are partly attributable to the pro-apoptotic activity of MUC-1 toward activated human T cells, as it enhances the efficient lysis of cytotoxic T cells, thereby contributing to immune evasion, and inhibits human T-cell proliferation as well as interactions between cytotoxic T cells and their target cells [354].

A study by Zhang et al. further demonstrated that MUC-1 can indirectly upregulate PD-L1 expression on tumor cells through the recruitment of pro-inflammatory cytokines, such as IL-17A and IFN- γ , which drive increased PD-L1 expression in tumor cells and tumor-infiltrating myeloid cells. This process subsequently suppresses antitumor immune responses via the PD-L1/PD-1 signaling pathway, enabling tumor cells to evade immune surveillance [355]. MUC-1 is also expressed in both major colorectal tumorigenesis pathways, namely the adenomatous polyp and the serrated polyp pathway [356].

A preclinical MUC1 vaccine in a colon cancer model overcame tolerance and induced IFN- γ -producing HTLs and CTLs, resulting in tumor rejection or reduced burden [357]. Another, preclinical studies show that CRC stem cell (CCSC) vaccine with MUC1 antigen significantly activated NK, T, and B cells, inhibited tumor growth, and required MUC1 expression for full antitumor efficacy in murine CRC models [358, 359]. In advanced CRC patients, MUC-1 vaccination induced robust anti-MUC-1 IgG responses and durable immunological memory, indicating a potential role in CRC prevention [360]. In a multicenter, randomized, double-blind, placebo-controlled trial of a MUC1 peptide vaccine for the prevention of recurrent colorectal adenoma, vaccination elicited measurable immune responses but did not significantly reduce overall adenoma recurrence compared to placebo. Among participants who developed a ≥ 2 -fold increase in anti-MUC1 IgG levels,

there was a trend toward lower adenoma recurrence and a 38 % absolute reduction in recurrence rates versus placebo, suggesting that immunogenic responders may derive clinical benefit. Importantly, the vaccine was immunogenic in a subset of recipients and was well tolerated without major safety concerns. These results indicate that while MUC1-targeted vaccination can induce antigen-specific immunity in high-risk individuals, boosting immunogenicity or combining with other immunomodulatory agents may be necessary to achieve meaningful preventive efficacy [361].

4.1.4 Epidermal Growth Factor Receptor

Epidermal growth factor receptor (EGFR) plays a central role in human tumorigenesis, and its dysregulation predominantly arises from receptor overexpression and mutations affecting distinct functional domains [362, 363]. Aberrant EGFR expression has been documented in a wide range of epithelial malignancies, including breast and triple-negative breast cancer, non-small cell lung cancer, head and neck cancers, CRC, and several other tumor types. Elevated EGFR expression in tumors is closely associated with increased tumor aggressiveness, unfavorable prognosis, reduced overall survival, diminished therapeutic responsiveness, and the development of treatment resistance [364]. Moreover, EGFR can form heterodimers with other members of the human epidermal growth factor receptor (HER) family, such as HER2, HER3, and HER4, leading to enhanced oncogenic signaling and the emergence of more aggressive cancer phenotypes with poorer clinical outcomes [365].

Foy et al. evaluated peptide vaccines targeting the EGFR as a tumor antigen platform. They developed immunogens corresponding to specific EGFR B-cell epitopes designed to elicit humoral and cellular anti-tumor immune responses. In preclinical models, vaccination with these EGFR peptide constructs significantly suppressed tumor growth and improved antitumor efficacy, demonstrating that antigen-specific immunization against EGFR can modulate tumor progression through enhanced immune recognition and effector activity. This work supports the feasibility of targeting EGFR not only with monoclonal antibodies but also through active immunization strategies that recruit both antibody-mediated and T-cell-dependent mechanisms of tumor control [364].

4.1.5 Survivin

Survivin, one of the most extensively studied members of the inhibitor of apoptosis protein (IAP) family, is overexpressed in malignant tumors and is strongly associated with a poor prognosis [366]. Beyond its

canonical anti-apoptotic function, survivin also promotes tumor cell invasion and migration through activation of NF- κ B-dependent signaling pathways, thereby contributing to metastatic dissemination [367]. Under normal physiological conditions, survivin expression is largely confined to proliferating fetal tissues and is absent from most differentiated adult cells [368]. In line with these findings, analysis of human transcriptomic datasets has identified survivin among a limited group of approximately 40 genes that are highly expressed in malignant tissues but show minimal or no expression in normal cells [369].

Sarela et al. have demonstrated marked upregulation of survivin across a wide range of human cancers, including CRC, where its expression is consistently associated with unfavorable clinical outcomes and poor prognosis [370]. Results from a meta-analysis encompassing 11 studies that conducted survival analyses demonstrated a significant association between survivin expression and poor prognosis in CRC patients. Furthermore, pooled analyses revealed that survivin expression was significantly associated with adverse pathological features, including lymph node metastasis and vascular invasion. Collectively, these findings indicate that survivin expression is linked to unfavorable clinical outcomes and a pro-metastatic phenotype in CRC, underscoring its potential utility as a candidate target for survivin-directed therapeutic strategies [371].

In the context of CRC immunotherapy, survivin has shown promise as a vaccine antigen. Clinical studies have demonstrated that CRC patients receiving DCs pulsed with survivin-derived peptides exhibited elevated frequencies of survivin-specific CTLs [372, 373]. In a subset of these patients, this immune activation was accompanied by reductions in tumor marker levels and, in some cases, measurable decreases in overall tumor burden [373].

4.1.6 Wilms' Tumor 1

Wild-type Wilms' Tumor 1 (WT1) protein is a TAA with broad expression across various malignancies, including colorectal adenocarcinomas, and play an important role in tumorigenesis of CRC, making it an attractive target for immunotherapy [374]. WT1 ranks at the top of a National Cancer Institute (NCI) prioritization list of cancer antigens based on criteria such as therapeutic relevance, immunogenicity, and potential for clinical benefit. WT1-targeted vaccines have been shown to induce specific immune responses in patients with both hematological and solid tumors, including measurable clinical activity [375]. In CRC tissues, WT1 expression is confirmed by immunohistochemistry alongside HLA class I and II

molecules, suggesting that WT1-derived peptides can be presented effectively to both CTLs and HTLs. A by Shimodaira et al. demonstrated that DC-based vaccine pulsed with WT1 is both safe, with only mild local and systemic reactions, and immunogenic in CRC patients and may contribute to durable antitumor immunity when integrated into multimodal treatment regimens [376].

4.1.7 Transmembrane 4 L Six Family Member 5

Transmembrane 4 L six family member 5 (TM4SF5), a tetraspanin-like cell surface glycoprotein with four transmembrane domains, is also overexpressed in CRC [377]. TM4SF5 promotes invasive behavior, enhances tumor cell motility, disrupts contact inhibition, and facilitates EMT [378–380]. Elevated TM4SF5 levels correlate with advanced disease and poor prognosis, highlighting its potential as a therapeutic target in CRC [3, 336, 337]. In a preclinical investigation of a peptide vaccine targeting TM4SF5 in a colon cancer model, Kwon et al. evaluated the therapeutic efficacy of a TM4SF5-specific peptide vaccine formulated with CpG-DNA and a liposome complex (Lipoplex) in tumor-bearing mice. Immunization with this vaccine elicited a robust production of TM4SF5-specific antibodies and was associated with significant suppression of tumor growth compared with control animals. The vaccinated mice also exhibited markedly reduced serum levels of VEGF, suggesting an anti-angiogenic effect contributing to tumor inhibition. These results demonstrate that a TM4SF5-targeting peptide vaccine can induce effective humoral immune responses and therapeutic antitumor activity in a murine model of colon cancer, supporting the feasibility of antigen-specific peptide vaccines in CRC immunotherapy research [381].

4.1.8 Protein Tyrosine Kinase 7

Protein tyrosine kinase 7 (PTK7) is a catalytically inactive transmembrane receptor that modulates key oncogenic pathways, including Wnt and VEGF signaling [382–393]. PTK7 is highly expressed in several malignancies, including CRC, and its expression correlates with poor differentiation, lymph node involvement, distant metastasis, and higher TNM stage [394–399]. PTK7 regulates tumor growth and dissemination and contributes to immune evasion through interactions with macrophage galactose-type lectin (MGL) [398, 400].

4.1.9 B7-H3

B7 Homolog 3 (B7-H3) is a transmembrane immune checkpoint molecule belonging to the B7 family of immunomodulatory proteins [401]. While minimally expressed in normal tissues, B7-H3 is markedly upregulated across multiple tumors. In CRC, elevated B7-

H3 levels are associated with tumor progression, metastatic spread, resistance to apoptosis, and unfavorable clinical outcomes [402–416]. B7-H3 functions as a T-cell co-inhibitory molecule, enhances tumor aggressiveness via JAK2–STAT3 activation, and reduces T-cell infiltration within the TME, collectively contributing to immune evasion [417–423].

4.1.10 Kirsten Rat Sarcoma Virus

Kirsten rat sarcoma virus (KRAS) is a key regulator of intracellular signaling pathways and is mutated in over 40% of CRC cases. During tumorigenesis, somatic mutations can arise throughout coding regions of the genome, giving rise to neoantigens. In CRC, mutations in KRAS represent a prototypical source of such neoantigens [424, 425]. Early studies demonstrated that peptides derived from mutant KRAS can effectively stimulate CTLs in vitro [426, 427], as well as in patients with CRC [428]. Subsequent investigations further confirmed the cytolytic capacity of CTLs induced by mutant KRAS peptides through the lysis of HLA-A2-positive target cells pulsed with mutant KRAS peptides [429].

Clinical vaccination trials using mutant KRAS-derived peptides have yielded evidence of potential therapeutic benefit. While objective immune responses were observed in a limited proportion of vaccinated CRC patients, a subset remained disease-free following vaccination [430]. In a separate case report, adoptive transfer of activated T cells targeting the KRAS G12D mutation induced regression of multiple lung metastases in a CRC patient, with durable tumor control lasting several months before progression of a single lesion [431].

To further enhance immunogenicity, mutant KRAS peptide vaccines have been combined with cytokine-based adjuvants such as IL-2 or GM-CSF. The most robust immune responses were observed in patients receiving GM-CSF, characterized by increased frequencies of interferon-producing, antigen-specific T cells. However, despite strong immunological activation, no objective clinical responses were achieved, and disease progression occurred in all patients. This lack of efficacy was accompanied by a concurrent expansion of Tregs, suggesting immune suppression as a likely mechanism underlying the discordance between immune activation and clinical outcome [432].

4.1.11 p53

Mutations affecting the tumor suppressor gene TP53, or genes regulating p53 activity, are frequently observed across multiple malignancies, including CRC, re

sulting in abnormal p53 expression [433]. These somatic mutations can generate neoantigenic peptides that are presented by MHC molecules and recognized by the immune system, providing a rationale for targeting mutant p53 in cancer immunotherapy. Early clinical evidence supporting this concept was provided by a phase I/II trial employing a recombinant canarypoxvirus (ALVAC) vaccine encoding wild-type human p53 in patients with advanced CRC, which demonstrated that p53-directed vaccination was feasible, safe, and capable of inducing p53-specific cellular immune responses. In this study, IFN- γ -secreting p53-specific T cells were detected, particularly at higher vaccine doses, indicating preferential induction of Th1-polarized immunity despite robust anti-vector responses [434].

In a phase I/II clinical trial, a synthetic long peptide (SLP) vaccine targeting the tumor-associated self-antigen p53 in patients with mCRC demonstrated that the was well tolerated, with toxicity limited to mild (grade 1–2) local reactions. Importantly, vaccine-induced p53-specific T-cell responses were detected in the majority patients, thereby supporting the feasibility, safety, and immunogenic potential of p53-directed vaccination in CRC patients [435]. In a phase I/II clinical trial evaluating a p53-targeted SLP vaccine in combination with IFN- α in metastatic colorectal cancer patients, the addition of IFN- α significantly enhanced p53-specific cellular immunity compared with historical results from p53-SLP vaccination alone. Marked increases in IFN- γ -producing, p53-specific T cells, indicate a stronger type-1 immune response than that achieved with p53 peptide vaccination alone. The combination regimen was also well tolerated and safe [436].

4.1.12 Melanoma-Associated Antigen

Melanoma-associated antigen (MAGE), originally identified in melanoma, belongs to the cancer-testis antigen family, a subgroup of TAAs characterized by restricted expression in immune-privileged testicular tissue and malignant cells. Subsequent studies have demonstrated MAGE expression across a broad range of adenocarcinomas, including CRC. However, reported frequencies of MAGE expression in CRC vary substantially depending on the study and specific MAGE isoform, with positivity rates of approximately 14% for MAGE-A, 51% for MAGE-A1–6, and 28% for MAGE-A3 [437–439].

Clinical evidence supporting the therapeutic potential of MAGE-targeted vaccination remains limited. Takahashi et al. reported that vaccination with a synthetic long peptide containing a helper/killer hybrid epitope derived from MAGE-A4 elicited coordinated HTL and CTL responses, resulting in modest tumor growth suppression and

disease stabilization in a single patient [440]. In contrast, a broader vaccination study evaluating multiple TAAs demonstrated enhanced antigen-specific CTL responses without translating into measurable clinical benefit [441].

Despite extensive efforts, the identification of optimal tumor antigens remains challenging, primarily due to the inherently low immunogenicity of many human cancers [442]. Consequently, additional immune stimulation is often required in designing vaccines based on unmodified tumor cells [443, 444]. Although single-peptide vaccines can induce robust immune activation, their impact on tumor regression is generally modest [178]. Current vaccine research therefore favors multi-epitope formulations composed of several multiple tumor-related peptides to achieve more durable and clinically meaningful outcomes.

5. Types of Vaccines and Advances in Clinical Trials and Therapeutic Outcomes

Figure 6 depicts various types of CRC vaccines, including peptide-based vaccine, DNA-based vaccine, RNA-/mRNA-based vaccine, cancer-cell based vaccine, dendritic cell vaccine, and virus- or vector-based vaccine. Each of which is discussed in detail in this section.

5.1 Molecular-Based Vaccines

Molecular-based CRC vaccines include peptide or full-length protein formulations as well as nucleic acid platforms such as RNA and DNA vaccines [286]. Peptide-based vaccines are engineered from immunogenic epitopes derived from tumor antigens, which are subsequently processed and presented through MHC class I and II pathways to activate T lymphocytes and establish durable antigen-specific immune memory against malignant cells [445]. These vaccines offer notable advantages, including high target specificity and relatively low manufacturing and storage costs [446, 447]. Despite their simplicity, affordability, and ease of production, no peptide-based cancer vaccine has yet achieved regulatory approval or commercialization. This is likely due to their inherently low immunogenicity of short peptide vaccine, HLA restriction that severely limits population coverage, limited capacity to induce sustained immune memory, and the requirement for booster dosing [28, 273]. Moreover, their clinical efficacy may be compromised by tumor-driven immune escape and antigen loss, both of which contribute to disease recurrence [448].

The use of minimal antigenic peptides necessitates further refinement, including precise epitope prioritization to ensure optimal targeting of CRC pathogenic pathways. Peptide vaccines also typically

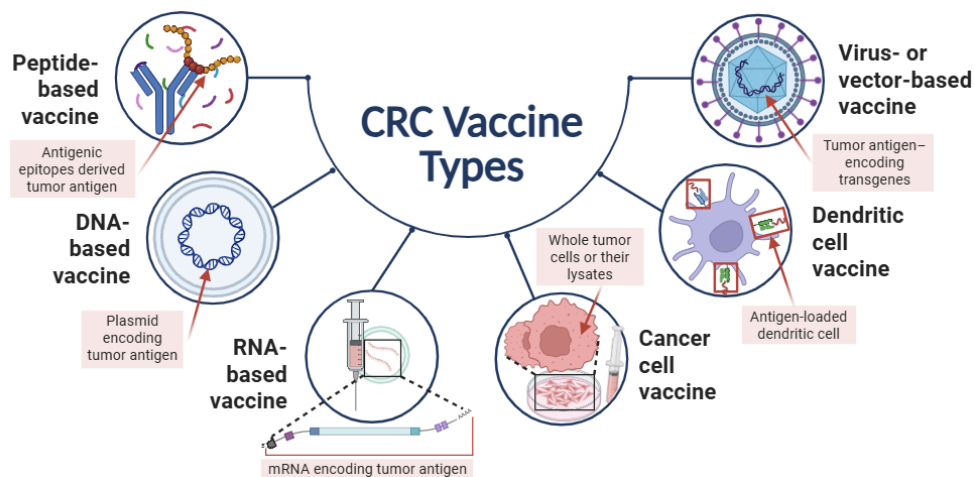


Figure 6. CRC vaccine types.

require potent adjuvants to augment immunogenicity [448]. Consequently, current clinical development efforts are increasingly focused on multi-antigen peptide vaccines formulated with strong adjuvants such as GM-CSF or Montanide, which enhance antigen presentation and T-cell priming [445, 446, 449–451]. Given the potential for antigen downregulation in CRC, combination immunotherapy, particularly with ICIs, has become an attractive strategy to counteract resistance mechanisms and amplify clinical benefit [27, 452, 453].

Although no peptide vaccines have reached global approval, multiple candidates have progressed into clinical development [273, 454]. A phase Ib trial evaluating PolyPEP11018, a multi-epitope vaccine incorporating 12 peptides from seven mCRC-associated tumor antigens, demonstrated safety, favorable tolerability, and robust HTL and CTL activation [455]. Another candidate, ATP128, currently being evaluated in MSS/pMMR stage IV CRC, has been shown to activate dendritic cells via TLR2 and TLR4 signaling, stimulate cytokine release, and upregulate costimulatory molecules essential for effective T-cell priming [456].

DNA vaccines are plasmid-based platforms that encode one or more tumor antigens. Following delivery into host cells, these plasmids undergo transcription and translation to generate antigenic proteins, which are subsequently processed and presented via MHC class I and II pathways while concurrently activating innate immune sensors through cytosolic receptors [457, 458]. This mechanism distinguishes DNA vaccines from peptide- or protein-based formulations, which rely on antigen presentation through phagocytosis, and from cellular vaccines, in which APCs directly display antigens upon administration [459].

One of the aberrantly expressed oncoproteins in CRC, MYB, has been investigated as a DNA vaccine target [460, 461]. A MYB-encoded DNA vaccine demonstrated robust prophylactic and therapeutic activity in a transgenic mouse model, reflecting the biological relevance of MYB, which is implicated in promoting cell proliferation, suppressing differentiation, and enhancing resistance to apoptosis [460–462]. Despite these supportive preclinical findings and the theoretical advantages of DNA vaccine platforms, several limitations hinder clinical translation [463]. The major challenges include the lack of optimized delivery systems, suboptimal immunogenicity, intratumoral heterogeneity, and restricted nuclear entry of plasmid DNA, coupled with potential risks of plasmid integration into the host genome. Current strategies to improve efficacy focus on rational antigen selection, plasmid optimization, and combinatorial regimens aimed at reducing TME-mediated immunosuppression and enhancing immune effector function [286, 463, 464].

mRNA-based vaccines have similarly gained prominence as preventive and therapeutic modalities. These vaccines are synthesized *in vitro* and encode tumor antigens in the form of mRNA. Unlike DNA vaccines, mRNA requires no nuclear entry and undergoes immediate cytoplasmic translation, conferring advantages in efficiency, modifiability, and production speed [465, 466]. Once internalized, the mRNA is translated into antigenic proteins that are presented by APCs through MHC class I, inducing strong CTL responses [467].

Moderna Therapeutics has advanced several mRNA vaccine candidates for oncology, including CRC. In phase I/II trials, mRNA-4650 delivered intramuscularly elicited both CTL and HTL responses against neoantigens without significant adverse effects or tumor recurrence [468]. Another candidate, mRNA-4157, demonstrated safety, tolerability, and preliminary clinical activity in MSI-H CRC

when combined with ICIs in a phase I study, leading to its progression into phase II trials [469]. Additionally, mRNA-5671, designed to target KRAS mutations, is currently being evaluated in phase I trials in combination with ICIs among non-MSI-H patients (NCT03948763). Preclinical evidence indicates that mRNA-5671 can substantially enhance CTL responses, both as monotherapy and in combination strategies [470].

Despite the substantial promise of nucleic acid-based cancer vaccines, DNA and mRNA modalities are constrained by several key challenges. These include: (1) the high cost and extended timelines required for good manufacturing practice (GMP)-compliant production, and (2) stability issues, particularly with mRNA, which necessitates ultra-low-temperature storage and complicates broad clinical distribution [471]. Addressing these limitations is essential for enabling large-scale implementation and optimizing their therapeutic potential.

5.2 Cancer Cell Vaccines

Cancer cell vaccines employ whole tumor cells or their lysates as antigen sources and are designed to prime broad immune responses [472]. These vaccines may be autologous, offering individualized specificity by using patient-derived tumor cells, or allogeneic, derived from standardized cell lines that facilitate scalable production [442, 473]. Although whole-cell antigen presentation can reduce the likelihood of tumor immune escape, this approach carries an inherent risk of autoimmunity due to shared antigen expression in healthy tissues [286].

Among recent developments, OncoVax, an autologous tumor cell vaccine combined with bacille Calmette-Guérin (BCG), has demonstrated acceptable safety when administered alongside 5-fluorouracil (5-FU) and leucovorin. However, early trials integrating OncoVax with surgical resection failed to yield significant improvements in clinical outcomes [474]. Conversely, the allogeneic vaccine GVAX, engineered to secrete GM-CSF, has shown evidence of modulating anti-tumor immune responses in a phase II study involving patients with advanced pMMR CRC [475].

5.3 Dendritic Cell Vaccines

DC vaccines leverage the exceptional antigen-presenting capacity of DCs isolated via leukapheresis. Following ex vivo maturation with cytokines and pulsing with tumor peptides or lysates, these cells are reinfused to stimulate potent anti-tumor immunity. DC vaccines, which have shown notable clinical efficacy in melanoma and prostate cancer, have also been explored extensively in CRC [476].

A randomized controlled trial by Rodriguez et al. demonstrated that CRC patients receiving postoperative DC vaccination following chemotherapy exhibited a markedly prolonged mean disease-free survival (25.26 months) relative to controls (9.53 months) [477]. MelCancerVac, a DC vaccine pulsed with allogeneic melanoma lysate (DDM-1.13), showed modest median overall survival (7.4 months) in a phase II study of stage IV CRC patients; however, a subset of patients exhibited exceptional outcomes, including progression-free intervals exceeding 6 months, with two individuals achieving disease control for more than 27 and 37 months, respectively [478].

5.4 Vector-Based Vaccines

Vector-based vaccines utilize viruses, attenuated bacteria, or yeast as delivery vehicles to introduce tumor antigen-encoding transgenes and elicit vigorous immune responses. These vectors intrinsically contain PAMPs recognized by host PRRs, thereby activating innate immunity and facilitating robust adaptive responses. Recombinant viral vectors are particularly attractive due to their ability to co-deliver immunogenic TAAs and innate immune activators, enhancing the magnitude and specificity of anti-tumor immunity [479, 480].

Despite decades of research, no viral vector-based CRC vaccine has advanced to phase III trials or received regulatory approval. This translational failure reflects a convergence of biological, immunological, and clinical trial-related limitations rather than a lack of intrinsic vaccine potency. Key contributing factors include suboptimal trial design, inadequate patient stratification, and enrollment of heavily pretreated patients with advanced disease, in whom profound tumor-induced immunosuppression limits vaccine efficacy. Moreover, conventional response criteria and short-term tumor shrinkage endpoints are poorly suited to capture the delayed and cumulative benefits of immunotherapy, leading to premature classification of clinical failure despite evidence of vaccine-induced immune activation [481].

Dosing strategies and repeated vector administration have further constrained efficacy by inducing anti-vector immunity, thereby reducing antigen expression and limiting effective boosting. Importantly, immunological analyses have revealed clear differences between responders and non-responders, with clinical benefit associated with sustained, polyfunctional antigen-specific T-cell responses and memory formation, whereas non-responders exhibit transient activation, T-cell exhaustion, or dominance of immunosuppressive cell populations. Collectively, these lessons explain why

robust preclinical immunogenicity has not consistently translated into meaningful clinical benefit and underscore the need for optimized trial design, refined immune monitoring, rational patient selection, and combination strategies to overcome tumor-mediated immune resistance [481].

In a phase II trial, Redman et al. evaluated a modified adenovirus subtype 5 vector, Ad5 (E1-, E2b-)-CEA (AdCEA), in patients with MSS mCRC. When combined with standard-of-care therapy, AdCEA produced a modest improvement in progression-free survival (10.1 vs. 8.8 months) and elicited significantly stronger HTL and CTL responses compared with standard therapy alone (8/11 patients vs. 1/8 patients) [482]. The overall limited clinical efficacy likely reflects suboptimal induction of durable anti-tumor immunity when used as monotherapy, underscoring the need for rational combinatorial strategies to unlock the full therapeutic potential of vector-based vaccination in CRC [479].

6. Potentials for the Therapeutic Development of Cancer Vaccines

Vaccines have demonstrated substantial success in preventing virus-associated malignancies, such as hepatitis B virus-related hepatocellular carcinoma and human papillomavirus (HPV)-driven cervical cancer, by targeting viral proteins that directly contribute to oncogenesis. This success has extended into therapeutic settings, where HPV vaccines administered in combination with ICIs have shown superior clinical responses compared with ICI monotherapy. These findings strengthen the rationale for applying similar antigen-targeted vaccination strategies to CRC, particularly through the development of multi-antigen vaccines incorporating well-defined, biologically relevant, and immunogenic tumor proteins [483].

Therapeutic cancer vaccines are increasingly recognized as a valuable component of cancer management, as they include durable, antigen-specific immune responses capable of controlling minimal residual disease and preventing tumor recurrence. Although immunogenicity may be attenuated in advanced disease, evidence from phase III clinical trials demonstrates that therapeutic vaccines can improve disease-free survival and overall survival in malignancies such as prostate cancer, melanoma, and follicular lymphoma [484–486]. As a modality designed to augment antitumor immunity, therapeutic vaccines promote tumor regression and elimination while maintaining a highly favorable safety profile. Their antigen-specific activity also reduces the likelihood of off-target immune activation. By selectively delivering TAAs or TSAs, vaccines stimulate robust

immune responses, enhance TIL accumulation, and activate cytotoxic mechanisms targeting antigen-expressing malignant cells [487]. When paired with immunostimulatory adjuvants, cancer vaccines can effectively potentiate T-cell-mediated immunity, a central determinant of tumor recognition and destruction [488–490].

In CRC, therapeutic cancer vaccines hold emerging promise, particularly as immunomodulatory agents capable of converting immunologically “cold” tumors into more inflamed and treatment-responsive lesions. Although CRC has historically shown limited responsiveness to vaccine monotherapy, multiple CRC-specific clinical and preclinical studies provide proof-of-concept that antigen-targeted vaccination can induce measurable antitumor immunity. Early-phase clinical trials using DC-based vaccines loaded with CRC-associated antigens such as survivin, WT1, CEA, and MUC-1 have consistently demonstrated safety and the induction of antigen-specific CTLs and HTLs responses in patients with advanced CRC or mCRC [373, 376, 491]. In some patients, vaccine-induced immune activation was accompanied by reductions in tumor markers or stabilization of disease [491].

Importantly, CRC-specific neoantigen vaccines targeting recurrent driver mutations such as KRAS and TP53 have provided further evidence of therapeutic potential. Peptide vaccines and adoptive transfer strategies targeting mutant KRAS have induced functional, mutation-specific T-cell responses in CRC patients, with isolated but striking clinical responses reported in metastatic disease [265, 431]. Similarly, p53-derived SLP vaccines have demonstrated favorable safety profiles and robust cellular immunogenicity in metastatic CRC patients, particularly when combined with immune adjuvants such as IFN- α , highlighting the feasibility of targeting shared tumor-associated antigens in CRC [434, 435].

Also, as explained in previous subsections, therapeutic vaccines alone may be insufficient in unselected CRC populations. They possess clear therapeutic potential when deployed in rational combination strategies, particularly alongside ICIs, chemotherapy, induced immunogenic cell death, or TME-modulating agents. Thus, CRC-directed cancer vaccines should be viewed not as standalone curative interventions, but as integral components of multimodal immunotherapy strategies tailored to tumor immunogenicity, molecular subtype, and host immune context.

Compared with classical drug-based therapies, therapeutic vaccines offer several key advantages:

(1) short treatment duration without daily dosing, (2) compatibility with ICIs to potentiate clinical outcomes, (3) the ability to induce durable immunological memory that prevents recurrence and can be boosted periodically, and (4) a highly favorable safety profile [265, 326]. Supporting this favorable safety profile, an immunoprevention study by Kimura et al. demonstrated that a single-antigen vaccine targeting the TAA MUC-1 elicited strong immunogenicity, inducing high titers of anti-MUC1 IgG and generating long-lasting immune memory, while remaining well tolerated with no evidence of autoimmunity [360]. Collectively, these findings indicate that while therapeutic cancer vaccines alone may be insufficient in unselected CRC populations, they represent a safe and biologically active platform that can be leveraged within multimodal treatment strategies to enhance antitumor immunity and improve long-term disease control.

7. Challenges in the Development of CRC Vaccines

As previously discussed, the development of therapeutic vaccines for CRC is hindered by the profoundly immunosuppressive TME and the sophisticated immune evasion strategies that characterize CRC immunopathology. Immunosuppression within the TME remains a major limiting factor affecting the clinical efficacy of vaccines in improving patient survival [492–495]. According to the cancer immunoediting paradigm, interactions between tumor cells and the immune system evolve through three stages: elimination, equilibrium, and escape. During the escape phase, tumors establish a highly suppressive microenvironment that is difficult to revert, thereby severely impairing vaccine-induced antitumor immunity [496]. Upregulation of immune checkpoint proteins such as PD-1 and PD-L1 further exacerbates immune escape, directly restraining the cytotoxic activity of vaccine-primed effector T cells [492–495]. Beyond these broadly recognized immunological barriers, CRC exhibits several unique biological, anatomical, and clinical features that impose additional and distinct challenges for the development of effective therapeutic vaccines.

As explained previously, MSS and pMMR tumors account for approximately 85% of CRC cases. These tumors are characterized by low TMB, limited neoantigen diversity, poor T-cell infiltration, and weak baseline interferon signaling, collectively resulting in an immunologically “cold” TME that is intrinsically resistant to both immune checkpoint inhibition and vaccine-induced immune priming [61, 497–499]. Consequently, vaccine strategies that rely on pre-existing antitumor immunity or robust neoantigen presentation face fundamental limitations in the majority of CRC patients. Metastatic patterns further

compound these challenges. The liver, the most common site of CRC metastasis, represents a uniquely tolerogenic immune organ enriched with immunosuppressive Kupffer cells, liver sinusoidal endothelial cells, Tregs, and myeloid-derived suppressor cells. This hepatic immune milieu actively promotes T-cell dysfunction, deletion, and exhaustion, thereby attenuating systemic and immunotherapy-induced antitumor responses [500–502]. As a result, immunotherapy that demonstrate immunogenicity in primary cancer lesions may fail to generate meaningful clinical benefit in patients with liver-dominant metastatic disease [503].

In addition, CRC is uniquely influenced by the gut microbiome, which plays a critical role in shaping mucosal immunity, antigen presentation, and systemic immune responsiveness. Dysbiosis, microbial-derived metabolites, and bacterial translocation can modulate DC function, T-cell polarization, and vaccine adjuvant efficacy. Emerging evidence indicates that specific microbial taxa can either enhance or suppress responses to cancer immunotherapies, including vaccines, highlighting the microbiome as both a confounding variable and a potential therapeutic target in CRC vaccine development [504–507]. This layer of complexity is particularly relevant to CRC and less pronounced in non-gastrointestinal malignancies. Beyond microbiome-mediated immunomodulation, anatomical and physiological features of the colorectum also impose constraints on immune surveillance and vaccine delivery. The intestinal immune system is inherently biased toward tolerance due to constant exposure to dietary antigens and commensal microbes. This tolerogenic baseline can dampen vaccine-induced immune activation and necessitates strategies capable of overcoming mucosal immune suppression without triggering excessive inflammation or autoimmunity [508, 509]. Furthermore, regional lymphatic drainage and local immune compartmentalization may limit effective trafficking of vaccine-primed effector cells to tumor sites.

Finally, CRC vaccine development must contend with intense competition from well-established standard-of-care therapies, including cytotoxic regimens such as FOLFOX and FOLFIRI, molecularly targeted agents (anti-EGFR, anti-VEGF), and selected immunotherapies for MSI-H/dMMR tumors. These treatments set a high benchmark for clinical efficacy, particularly in terms of progression-free and overall survival, thereby raising the evidentiary threshold for vaccine-based interventions to demonstrate additive or synergistic benefit [510, 511]. In this therapeutic landscape, cancer vaccines must not only be safe and immunogenic but also provide clear clinical

advantages over, or in rational combination with, existing modalities.

In addition to these CRC-specific biological and anatomical constraints, therapeutic vaccine development also faces challenges involving the balance between personalization and scalability. Neoantigen-based vaccines are inherently patient-specific, yet their production requires extensive processing from tumor tissue acquisition and genomic characterization to individualized vaccine formulation, resulting in substantial cost and time burdens. These limitations are particularly consequential in mCRC, where rapid disease progression demands timely therapeutic intervention [512]. Tumor heterogeneity presents an additional barrier, enabling subclones to evade immune detection and limiting the durability and consistency of vaccine responses across patients [178].

Moreover, determining the cause of suboptimal vaccine immunogenicity in cancer patients can be challenging. Reduced immune responses may result from inappropriate antigen selection (e.g., TAAs that largely resemble host molecules and are therefore subject to self-tolerance), suboptimal vaccine design (such as weak adjuvants), profound immunosuppression within the TME, the deleterious effects of prior therapy, advanced patient age, or a combination of these factors [360]. Despite considerable advances, identifying optimal tumor antigens for vaccine development remains difficult in most malignancies, including CRC, underscoring the need for refined antigen discovery approaches and rigorous antigen selection criteria [442].

Beyond biological barriers, financial constraints also significantly influence the development, accessibility, and commercial viability of cancer vaccines. The economic sustainability of these products depends on an appropriate balance between manufacturing costs, pricing strategies, and demonstrable clinical benefit. High-cost vaccines must provide substantial improvements in survival outcomes to justify premium pricing. For example, Sipuleucel-T, the first FDA-approved therapeutic cancer vaccine, continues to face adoption challenges due to its high-cost relative to its modest clinical benefit [513]. Its complex manufacturing workflow, involving leukapheresis, ex vivo antigen loading of autologous dendritic cells, and reinfusion, contributes to treatment costs exceeding \$90,000 per patient [514, 515]. By contrast, ICIs such as nivolumab and pembrolizumab remain widely used despite their high prices, largely due to their proven durable clinical responses and compatibility with combination therapy regimens [516, 517]. These market dynamics demonstrate that pricing strategies in oncology are

strongly shaped by perceived therapeutic benefit and competitive alternatives within the treatment landscape [518].

8. Utilization of Bioinformatics in Cancer Vaccine Development

Bioinformatics is an interdisciplinary field that integrates computer science, information technology, mathematics, and statistics to analyze, interpret, and model biological data, including predictions of gene regulatory networks [519, 520]. By enabling high-resolution interpretation of complex biological information, bioinformatics provides powerful computational frameworks for solving scientific challenges across diverse biomedical domains [521, 522]. In the healthcare sector, bioinformatics has become indispensable for overcoming the limitations of traditional drug discovery pipelines, which are often hindered by high development costs, prolonged timelines, and discrepancies between preclinical predictions and clinical outcomes. These capabilities are further strengthened by the rapid expansion of biological databases and the development of increasingly sophisticated, internationally validated computational platforms that support accurate prediction of pharmacological, toxicological, and molecular characteristics of therapeutic candidates [523].

A notable example of bioinformatics-driven therapeutic innovation is the development of imatinib mesylate (Gleevec), a targeted therapy for chronic myeloid leukemia designed to inhibit abnormal protein production resulting from the BCR-ABL fusion gene. Bioinformatics tools were instrumental in identifying and validating the relevant genetic drivers, enabling the rational design of this highly effective drug. Additionally, in time-sensitive global health emergencies, such as the COVID-19 pandemic, bioinformatics played a central role in rapidly characterizing the SARS-CoV-2 genome, predicting viral protein structures, identifying therapeutic targets, and accelerating vaccine development, thereby establishing foundational biological insights and underscoring its critical contribution to modern biomedical research [523].

Immunoinformatics, a specialized subfield of bioinformatics, integrates computational tools with immunological principles to analyze and model immune system behavior, predict disease mechanisms, and identify potential antigenic determinants [520, 524–526]. This approach supports the prediction of immunogenicity, allergenicity, and antigenicity of candidate antigens to ensure the generation of effective humoral and cellular immune responses [527–529]. Immunoinformatics significantly reduces the time and

resources required for vaccine target identification, often shortening the process to approximately 1–2 years compared with conventional vaccine development strategies [524, 530–532]. Immunoinformatics-guided vaccine research has demonstrated success across preclinical, clinical, and post-vaccination stages, highlighting its transformative impact on rational vaccine design and precision immunotherapy [519, 520, 526].

In the CRC context, these computational strategies directly address several key themes discussed in earlier sections of this review. For tumor antigens, bioinformatics enables systematic evaluation of gene and protein expression across CRC cohorts, *in silico* epitope mapping, and population-level HLA coverage analysis, thereby refining antigen selection and minimizing off-target toxicity [519, 533]. Moreover, bioinformatics provides the computational foundation for the neoantigen concept introduced throughout this manuscript, offering a rational solution to the antigen selection challenges highlighted in Section 7 [534–536]. Importantly, the personalized vaccine approaches discussed in Section 5, including mRNA-based platforms such as mRNA-4157 and mRNA-5671, are fundamentally dependent on bioinformatics pipelines for patient-specific mutation calling, epitope prioritization, and vaccine construct design, underscoring the integrative role of bioinformatics across CRC vaccine development [537, 538].

Neoantigen prediction pipelines have emerged as a critical application of bioinformatics in CRC vaccine development. Computational frameworks such as pVACtools, MuPeXI, and NeoPredPipe integrate whole-exome or whole-genome sequencing data with RNA sequencing to identify nonsynonymous somatic mutations, predict mutant peptide sequences, and rank candidate neoantigens based on their likelihood of being presented by patient-specific HLA molecules [534, 539–541]. These pipelines are particularly relevant for CRC, where MSI-H tumors exhibit elevated TMB and increased neoantigen load, making them more amenable to neoantigen-based vaccine strategies compared with MSS tumors [497, 499].

Accurate HLA typing and epitope–MHC binding prediction constitute another indispensable component of CRC vaccine bioinformatics. Tools such as NetMHCpan, MHCflurry, and MixMHCpred are widely used to predict peptide binding affinity and stability across diverse HLA class I and II alleles, enabling prioritization of epitopes capable of eliciting robust CTL and HTL responses [542–545]. These predictions are particularly important in CRC, given the extensive interpatient HLA diversity and the

need to design vaccines with sufficient population coverage or individualized specificity.

Beyond *in silico* prediction, bioinformatics-driven immunopeptidomics approaches have gained increasing importance in validating CRC neoantigens. Mass spectrometry-based profiling of HLA-bound peptides directly isolated from CRC tissues enables empirical confirmation of naturally presented tumor antigens, thereby refining computational predictions and reducing false-positive epitope selection [546, 547]. Integration of immunopeptidomics data with sequencing-based pipelines has been shown to improve the translational relevance of candidate vaccine epitopes and enhance the likelihood of clinical efficacy.

Importantly, bioinformatics-guided CRC vaccine development has already progressed into early-phase clinical evaluation. Several personalized neoantigen vaccines for CRC, often developed using integrated pipelines combining mutation calling, HLA binding prediction, and epitope prioritization, have demonstrated feasibility, safety, and immunogenicity in phase I trials, particularly in MSI-H CRC and in combination with immune checkpoint inhibitors [537, 538, 548]. These studies highlight how bioinformatics not only accelerates antigen discovery but also enables precision stratification of CRC patients most likely to benefit from vaccine-based immunotherapy.

Collectively, these advances underscore the indispensable role of bioinformatics and immunoinformatics in overcoming CRC-specific barriers to vaccine development. By enabling systematic neoantigen discovery, epitope validation, and patient stratification, computational approaches provide the conceptual and technical framework required for the next generation of effective CRC vaccines.

9. Conclusions

CRC represents an immunologically complex malignancy in which tumor progression, metastatic dissemination, and therapeutic resistance are strongly shaped by the TME. Accumulating evidence indicates that immunosuppressive cellular and stromal networks, including TAMs, Tregs, impaired DCs, CAFs, TANs, endothelial abnormalities, and ECM remodeling, collectively attenuate anti-tumor immunity and restrict the efficacy of current immunotherapeutic approaches. These features provide an important biological context for understanding the limited clinical efficacy observed with current vaccine-based strategies in CRC.

Therapeutic cancer vaccines are supported by a strong biological rationale, particularly their capacity to induce

antigen-specific T-cell responses and modulate tumor immunogenicity. This review has summarized key classes of CRC-associated antigens, including selected TAAs and neoantigen-based approaches, as well as major vaccine platforms evaluated to date. However, available clinical evidence remains largely confined to early-phase studies, with immunogenicity often exceeding demonstrated clinical benefit, underscoring the persistent translational gap in CRC vaccine development.

Synthesis of the current literature suggests that this gap reflects CRC-specific immunopathological constraints rather than insufficient antigen immunogenicity alone. The predominance of immunologically cold MSS/pMMR tumors, early immune exclusion, and tolerogenic pressures within metastatic niches, particularly the liver, distinguish CRC from more immunotherapy-responsive malignancies and limit the impact of vaccine monotherapy. Within this framework, vaccines appear to function primarily as immune primers, necessitating additional interventions to enable effective T-cell infiltration, persistence, and effector function within the TME.

Consistent with this concept, emerging data indicate that rational combination strategies, especially those integrating cancer vaccines with immune checkpoint inhibition, may partially restore T-cell functionality, although robust and durable clinical benefit has yet to be conclusively demonstrated. In parallel, computational and bioinformatics-based approaches have increasingly been applied to support antigen prioritization, neoantigen identification, and patient stratification, offering conceptual solutions to several challenges discussed in this review, while their clinical utility in CRC requires further prospective validation.

In summary, while meaningful progress has been made in elucidating CRC immunobiology and exploring vaccine-based immunotherapy, substantial challenges remain. Future advances will likely depend on improved and CRC-tailored antigen selection strategies, deeper integration of immunopathological insights into vaccine design, rational combination regimens that directly address dominant immunosuppressive mechanisms, and rigorous clinical evaluation incorporating robust immune monitoring. A mechanistically informed and critically assessed translational framework will be essential to define the ultimate clinical role of therapeutic cancer vaccines in CRC.

Author Contributions: Conceptualization, T.E.T. and S.B.V.S.; methodology, T.E.T.; software, S.B.V.S.; validation, T.E.T., S.B.V.S. and T.E.T.; formal analysis, S.B.V.S. and G.L.A.T.; investigation, S.B.V.S. and G.L.A.T.; resources, T.E.T.; data curation, S.B.V.S.; writing—original draft preparation, S.B.V.S. and G.L.A.T.;

writing—review and editing, S.B.V.S. and G.L.A.T.; visualization, T.E.T. and S.B.V.S.; supervision, T.E.T. and G.L.A.T.; project administration, T.E.T.; funding acquisition, T.E.T. and S.B.V.S. All authors have read and agreed to the published version of the manuscript.

Funding: This study does not receive external funding.

Data Availability Statement: Data that support the findings of this study are available from the corresponding author upon reasonable request.

Acknowledgments: All the authors would like to express their sincere gratitude to all individuals whose assistance, insights, and encouragement contributed to the completion of this study.

Conflicts of Interest: All the authors declare no conflicts of interest.

References

1. Wu, Z., Yang, M., and Cao, Y. (2022). Tumor Antigens and Vaccines in Colorectal Cancer, *Medicine in Drug Discovery*, Vol. 16, 100144. doi:10.1016/j.medidd.2022.100144.
2. Ferlay, J., Colombet, M., Soerjomataram, I., Mathers, C., Parkin, D. M., Piñeros, M., Znaor, A., and Bray, F. (2019). Estimating the Global Cancer Incidence and Mortality in 2018: GLOBOCAN Sources and Methods, *International Journal of Cancer*, Vol. 144, No. 8, 1941–1953. doi:10.1002/ijc.31937.
3. Xie, Y.-H., Chen, Y.-X., and Fang, J.-Y. (2020). Comprehensive Review of Targeted Therapy for Colorectal Cancer, *Signal Transduction and Targeted Therapy*, Vol. 5, No. 1, 22. doi:10.1038/s41392-020-0116-z.
4. Zhou, J., Yang, Q., Zhao, S., Sun, L., Li, R., Wang, J., Wang, L., and Wang, D. (2025). Evolving Landscape of Colorectal Cancer: Global and Regional Burden, Risk Factor Dynamics, and Future Scenarios (the Global Burden of Disease 1990–2050), *Ageing Research Reviews*, Vol. 104, 102666. doi:10.1016/j.arr.2025.102666.
5. Bray, F., Laversanne, M., Sung, H., Ferlay, J., Siegel, R. L., Soerjomataram, I., and Jemal, A. (2024). Global Cancer Statistics 2022: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries, *CA: A Cancer Journal for Clinicians*, Vol. 74, No. 3, 229–263. doi:10.3322/caac.21834.
6. Jeo, W., and Subrata, F. (2020). The Survival Rate of Colorectal Cancer in Dr. Cipto Mangunkusumo Hospital, *The New Ropanasuri: Journal of Surgery*, Vol. 5, No. 2, 13–17. doi:10.7454/nrjs.v5i2.1081.
7. Connell, L. C., Mota, J. M., Braghiroli, M. I., and Hoff, P. M. (2017). The Rising Incidence of Younger Patients with Colorectal Cancer: Questions about Screening, Biology, and Treatment, *Current Treatment Options in Oncology*, Vol. 18, No. 4, 23. doi:10.1007/s11864-017-0463-3.
8. Veruttipong, D. (2012). Age Distribution, Polyps and Rectal Cancer in the Egyptian Population-Based Cancer Registry, *World Journal of Gastroenterology*, Vol. 18, No. 30, 3997. doi:10.3748/wjg.v18.i30.3997.
9. Naganna, S. M., Vidyavathi, K., Kumar, H. M., and Bhaskaran, A. (2016). Histomorphological Characteristics of Colorectal Carcinoma in the Young and Elderly: Is There a Difference?, *Indian Journal of Pathology and Oncology*, Vol. 3, No. 2, 293. doi:10.5958/2394-6792.2016.00056.9.
10. Carethers, J. M. (2016). The Increasing Incidence of Colorectal Cancers Diagnosed in Subjects under Age 50 among Races:

- CRaCking the Conundrum, *Digestive Diseases and Sciences*, Vol. 61, No. 10, 2767–2769. doi:10.1007/s10620-016-4268-1.
11. Arima, K., Zhong, R., Ugai, T., Zhao, M., Haruki, K., Akimoto, N., Lau, M. C., Okadome, K., Mehta, R. S., Väyrynen, J. P., Kishikawa, J., Twombly, T. S., Shi, S., Fujiyoshi, K., Kosumi, K., Ogata, Y., Baba, H., Wang, F., Wu, K., Song, M., Zhang, X., Fuchs, C. S., Sears, C. L., Willett, W. C., Giovannucci, E. L., Meyerhardt, J. A., Garrett, W. S., Huttenhower, C., Chan, A. T., Nowak, J. A., Giannakis, M., and Ogino, S. (2022). Western-Style Diet, Pks Island-Carrying *Escherichia Coli*, and Colorectal Cancer: Analyses from Two Large Prospective Cohort Studies, *Gastroenterology*, Vol. 163, No. 4, 862–874. doi:10.1053/j.gastro.2022.06.054.
 12. Rothwell, J. A., Murphy, N., Bešević, J., Kliemann, N., Jenab, M., Ferrari, P., Achaintre, D., Gicquiau, A., Vozar, B., Scalbert, A., Huybrechts, I., Freisling, H., Prehn, C., Adamski, J., Cross, A. J., Pala, V. M., Boutron-Ruault, M.-C., Dahm, C. C., Overvad, K., Gram, I. T., Sandanger, T. M., Skeie, G., Jakszyn, P., Tsilidis, K. K., Aleksandrova, K., Schulze, M. B., Hughes, D. J., van Guelpen, B., Bodén, S., Sánchez, M.-J., Schmidt, J. A., Katzke, V., Kühn, T., Colorado-Yohar, S., Tumino, R., Bueno-de-Mesquita, B., Vineis, P., Masala, G., Panico, S., Eriksen, A. K., Tjønneland, A., Aune, D., Weiderpass, E., Severi, G., Chajès, V., and Gunter, M. J. (2022). Metabolic Signatures of Healthy Lifestyle Patterns and Colorectal Cancer Risk in a European Cohort, *Clinical Gastroenterology and Hepatology*, Vol. 20, No. 5, e1061–e1082. doi:10.1016/j.cgh.2020.11.045.
 13. Patel, S. G., Karltz, J. J., Yen, T., Lieu, C. H., and Boland, C. R. (2022). The Rising Tide of Early-Onset Colorectal Cancer: A Comprehensive Review of Epidemiology, Clinical Features, Biology, Risk Factors, Prevention, and Early Detection, *The Lancet Gastroenterology & Hepatology*, Vol. 7, No. 3, 262–274. doi:10.1016/S2468-1253(21)00426-X.
 14. Bartnik, A., Nirmal, A. J., and Yang, S.-Y. (2012). Peptide Vaccine Therapy in Colorectal Cancer, *Vaccines*, Vol. 1, No. 1, 1–16. doi:10.3390/vaccines1010001.
 15. Zhang, Y., and Zhang, Z. (2020). The History and Advances in Cancer Immunotherapy: Understanding the Characteristics of Tumor-Infiltrating Immune Cells and Their Therapeutic Implications, *Cellular & Molecular Immunology*, Vol. 17, No. 8, 807–821. doi:10.1038/s41423-020-0488-6.
 16. Haslam, A., and Prasad, V. (2019). Estimation of the Percentage of US Patients with Cancer Who Are Eligible for and Respond to Checkpoint Inhibitor Immunotherapy Drugs, *JAMA Network Open*, Vol. 2, No. 5, e192535. doi:10.1001/jamanetworkopen.2019.2535.
 17. Giraldo, N. A., and Taube, J. M. (2018). PD-L1 and Other Immunological Diagnosis Tools, *Oncoimmunology*, Springer International Publishing, Cham, 371–385. doi:10.1007/978-3-319-62431-0_23.
 18. Oladejo, M., Paulishak, W., and Wood, L. (2023). Synergistic Potential of Immune Checkpoint Inhibitors and Therapeutic Cancer Vaccines, *Seminars in Cancer Biology*, Vol. 88, 81–95. doi:10.1016/j.semcan.2022.12.003.
 19. Lin, M. J., Svensson-Arvelund, J., Lubitz, G. S., Marabelle, A., Melero, I., Brown, B. D., and Brody, J. D. (2022). Cancer Vaccines: The next Immunotherapy Frontier, *Nature Cancer*, Vol. 3, No. 8, 911–926. doi:10.1038/s43018-022-00418-6.
 20. Rosenberg, S. A. (2014). IL-2: The First Effective Immunotherapy for Human Cancer, *The Journal of Immunology*, Vol. 192, No. 12, 5451–5458. doi:10.4049/jimmunol.1490019.
 21. Riley, R. S., June, C. H., Langer, R., and Mitchell, M. J. (2019). Delivery Technologies for Cancer Immunotherapy, *Nature Reviews Drug Discovery*, Vol. 18, No. 3, 175–196. doi:10.1038/s41573-018-0006-z.
 22. Kciuk, M., Yahya, E. B., Mohamed Ibrahim Mohamed, M., Rashid, S., Iqbal, M. O., Kontek, R., Abdulsamad, M. A., and Allaq, A. A. (2023). Recent Advances in Molecular Mechanisms of Cancer Immunotherapy, *Cancers*, Vol. 15, No. 10, 2721. doi:10.3390/cancers15102721.
 23. Belli, C., Trapani, D., Viale, G., D'Amico, P., Duso, B. A., Della Vigna, P., Orsi, F., and Curigliano, G. (2018). Targeting the Microenvironment in Solid Tumors, *Cancer Treatment Reviews*, Vol. 65, 22–32. doi:10.1016/j.ctrv.2018.02.004.
 24. Taube, J. M., Galon, J., Sholl, L. M., Rodig, S. J., Cottrell, T. R., Giraldo, N. A., Baras, A. S., Patel, S. S., Anders, R. A., Rimm, D. L., and Cimino-Mathews, A. (2018). Implications of the Tumor Immune Microenvironment for Staging and Therapeutics, *Modern Pathology*, Vol. 31, No. 2, 214–234. doi:10.1038/modpathol.2017.156.
 25. Mlecnik, B., Bindea, G., Angell, H. K., Maby, P., Angelova, M., Tougeron, D., Church, S. E., Lafontaine, L., Fischer, M., Fredriksen, T., Sasso, M., Bilocq, A. M., Kirilovsky, A., Obenauf, A. C., Hamieh, M., Berger, A., Bruneval, P., Tuech, J.-J., Sabourin, J.-C., Le Pessot, F., Mauillon, J., Rafii, A., Laurent-Puig, P., Speicher, M. R., Trajanoski, Z., Michel, P., Sesboüe, R., Frebourg, T., Pagès, F., Valge-Archer, V., Latouche, J.-B., and Galon, J. (2016). Integrative Analyses of Colorectal Cancer Show Immunoscore Is a Stronger Predictor of Patient Survival than Microsatellite Instability, *Immunity*, Vol. 44, No. 3, 698–711. doi:10.1016/j.immuni.2016.02.025.
 26. Giraldo, N. A., Sanchez-Salas, R., Peske, J. D., Vano, Y., Becht, E., Petitprez, F., Validire, P., Ingels, A., Cathelineau, X., Fridman, W. H., and Sautès-Fridman, C. (2019). The Clinical Role of the TME in Solid Cancer, *British Journal of Cancer*, Vol. 120, No. 1, 45–53. doi:10.1038/s41416-018-0327-z.
 27. Obara, W., Kanehira, M., Katagiri, T., Kato, R., Kato, Y., and Takata, R. (2018). Present Status and Future Perspective of Peptide-based Vaccine Therapy for Urological Cancer, *Cancer Science*, Vol. 109, No. 3, 550–559. doi:10.1111/cas.13506.
 28. Runtunuwu, S. V. B. E., Tallei, T. E., and Turalaki, G. L. A. (2025). Prostate Cancer Vaccines: Progress, Challenges, and Future Directions, *Heca Journal of Applied Sciences*, Vol. 3, No. 1, 30–55. doi:10.60084/hjas.v3i1.263.
 29. Gallio, C., Esposito, L., and Passardi, A. (2025). Therapeutic Cancer Vaccines in Colorectal Cancer: Platforms, Mechanisms, and Combinations, *Cancers*, Vol. 17, No. 15, 2582. doi:10.3390/cancers17152582.
 30. Qi, G.-X., Zhao, R.-X., Gao, C., Ma, Z.-Y., Wang, S., and Xu, J. (2025). Recent Advances and Challenges in Colorectal Cancer: From Molecular Research to Treatment, *World Journal of Gastroenterology*, Vol. 31, No. 21. doi:10.3748/wjg.v31.i21.106964.
 31. Worthley, D. L., and Leggett, B. A. (2010). Colorectal Cancer: Molecular Features and Clinical Opportunities., *The Clinical Biochemist. Reviews*, Vol. 31, No. 2, 31–8.
 32. Jiang, C., Zhou, Q., Yi, K., Yuan, Y., and Xie, X. (2024). Colorectal Cancer Initiation: Understanding Early-Stage Disease for Intervention, *Cancer Letters*, Vol. 589, 216831. doi:10.1016/j.canlet.2024.216831.
 33. Milkva, I. (2016). Genetics of Colorectal Tumorigenesis (Possibilities of Testing and Screening Prediction of Hereditary Form of Colorectal Cancer – Lynch Syndrome), *Klinicka Onkologie*, Vol. 29, No. Suppl 1, S55–S61. doi:10.14735/amko2016555.
 34. Al-Sohaily, S., Biankin, A., Leong, R., Kohonen-Corish, M., and Warusavitarne, J. (2012). Molecular Pathways in Colorectal Cancer, *Journal of Gastroenterology and Hepatology*, Vol. 27, No. 9, 1423–1431. doi:10.1111/j.1440-1746.2012.07200.x.

35. Balchen, V., and Simon, K. (2016). Colorectal Cancer Development and Advances in Screening, *Clinical Interventions in Aging*, Vol. 11, 967–976. doi:10.2147/CIA.S109285.
36. Ogino, S., and Goel, A. (2008). Molecular Classification and Correlates in Colorectal Cancer, *The Journal of Molecular Diagnostics*, Vol. 10, No. 1, 13–27. doi:10.2353/jmoldx.2008.070082.
37. Jaspersion, K. W., Tuohy, T. M., Neklason, D. W., and Burt, R. W. (2010). Hereditary and Familial Colon Cancer, *Gastroenterology*, Vol. 138, No. 6, 2044–2058. doi:10.1053/j.gastro.2010.01.054.
38. Weisenberger, D. J., Siegmund, K. D., Campan, M., Young, J., Long, T. I., Faasse, M. A., Kang, G. H., Widschwendter, M., Weener, D., Buchanan, D., Koh, H., Simms, L., Barker, M., Leggett, B., Levine, J., Kim, M., French, A. J., Thibodeau, S. N., Jass, J., Haile, R., and Laird, P. W. (2006). CpG Island Methylator Phenotype Underlies Sporadic Microsatellite Instability and Is Tightly Associated with BRAF Mutation in Colorectal Cancer, *Nature Genetics*, Vol. 38, No. 7, 787–793. doi:10.1038/ng1834.
39. Keum, N., and Giovannucci, E. (2019). Global Burden of Colorectal Cancer: Emerging Trends, Risk Factors and Prevention Strategies, *Nature Reviews Gastroenterology & Hepatology*, Vol. 16, No. 12, 713–732. doi:10.1038/s41575-019-0189-8.
40. Chandra, R., Karalis, J. D., Liu, C., Murimwa, G. Z., Voth Park, J., Heid, C. A., Reznik, S. I., Huang, E., Minna, J. D., and Brekken, R. A. (2021). The Colorectal Cancer Tumor Microenvironment and Its Impact on Liver and Lung Metastasis, *Cancers*, Vol. 13, No. 24, 6206. doi:10.3390/cancers13246206.
41. Li, J., Chen, D., and Shen, M. (2022). Tumor Microenvironment Shapes Colorectal Cancer Progression, Metastasis, and Treatment Responses, *Frontiers in Medicine*, Vol. 9. doi:10.3389/fmed.2022.869010.
42. Whiteside, T. L. (2008). The Tumor Microenvironment and Its Role in Promoting Tumor Growth, *Oncogene*, Vol. 27, No. 45, 5904–5912. doi:10.1038/onc.2008.271.
43. Galon, J., Mlecnik, B., Bindea, G., Angell, H. K., Berger, A., Lagorce, C., Lugli, A., Zlobec, I., Hartmann, A., Bifulco, C., Nagtegaal, I. D., Palmqvist, R., Masucci, G. V., Botti, G., Tatangelo, F., Delrio, P., Maio, M., Laghi, L., Grizzi, F., Asslaber, M., D'Arrigo, C., Vidal-Vanaclocha, F., Zavadvova, E., Chouchane, L., Ohashi, P. S., Hafezi-Bakhtiari, S., Wouters, B. G., Roehrl, M., Nguyen, L., Kawakami, Y., Hazama, S., Okuno, K., Ogino, S., Gibbs, P., Waring, P., Sato, N., Torigoe, T., Itoh, K., Patel, P. S., Shukla, S. N., Wang, Y., Kopetz, S., Sinicrope, F. A., Scripcariu, V., Ascierto, P. A., Marincola, F. M., Fox, B. A., and Pagès, F. (2014). Towards the Introduction of the 'Immunoscore' in the Classification of Malignant Tumours, *The Journal of Pathology*, Vol. 232, No. 2, 199–209. doi:10.1002/path.4287.
44. Taube, J. M., Galon, J., Sholl, L. M., Rodig, S. J., Cottrell, T. R., Giraldo, N. A., Baras, A. S., Patel, S. S., Anders, R. A., Rimm, D. L., and Cimino-Mathews, A. (2018). Implications of the Tumor Immune Microenvironment for Staging and Therapeutics, *Modern Pathology*, Vol. 31, No. 2, 214–234. doi:10.1038/modpathol.2017.156.
45. Zhou, Z., Wang, J., Wang, J., Yang, S., Wang, R., Zhang, G., Li, Z., Shi, R., Wang, Z., and Lu, Q. (2024). Deciphering the Tumor Immune Microenvironment from a Multidimensional Omics Perspective: Insight into next-Generation CAR-T Cell Immunotherapy and Beyond, *Molecular Cancer*, Vol. 23, No. 1, 131. doi:10.1186/s12943-024-02047-2.
46. Kim, H.-D., Kim, J. H., Ryu, Y.-M., Kim, D., Lee, S., Shin, J., Hong, S.-M., Kim, K.-H., Jung, D., Song, G., Hwang, D. W., Lee, J. H., Song, K. B., Ryoo, B.-Y., Jeong, J. H., Kim, K., Kim, S.-Y., and Yoo, C. (2021). Spatial Distribution and Prognostic Implications of Tumor-Infiltrating FoxP3- CD4+ T Cells in Biliary Tract Cancer, *Cancer Research and Treatment*, Vol. 53, No. 1, 162–171. doi:10.4143/crt.2020.704.
47. Wu, Y., Cheng, Y., Wang, X., Fan, J., and Gao, Q. (2022). Spatial Omics: Navigating to the Golden Era of Cancer Research, *Clinical and Translational Medicine*, Vol. 12, No. 1. doi:10.1002/ctm2.696.
48. Liu, M., Kuo, F., Capistrano, K. J., Kang, D., Nixon, B. G., Shi, W., Chou, C., Do, M. H., Stamatiades, E. G., Gao, S., Li, S., Chen, Y., Hsieh, J. J., Hakimi, A. A., Taniuchi, I., Chan, T. A., and Li, M. O. (2020). TGF- β Suppresses Type 2 Immunity to Cancer, *Nature*, Vol. 587, No. 7832, 115–120. doi:10.1038/s41586-020-2836-1.
49. Rodriguez, A. B., and Engelhard, V. H. (2020). Insights into Tumor-Associated Tertiary Lymphoid Structures: Novel Targets for Antitumor Immunity and Cancer Immunotherapy, *Cancer Immunology Research*, Vol. 8, No. 11, 1338–1345. doi:10.1158/2326-6066.CIR-20-0432.
50. Sautès-Fridman, C., Petitprez, F., Calderaro, J., and Fridman, W. H. (2019). Tertiary Lymphoid Structures in the Era of Cancer Immunotherapy, *Nature Reviews Cancer*, Vol. 19, No. 6, 307–325. doi:10.1038/s41568-019-0144-6.
51. Di Caro, G., Bergomas, F., Grizzi, F., Doni, A., Bianchi, P., Malesci, A., Laghi, L., Allavena, P., Mantovani, A., and Marchesi, F. (2014). Occurrence of Tertiary Lymphoid Tissue Is Associated with T-Cell Infiltration and Predicts Better Prognosis in Early-Stage Colorectal Cancers, *Clinical Cancer Research*, Vol. 20, No. 8, 2147–2158. doi:10.1158/1078-0432.CCR-13-2590.
52. Bergomas, F., Grizzi, F., Doni, A., Pesce, S., Laghi, L., Allavena, P., Mantovani, A., and Marchesi, F. (2011). Tertiary Intratumor Lymphoid Tissue in Colo-Rectal Cancer, *Cancers*, Vol. 4, No. 1, 1–10. doi:10.3390/cancers4010001.
53. Dai, N.-N., Hu, M.-Y., Wang, J.-P., Dai, Z.-H., Xu, L., and Ye, T.-W. (2025). Tertiary Lymphoid Structures in the Microenvironment of Colorectal Cancer: Exploring New Therapeutic Targets, *Cancer Immunology, Immunotherapy*, Vol. 74, No. 8, 245. doi:10.1007/s00262-025-04108-x.
54. Lv, J., Zhang, X., Zhou, M., Yan, J., Chao, G., and Zhang, S. (2024). Tertiary Lymphoid Structures in Colorectal Cancer, *Annals of Medicine*, Vol. 56, No. 1. doi:10.1080/07853890.2024.2400314.
55. Gajewski, T. F., Schreiber, H., and Fu, Y.-X. (2013). Innate and Adaptive Immune Cells in the Tumor Microenvironment, *Nature Immunology*, Vol. 14, No. 10, 1014–1022. doi:10.1038/ni.2703.
56. Vesely, M. D., Kershaw, M. H., Schreiber, R. D., and Smyth, M. J. (2011). Natural Innate and Adaptive Immunity to Cancer, *Annual Review of Immunology*, Vol. 29, No. 1, 235–271. doi:10.1146/annurev-immunol-031210-101324.
57. Nagtegaal, I. D., Quirke, P., and Schmolli, H.-J. (2012). Has the New TNM Classification for Colorectal Cancer Improved Care?, *Nature Reviews Clinical Oncology*, Vol. 9, No. 2, 119–123. doi:10.1038/nrclinonc.2011.157.
58. Fridman, W. H., Pagès, F., Sautès-Fridman, C., and Galon, J. (2012). The Immune Contexture in Human Tumours: Impact on Clinical Outcome, *Nature Reviews Cancer*, Vol. 12, No. 4, 298–306. doi:10.1038/nrc3245.
59. Galon, J., Costes, A., Sanchez-Cabo, F., Kirilovsky, A., Mlecnik, B., Lagorce-Pagès, C., Tosolini, M., Camus, M., Berger, A., Wind, P., Zinzindohoué, F., Bruneval, P., Cugnenc, P.-H., Trajanoski, Z., Fridman, W.-H., and Pagès, F. (2006). Type, Density, and Location of Immune Cells within Human Colorectal Tumors Predict Clinical Outcome, *Science*, Vol. 313, No. 5795, 1960–1964. doi:10.1126/science.1129139.
60. Galon, J., Mlecnik, B., Marliot, F., Ou, F.-S., Bifulco, C. B., Lugli, A., Zlobec, I., Rau, T. T., Hartmann, A., Masucci, G. V., Zavadvova, E., Ohashi, P., Roehrl, M. H. A., Kawakami, Y., Torigoe, T., Ascierto, P. A., Marincola, F., Sargent, D. J., Fox, B. A., and Pages, F. (2016). Validation of the Immunoscore (IM) as a Prognostic Marker in

- Stage I/II/III Colon Cancer: Results of a Worldwide Consortium-Based Analysis of 1,336 Patients., *Journal of Clinical Oncology*, Vol. 34, No. 15_suppl, 3500-3500. doi:10.1200/JCO.2016.34.15_suppl.3500.
61. Pagès, F., Mlecnik, B., Marliot, F., Bindea, G., Ou, F.-S., Bifulco, C., Lugli, A., Zlobec, I., Rau, T. T., Berger, M. D., Nagtegaal, I. D., Vink-Börger, E., Hartmann, A., Geppert, C., Kolwelter, J., Merkel, S., Grützmann, R., Van den Eynde, M., Jouret-Mourin, A., Kartheuser, A., Léonard, D., Remue, C., Wang, J. Y., Bavi, P., Roehrl, M. H. A., Ohashi, P. S., Nguyen, L. T., Han, S., MacGregor, H. L., Hafezi-Bakhtiari, S., Wouters, B. G., Masucci, G. V., Andersson, E. K., Zavadova, E., Vocka, M., Spacek, J., Petruzelka, L., Konopasek, B., Dunder, P., Skalova, H., Nemejcova, K., Botti, G., Tatangelo, F., Delrio, P., Ciliberto, G., Maio, M., Laghi, L., Grizzi, F., Fredriksen, T., Buttard, B., Angelova, M., Vasaturo, A., Maby, P., Church, S. E., Angell, H. K., Lafontaine, L., Bruni, D., El Sissy, C., Haicheur, N., Kirilovsky, A., Berger, A., Lagorce, C., Meyers, J. P., Paustian, C., Feng, Z., Ballesteros-Merino, C., Dijkstra, J., van de Water, C., van Lent-van Vliet, S., Knijn, N., Mušinā, A.-M., Scripcariu, D.-V., Popivanova, B., Xu, M., Fujita, T., Hazama, S., Suzuki, N., Nagano, H., Okuno, K., Torigoe, T., Sato, N., Furuhashi, T., Takemasa, I., Itoh, K., Patel, P. S., Vora, H. H., Shah, B., Patel, J. B., Rajvik, K. N., Pandya, S. J., Shukla, S. N., Wang, Y., Zhang, G., Kawakami, Y., Marincola, F. M., Ascierto, P. A., Sargent, D. J., Fox, B. A., and Galon, J. (2018). International Validation of the Consensus Immunoscore for the Classification of Colon Cancer: A Prognostic and Accuracy Study, *The Lancet*, Vol. 391, No. 10135, 2128-2139. doi:10.1016/S0140-6736(18)30789-X.
 62. Mlecnik, B., Tosolini, M., Kirilovsky, A., Berger, A., Bindea, G., Meatchi, T., Bruneval, P., Trajanoski, Z., Fridman, W.-H., Pagès, F., and Galon, J. (2011). Histopathologic-Based Prognostic Factors of Colorectal Cancers Are Associated with the State of the Local Immune Reaction, *Journal of Clinical Oncology*, Vol. 29, No. 6, 610-618. doi:10.1200/JCO.2010.30.5425.
 63. Pagès, F., Kirilovsky, A., Mlecnik, B., Asslaber, M., Tosolini, M., Bindea, G., Lagorce, C., Wind, P., Marliot, F., Bruneval, P., Zatloukal, K., Trajanoski, Z., Berger, A., Fridman, W.-H., and Galon, J. (2009). In Situ Cytotoxic and Memory T Cells Predict Outcome in Patients with Early-Stage Colorectal Cancer, *Journal of Clinical Oncology*, Vol. 27, No. 35, 5944-5951. doi:10.1200/JCO.2008.19.6147.
 64. Angell, H., and Galon, J. (2013). From the Immune Contexture to the Immunoscore: The Role of Prognostic and Predictive Immune Markers in Cancer, *Current Opinion in Immunology*, Vol. 25, No. 2, 261-267. doi:10.1016/j.coi.2013.03.004.
 65. Asaoka, Y., Ijichi, H., and Koike, K. (2015). PD-1 Blockade in Tumors with Mismatch-Repair Deficiency, *New England Journal of Medicine*, Vol. 373, No. 20, 1979-1979. doi:10.1056/NEJMc1510353.
 66. Overman, M. J., McDermott, R., Leach, J. L., Lonardi, S., Lenz, H.-J., Morse, M. A., Desai, J., Hill, A., Axelson, M., Moss, R. A., Goldberg, M. V., Cao, Z. A., Ledezne, J.-M., Maglente, G. A., Kopetz, S., and André, T. (2017). Nivolumab in Patients with Metastatic DNA Mismatch Repair-Deficient or Microsatellite Instability-High Colorectal Cancer (CheckMate 142): An Open-Label, Multicentre, Phase 2 Study, *The Lancet Oncology*, Vol. 18, No. 9, 1182-1191. doi:10.1016/S1470-2045(17)30422-9.
 67. Carlsen, L., Huntington, K. E., and El-Deiry, W. S. (2022). Immunotherapy for Colorectal Cancer: Mechanisms and Predictive Biomarkers, *Cancers*, Vol. 14, No. 4, 1028. doi:10.3390/cancers14041028.
 68. André, T., Shiu, K.-K., Kim, T. W., Jensen, B. V., Jensen, L. H., Punt, C., Smith, D., Garcia-Carbonero, R., Benavides, M., Gibbs, P., de la Fouchardiere, C., Rivera, F., Elez, E., Bendell, J., Le, D. T., Yoshino, T., Van Cutsem, E., Yang, P., Farooqui, M. Z. H., Marinello, P., and Diaz, L. A. (2020). Pembrolizumab in Microsatellite-Instability-High Advanced Colorectal Cancer, *New England Journal of Medicine*, Vol. 383, No. 23, 2207-2218. doi:10.1056/NEJMoa2017699.
 69. Berry, J., Vreeland, T., Trappey, A., Hale, D., Peace, K., Tyler, J., Walker, A., Brown, R., Herbert, G., Yi, F., Jackson, D., Clifton, G., and Peoples, G. E. (2017). Cancer Vaccines in Colon and Rectal Cancer over the Last Decade: Lessons Learned and Future Directions, *Expert Review of Clinical Immunology*, Vol. 13, No. 3, 235-245. doi:10.1080/1744666X.2016.1226132.
 70. Huang, E.-Y., Chang, J.-C., Chen, H.-H., Hsu, C.-Y., Hsu, H.-C., and Wu, K.-L. (2018). Carcinoembryonic Antigen as a Marker of Radioresistance in Colorectal Cancer: A Potential Role of Macrophages, *BMC Cancer*, Vol. 18, No. 1, 321. doi:10.1186/s12885-018-4254-4.
 71. Orecchioni, M., Ghosheh, Y., Pramod, A. B., and Ley, K. (2019). Macrophage Polarization: Different Gene Signatures in M1(LPS+) vs. Classically and M2(LPS-) vs. Alternatively Activated Macrophages, *Frontiers in Immunology*, Vol. 10. doi:10.3389/fimmu.2019.01084.
 72. Cheng, Y., Zhu, Y., Xu, W., Xu, J., Yang, M., Chen, P., Zhao, J., Geng, L., and Gong, S. (2018). PKC α in Colon Cancer Cells Promotes M1 Macrophage Polarization via MKK3/6-P38 MAPK Pathway, *Molecular Carcinogenesis*, Vol. 57, No. 8, 1017-1029. doi:10.1002/mc.22822.
 73. Zhu, W., Yu, J., Nie, Y., Shi, X., Liu, Y., Li, F., and Zhang, X. (2014). Disequilibrium of M1 and M2 Macrophages Correlates with the Development of Experimental Inflammatory Bowel Diseases, *Immunological Investigations*, Vol. 43, No. 7, 638-652. doi:10.3109/08820139.2014.909456.
 74. Schmitt, M., and Greten, F. R. (2021). The Inflammatory Pathogenesis of Colorectal Cancer, *Nature Reviews Immunology*, Vol. 21, No. 10, 653-667. doi:10.1038/s41577-021-00534-x.
 75. Grivennikov, S. I., Greten, F. R., and Karin, M. (2010). Immunity, Inflammation, and Cancer, *Cell*, Vol. 140, No. 6, 883-899. doi:10.1016/j.cell.2010.01.025.
 76. Sacdalan, D. B., and Lucero, J. A. (2021). The Association between Inflammation and Immunosuppression: Implications for ICI Biomarker Development, *OncoTargets and Therapy*, Vol. Volume 14, 2053-2064. doi:10.2147/OTT.S278089.
 77. Nigam, M., Mishra, A. P., Deb, V. K., Dimri, D. B., Tiwari, V., Bungau, S. G., Bungau, A. F., and Radu, A.-F. (2023). Evaluation of the Association of Chronic Inflammation and Cancer: Insights and Implications, *Biomedicine & Pharmacotherapy*, Vol. 164, 115015. doi:10.1016/j.biopha.2023.115015.
 78. Wang, D., and DuBois, R. N. (2015). Immunosuppression Associated with Chronic Inflammation in the Tumor Microenvironment, *Carcinogenesis*, Vol. 36, No. 10, 1085-1093. doi:10.1093/carcin/bgv123.
 79. Lu, H., Ouyang, W., and Huang, C. (2006). Inflammation, a Key Event in Cancer Development, *Molecular Cancer Research*, Vol. 4, No. 4, 221-233. doi:10.1158/1541-7786.MCR-05-0261.
 80. Lin, R., Zhang, C., Zheng, J., Tian, D., Lei, Z., Chen, D., Xu, Z., and Su, M. (2016). Chronic Inflammation-Associated Genomic Instability Paves the Way for Human Esophageal Carcinogenesis, *Oncotarget*, Vol. 7, No. 17, 24564-24571. doi:10.18632/oncotarget.8356.
 81. Han, Y., Liu, D., and Li, L. (2020). PD-1/PD-L1 Pathway: Current Researches in Cancer., *American Journal of Cancer Research*, Vol. 10, No. 3, 727-742.
 82. Wu, X., Gu, Z., Chen, Y., Chen, B., Chen, W., Weng, L., and Liu, X. (2019). Application of PD-1 Blockade in Cancer Immunotherapy, *Computational and Structural Biotechnology Journal*, Vol. 17, 661-674. doi:10.1016/j.csbj.2019.03.006.

83. Wu, Y., Chen, W., Xu, Z. P., and Gu, W. (2019). PD-L1 Distribution and Perspective for Cancer Immunotherapy—Blockade, Knockdown, or Inhibition, *Frontiers in Immunology*, Vol. 10. doi:10.3389/fimmu.2019.02022.
84. Goswami, K. K., Ghosh, T., Ghosh, S., Sarkar, M., Bose, A., and Baral, R. (2017). Tumor Promoting Role of Anti-Tumor Macrophages in Tumor Microenvironment, *Cellular Immunology*, Vol. 316, 1–10. doi:10.1016/j.cellimm.2017.04.005.
85. Zhang, W., Chen, L., Ma, K., Zhao, Y., Liu, X., Wang, Y., Liu, M., Liang, S., Zhu, H., and Xu, N. (2016). Polarization of Macrophages in the Tumor Microenvironment Is Influenced by EGFR Signaling within Colon Cancer Cells, *Oncotarget*, Vol. 7, No. 46, 75366–75378. doi:10.18632/oncotarget.12207.
86. Edin, S., Wikberg, M. L., Rutegård, J., Oldenborg, P.-A., and Palmqvist, R. (2013). Phenotypic Skewing of Macrophages in Vitro by Secreted Factors from Colorectal Cancer Cells, *PLoS ONE*, Vol. 8, No. 9, e74982. doi:10.1371/journal.pone.0074982.
87. Wu, Y., Yuan, L., Lu, Q., Xu, H., and He, X. (2018). Distinctive Profiles of Tumor-Infiltrating Immune Cells and Association with Intensity of Infiltration in Colorectal Cancer, *Oncology Letters*. doi:10.3892/ol.2018.7771.
88. Barbera-Guillem, E., Nyhus, J. K., Wolford, C. C., Friece, C. R., and Sampsel, J. W. (2002). Vascular Endothelial Growth Factor Secretion by Tumor-Infiltrating Macrophages Essentially Supports Tumor Angiogenesis, and IgG Immune Complexes Potentiate the Process., *Cancer Research*, Vol. 62, No. 23, 7042–9.
89. Suarez-Lopez, L., Sriram, G., Kong, Y. W., Morandell, S., Merrick, K. A., Hernandez, Y., Haigis, K. M., and Yaffe, M. B. (2018). MK2 Contributes to Tumor Progression by Promoting M2 Macrophage Polarization and Tumor Angiogenesis, *Proceedings of the National Academy of Sciences*, Vol. 115, No. 18. doi:10.1073/pnas.1722020115.
90. Burmeister, K., Quagliata, L., Andreozzi, M., Eppenberger-Castori, S., Matter, M. S., Perrina, V., Grobholz, R., Jochum, W., Horber, D., Moosmann, P., Lehmann, F., Köberle, D., Ng, C. K. Y., Piscuoglio, S., Tornillo, L., and Terracciano, L. M. (2017). Vascular Endothelial Growth Factor A Amplification in Colorectal Cancer Is Associated with Reduced M1 and M2 Macrophages and Diminished PD-1-Expressing Lymphocytes, *PLOS ONE*, Vol. 12, No. 4, e0175563. doi:10.1371/journal.pone.0175563.
91. Illemann, M., Bird, N., Majeed, A., Sehested, M., Laerum, O. D., Lund, L. R., Danø, K., and Nielsen, B. S. (2006). MMP-9 Is Differentially Expressed in Primary Human Colorectal Adenocarcinomas and Their Metastases, *Molecular Cancer Research*, Vol. 4, No. 5, 293–302. doi:10.1158/1541-7786.MCR-06-0003.
92. Cai, J., Xia, L., Li, J., Ni, S., Song, H., and Wu, X. (2019). Tumor-Associated Macrophages Derived TGF- β -induced Epithelial to Mesenchymal Transition in Colorectal Cancer Cells through Smad2,3-4/Snail Signaling Pathway, *Cancer Research and Treatment*, Vol. 51, No. 1, 252–266. doi:10.4143/crt.2017.613.
93. Wei, C., Yang, C., Wang, S., Shi, D., Zhang, C., Lin, X., Liu, Q., Dou, R., and Xiong, B. (2019). Crosstalk between Cancer Cells and Tumor Associated Macrophages Is Required for Mesenchymal Circulating Tumor Cell-Mediated Colorectal Cancer Metastasis, *Molecular Cancer*, Vol. 18, No. 1, 64. doi:10.1186/s12943-019-0976-4.
94. Afik, R., Zigmund, E., Vugman, M., Klepfish, M., Shimshoni, E., Pasmanik-Chor, M., Shenoy, A., Bassat, E., Halpern, Z., Geiger, T., Sagi, I., and Varol, C. (2016). Tumor Macrophages Are Pivotal Constructors of Tumor Collagenous Matrix, *Journal of Experimental Medicine*, Vol. 213, No. 11, 2315–2331. doi:10.1084/jem.20151193.
95. Wei, C., Yang, C., Wang, S., Shi, D., Zhang, C., Lin, X., and Xiong, B. (2019). M2 Macrophages Confer Resistance to 5-Fluorouracil in Colorectal Cancer through the Activation of CCL22/PI3K/AKT Signaling, *OncoTargets and Therapy*, Vol. Volume 12, 3051–3063. doi:10.2147/OTT.S198126.
96. WANG, W., LI, X., ZHENG, D., ZHANG, D., PENG, X., ZHANG, X., AI, F., WANG, X., MA, J., XIONG, W., LI, G., ZHOU, Y., and SHEN, S. (2015). Dynamic Changes and Functions of Macrophages and M1/M2 Subpopulations during Ulcerative Colitis-Associated Carcinogenesis in an AOM/DSS Mouse Model, *Molecular Medicine Reports*, Vol. 11, No. 4, 2397–2406. doi:10.3892/mmr.2014.3018.
97. Zhong, X., Chen, B., and Yang, Z. (2018). The Role of Tumor-Associated Macrophages in Colorectal Carcinoma Progression, *Cellular Physiology and Biochemistry*, Vol. 45, No. 1, 356–365. doi:10.1159/000486816.
98. Narayanan, S., Kawaguchi, T., Peng, X., Qi, Q., Liu, S., Yan, L., and Takabe, K. (2019). Tumor Infiltrating Lymphocytes and Macrophages Improve Survival in Microsatellite Unstable Colorectal Cancer, *Scientific Reports*, Vol. 9, No. 1, 13455. doi:10.1038/s41598-019-49878-4.
99. Lin, A., Zhang, J., and Luo, P. (2020). Crosstalk between the MSI Status and Tumor Microenvironment in Colorectal Cancer, *Frontiers in Immunology*, Vol. 11. doi:10.3389/fimmu.2020.02039.
100. Zhang, J., Zhu, H., Liu, W., Miao, J., Mao, Y., and Li, Q. (2025). Prognostic and Predictive Molecular Biomarkers in Colorectal Cancer, *Frontiers in Oncology*, Frontiers Media SA. doi:10.3389/fonc.2025.1532924.
101. Lv, Q., Zhang, Y., Gao, W., Wang, J., Hu, Y., Yang, H., Xie, Y., Lv, Y., Zhang, H., Wu, D., Hu, L., and Wang, J. (2024). CSF1R Inhibition Reprograms Tumor-Associated Macrophages to Potentiate Anti-PD-1 Therapy Efficacy against Colorectal Cancer, *Pharmacological Research*, Vol. 202, 107126. doi:10.1016/j.phrs.2024.107126.
102. Zhu, M., Bai, L., Liu, X., Peng, S., Xie, Y., Bai, H., Yu, H., Wang, X., Yuan, P., Ma, R., Lin, J., Wu, L., Huang, M., Li, Y., and Luo, Y. (2022). Silence of a Dependence Receptor CSF1R in Colorectal Cancer Cells Activates Tumor-Associated Macrophages, *Journal for ImmunoTherapy of Cancer*, Vol. 10, No. 12, e005610. doi:10.1136/jitc-2022-005610.
103. Shimizu, D., Yuge, R., Kitadai, Y., Ariyoshi, M., Miyamoto, R., Hiyama, Y., Takigawa, H., Urabe, Y., and Oka, S. (2024). Pexidartinib and Immune Checkpoint Inhibitors Combine to Activate Tumor Immunity in a Murine Colorectal Cancer Model by Depleting M2 Macrophages Differentiated by Cancer-Associated Fibroblasts, *International Journal of Molecular Sciences*, Vol. 25, No. 13, 7001. doi:10.3390/ijms25137001.
104. Wang, S., Sun, J., Chen, K., Ma, P., Lei, Q., Xing, S., Cao, Z., Sun, S., Yu, Z., Liu, Y., and Li, N. (2021). Perspectives of Tumor-Infiltrating Lymphocyte Treatment in Solid Tumors, *BMC Medicine*, Vol. 19, No. 1, 140. doi:10.1186/s12916-021-02006-4.
105. Ropponen, K. M., Eskelinen, M. J., Lippinen, P. K., Alhava, E., and Kosma, V. M. (1997). Prognostic Value of Tumour-Infiltrating Lymphocytes (TILs) in Colorectal Cancer., *The Journal of Pathology*, Vol. 182, No. 3, 318–24. doi:10.1002/(SICI)1096-9896(199707)182:3<318::AID-PATH862>3.0.CO;2-6.
106. Kuwahara, T., Hazama, S., Suzuki, N., Yoshida, S., Tomochika, S., Nakagami, Y., Matsui, H., Shindo, Y., Kanekiyo, S., Tokumitsu, Y., Iida, M., Tsunedomi, R., Takeda, S., Yoshino, S., Okayama, N., Suehiro, Y., Yamasaki, T., Fujita, T., Kawakami, Y., Ueno, T., and Nagano, H. (2019). Intratumoural-Infiltrating CD4+ and FOXP3+ T Cells as Strong Positive Predictive Markers for the Prognosis of Resectable Colorectal Cancer, *British Journal of*

- Cancer*, Vol. 121, No. 8, 659–665. doi:[10.1038/s41416-019-0559-6](https://doi.org/10.1038/s41416-019-0559-6).
107. Ahrends, T., Spanjaard, A., Pilzecker, B., Bąbała, N., Bovens, A., Xiao, Y., Jacobs, H., and Borst, J. (2017). CD4+ T Cell Help Confers a Cytotoxic T Cell Effector Program Including Coinhibitory Receptor Downregulation and Increased Tissue Invasiveness, *Immunity*, Vol. 47, No. 5, 848–861.e5. doi:[10.1016/j.immuni.2017.10.009](https://doi.org/10.1016/j.immuni.2017.10.009).
 108. Ling, A., Lundberg, I. V., Eklöf, V., Wikberg, M. L., Öberg, Å., Edin, S., and Palmqvist, R. (2016). The Infiltration, and Prognostic Importance, of Th1 Lymphocytes Vary in Molecular Subgroups of Colorectal Cancer, *The Journal of Pathology: Clinical Research*, Vol. 2, No. 1, 21–31. doi:[10.1002/cjp2.31](https://doi.org/10.1002/cjp2.31).
 109. Tosolini, M., Kirilovsky, A., Mlecnik, B., Fredriksen, T., Mauger, S., Bindea, G., Berger, A., Bruneval, P., Fridman, W.-H., Pagès, F., and Galon, J. (2011). Clinical Impact of Different Classes of Infiltrating T Cytotoxic and Helper Cells (Th1, Th2, Treg, Th17) in Patients with Colorectal Cancer, *Cancer Research*, Vol. 71, No. 4, 1263–1271. doi:[10.1158/0008-5472.CAN-10-2907](https://doi.org/10.1158/0008-5472.CAN-10-2907).
 110. De Simone, V., Pallone, F., Monteleone, G., and Stolfi, C. (2013). Role of TH17 Cytokines in the Control of Colorectal Cancer, *Oncolmmunology*, Vol. 2, No. 12, e26617. doi:[10.4161/onci.26617](https://doi.org/10.4161/onci.26617).
 111. Glaire, M. A., Domingo, E., Sveen, A., Bruun, J., Nesbakken, A., Nicholson, G., Novelli, M., Lawson, K., Oukrif, D., Kildal, W., Danielsen, H. E., Kerr, R., Kerr, D., Tomlinson, I., Lothe, R. A., and Church, D. N. (2019). Tumour-Infiltrating CD8+ Lymphocytes and Colorectal Cancer Recurrence by Tumour and Nodal Stage, *British Journal of Cancer*, Vol. 121, No. 6, 474–482. doi:[10.1038/s41416-019-0540-4](https://doi.org/10.1038/s41416-019-0540-4).
 112. Jochems, C., and Schlom, J. (2011). Tumor-Infiltrating Immune Cells and Prognosis: The Potential Link between Conventional Cancer Therapy and Immunity, *Experimental Biology and Medicine*, Vol. 236, No. 5, 567–579. doi:[10.1258/ebm.2011.011007](https://doi.org/10.1258/ebm.2011.011007).
 113. Angell, H. K., Bruni, D., Barrett, J. C., Herbst, R., and Galon, J. (2020). The Immunoscore: Colon Cancer and Beyond, *Clinical Cancer Research*, Vol. 26, No. 2, 332–339. doi:[10.1158/1078-0432.CCR-18-1851](https://doi.org/10.1158/1078-0432.CCR-18-1851).
 114. O'Malley, G., Treacy, O., Lynch, K., Naicker, S. D., Leonard, N. A., Lohan, P., Dunne, P. D., Ritter, T., Egan, L. J., and Ryan, A. E. (2018). Stromal Cell PD-L1 Inhibits CD8+ T-Cell Antitumor Immune Responses and Promotes Colon Cancer, *Cancer Immunology Research*, Vol. 6, No. 11, 1426–1441. doi:[10.1158/2326-6066.CIR-17-0443](https://doi.org/10.1158/2326-6066.CIR-17-0443).
 115. Shan, T., Chen, S., Wu, T., Yang, Y., Li, S., and Chen, X. (2019). PD-L1 Expression in Colon Cancer and Its Relationship with Clinical Prognosis., *International Journal of Clinical and Experimental Pathology*, Vol. 12, No. 5, 1764–1769.
 116. Zhao, T., Li, Y., Zhang, J., and Zhang, B. (2020). PD-L1 Expression Increased by IFN-γ via JAK2-STAT1 Signaling and Predicts a Poor Survival in Colorectal Cancer, *Oncology Letters*, Vol. 20, No. 2, 1127–1134. doi:[10.3892/ol.2020.11647](https://doi.org/10.3892/ol.2020.11647).
 117. Elfshawy, M., Abd-ELaziz, S. A., Hegazy, A., and El-Yasergy, D. F. (2020). Immunohistochemical Expression of Programmed Death Ligand-1 (PDL-1) in Colorectal Carcinoma and Its Correlation with Stromal Tumor Infiltrating Lymphocytes, *Asian Pacific Journal of Cancer Prevention*, Vol. 21, No. 1, 225–232. doi:[10.31557/APJCP.2020.21.1.225](https://doi.org/10.31557/APJCP.2020.21.1.225).
 118. Thomas, D. A., and Massagué, J. (2005). TGF-β Directly Targets Cytotoxic T Cell Functions during Tumor Evasion of Immune Surveillance, *Cancer Cell*, Vol. 8, No. 5, 369–380. doi:[10.1016/j.ccr.2005.10.012](https://doi.org/10.1016/j.ccr.2005.10.012).
 119. Zhao, S., Jiang, T., Zhang, L., Yang, H., Liu, X., Jia, Y., and Zhou, C. (2016). Clinicopathological and Prognostic Significance of Regulatory T Cells in Patients with Non-Small Cell Lung Cancer: A Systematic Review with Meta-Analysis, *Oncotarget*, Vol. 7, No. 24, 36065–36073. doi:[10.18632/oncotarget.9130](https://doi.org/10.18632/oncotarget.9130).
 120. Tang, Y., Xu, X., Guo, S., Zhang, C., Tang, Y., Tian, Y., Ni, B., Lu, B., and Wang, H. (2014). An Increased Abundance of Tumor-Infiltrating Regulatory T Cells Is Correlated with the Progression and Prognosis of Pancreatic Ductal Adenocarcinoma, *PLoS ONE*, Vol. 9, No. 3, e91551. doi:[10.1371/journal.pone.0091551](https://doi.org/10.1371/journal.pone.0091551).
 121. Merlo, A., Casalini, P., Carcangiu, M. L., Malventano, C., Triulzi, T., Mènard, S., Tagliabue, E., and Balsari, A. (2009). FOXP3 Expression and Overall Survival in Breast Cancer, *Journal of Clinical Oncology*, Vol. 27, No. 11, 1746–1752. doi:[10.1200/JCO.2008.17.9036](https://doi.org/10.1200/JCO.2008.17.9036).
 122. Fontenot, J. D., Gavin, M. A., and Rudensky, A. Y. (2003). Foxp3 Programs the Development and Function of CD4+CD25+ Regulatory T Cells, *Nature Immunology*, Vol. 4, No. 4, 330–336. doi:[10.1038/ni904](https://doi.org/10.1038/ni904).
 123. Hori, S., Nomura, T., and Sakaguchi, S. (2003). Control of Regulatory T Cell Development by the Transcription Factor Foxp3, *Science*, Vol. 299, No. 5609, 1057–1061. doi:[10.1126/science.1079490](https://doi.org/10.1126/science.1079490).
 124. Gavin, M. A., Rasmussen, J. P., Fontenot, J. D., Vasta, V., Manganiello, V. C., Beavo, J. A., and Rudensky, A. Y. (2007). Foxp3-Dependent Programme of Regulatory T-Cell Differentiation, *Nature*, Vol. 445, No. 7129, 771–775. doi:[10.1038/nature05543](https://doi.org/10.1038/nature05543).
 125. Zheng, Y., Josefowicz, S. Z., Kas, A., Chu, T.-T., Gavin, M. A., and Rudensky, A. Y. (2007). Genome-Wide Analysis of Foxp3 Target Genes in Developing and Mature Regulatory T Cells, *Nature*, Vol. 445, No. 7130, 936–940. doi:[10.1038/nature05563](https://doi.org/10.1038/nature05563).
 126. Olguín, J. E., Medina-Andrade, I., Rodríguez, T., Rodríguez-Sosa, M., and Terrazas, L. I. (2020). Relevance of Regulatory T Cells during Colorectal Cancer Development, *Cancers*, Vol. 12, No. 7, 1888. doi:[10.3390/cancers12071888](https://doi.org/10.3390/cancers12071888).
 127. Ji, D., Song, C., Li, Y., Xia, J., Wu, Y., Jia, J., Cui, X., Yu, S., and Gu, J. (2020). Combination of Radiotherapy and Suppression of Tregs Enhances Abscopal Antitumor Effect and Inhibits Metastasis in Rectal Cancer, *Journal for ImmunoTherapy of Cancer*, Vol. 8, No. 2, e000826. doi:[10.1136/jitc-2020-000826](https://doi.org/10.1136/jitc-2020-000826).
 128. Kuwahara, T., Hazama, S., Suzuki, N., Yoshida, S., Tomochika, S., Nakagami, Y., Matsui, H., Shindo, Y., Kanekiyo, S., Tokumitsu, Y., Iida, M., Tsunedomi, R., Takeda, S., Yoshino, S., Okayama, N., Suehiro, Y., Yamasaki, T., Fujita, T., Kawakami, Y., Ueno, T., and Nagano, H. (2019). Intratumoural-Infiltrating CD4+ and FOXP3+ T Cells as Strong Positive Predictive Markers for the Prognosis of Resectable Colorectal Cancer, *British Journal of Cancer*, Vol. 121, No. 8, 659–665. doi:[10.1038/s41416-019-0559-6](https://doi.org/10.1038/s41416-019-0559-6).
 129. Salama, P., Phillips, M., Grieu, F., Morris, M., Zeps, N., Joseph, D., Platell, C., and Iacopetta, B. (2009). Tumor-Infiltrating FOXP3+ T Regulatory Cells Show Strong Prognostic Significance in Colorectal Cancer, *Journal of Clinical Oncology*, Vol. 27, No. 2, 186–192. doi:[10.1200/JCO.2008.18.7229](https://doi.org/10.1200/JCO.2008.18.7229).
 130. van den Bulk, J., de Miranda, N. F. C. C., and ten Dijke, P. (2021). Therapeutic Targeting of TGF-β in Cancer: Hacking a Master Switch of Immune Suppression, *Clinical Science*, Vol. 135, No. 1, 35–52. doi:[10.1042/CS20201236](https://doi.org/10.1042/CS20201236).
 131. Colak, S., and ten Dijke, P. (2017). Targeting TGF-β Signaling in Cancer, *Trends in Cancer*, Vol. 3, No. 1, 56–71. doi:[10.1016/j.trecan.2016.11.008](https://doi.org/10.1016/j.trecan.2016.11.008).
 132. Principe, D. R., Doll, J. A., Bauer, J., Jung, B., Munshi, H. G., Bartholin, L., Pasche, B., Lee, C., and Grippo, P. J. (2014). TGF- :

- Duality of Function between Tumor Prevention and Carcinogenesis, *JNCI Journal of the National Cancer Institute*, Vol. 106, No. 2, djt369–djt369. doi:10.1093/jnci/djt369.
133. Heldin, C.-H., Vanlandewijck, M., and Moustakas, A. (2012). Regulation of EMT by TGF β in Cancer, *FEBS Letters*, Vol. 586, No. 14, 1959–1970. doi:10.1016/j.febslet.2012.02.037.
134. Du, B., and Shim, J. (2016). Targeting Epithelial–Mesenchymal Transition (EMT) to Overcome Drug Resistance in Cancer, *Molecules*, Vol. 21, No. 7, 965. doi:10.3390/molecules21070965.
135. Mariathasan, S., Turley, S. J., Nickles, D., Castiglioni, A., Yuen, K., Wang, Y., Kadel III, E. E., Koeppen, H., Astarita, J. L., Cubas, R., Jhunjhunwala, S., Banchereau, R., Yang, Y., Guan, Y., Chalouni, C., Ziai, J., Şenbabaoğlu, Y., Santoro, S., Sheinson, D., Hung, J., Giltmane, J. M., Pierce, A. A., Mesh, K., Lianoglou, S., Riegler, J., Carano, R. A. D., Eriksson, P., Höglund, M., Somarriba, L., Halligan, D. L., van der Heijden, M. S., Loriot, Y., Rosenberg, J. E., Fong, L., Mellman, I., Chen, D. S., Green, M., Derleth, C., Fine, G. D., Hegde, P. S., Bourgon, R., and Powles, T. (2018). TGF β Attenuates Tumour Response to PD-L1 Blockade by Contributing to Exclusion of T Cells, *Nature*, Vol. 554, No. 7693, 544–548. doi:10.1038/nature25501.
136. Ferrari, G., Cook, B. D., Terushkin, V., Pintucci, G., and Mignatti, P. (2009). Transforming Growth Factor-beta 1 (TGF- β 1) Induces Angiogenesis through Vascular Endothelial Growth Factor (VEGF)-mediated Apoptosis, *Journal of Cellular Physiology*, Vol. 219, No. 2, 449–458. doi:10.1002/jcp.21706.
137. Ghahremanifard, P., Chanda, A., Bonni, S., and Bose, P. (2020). TGF- β Mediated Immune Evasion in Cancer—Spotlight on Cancer-Associated Fibroblasts, *Cancers*, Vol. 12, No. 12, 3650. doi:10.3390/cancers12123650.
138. Saito, T., Nishikawa, H., Wada, H., Nagano, Y., Sugiyama, D., Atarashi, K., Maeda, Y., Hamaguchi, M., Ohkura, N., Sato, E., Nagase, H., Nishimura, J., Yamamoto, H., Takiguchi, S., Tanoue, T., Suda, W., Morita, H., Hattori, M., Honda, K., Mori, M., Doki, Y., and Sakaguchi, S. (2016). Two FOXP3+CD4+ T Cell Subpopulations Distinctly Control the Prognosis of Colorectal Cancers, *Nature Medicine*, Vol. 22, No. 6, 679–684. doi:10.1038/nm.4086.
139. Bai, Z., Zhou, Y., Ye, Z., Xiong, J., Lan, H., and Wang, F. (2022). Tumor-Infiltrating Lymphocytes in Colorectal Cancer: The Fundamental Indication and Application on Immunotherapy, *Frontiers in Immunology*, Vol. 12. doi:10.3389/fimmu.2021.808964.
140. Ooki, A., Shinozaki, E., and Yamaguchi, K. (2021). Immunotherapy in Colorectal Cancer: Current and Future Strategies, *Journal of the Anus, Rectum and Colon*, Vol. 5, No. 1, 11–24. doi:10.23922/jarc.2020-064.
141. Arrichiello, G., Poliero, L., Borrelli, C., Paragliola, F., Nacca, V., Napolitano, S., Corte, C. M. Della, Martini, G., and Martinelli, E. (2021). Immunotherapy in Colorectal Cancer: Is the Long-Awaited Revolution Finally Happening?, *Cancer Treatment and Research Communications*, Vol. 28, 100442. doi:10.1016/j.ctarc.2021.100442.
142. Wang, Q., Yu, M., and Zhang, S. (2025). The Characteristics of the Tumor Immune Microenvironment in Colorectal Cancer with Different MSI Status and Current Therapeutic Strategies, *Frontiers in Immunology*, Vol. 15. doi:10.3389/fimmu.2024.1440830.
143. Nair, R., Lannagan, T. R. M., Jackstadt, R., Andrusaita, A., Cole, J., Boyne, C., Nibbs, R. J. B., Sansom, O. J., and Milling, S. (2024). Co-Inhibition of TGF- β and PD-L1 Pathways in a Metastatic Colorectal Cancer Mouse Model Triggers Interferon Responses, Innate Cells and T Cells, alongside Metabolic Changes and Tumor Resistance, *Onc Immunology*, Vol. 13, No. 1. doi:10.1080/2162402X.2024.2330194.
144. Gulley, J. L., Schlom, J., Barcellos-Hoff, M. H., Wang, X., Seoane, J., Audhuy, F., Lan, Y., Dussault, I., and Moustakas, A. (2022). Dual Inhibition of TGF- β and PD-L1: A Novel Approach to Cancer Treatment, *Molecular Oncology*, Vol. 16, No. 11, 2117–2134. doi:10.1002/1878-0261.13146.
145. Kobie, J. J., Wu, R. S., Kurt, R. A., Lou, S., Adelman, M. K., Whitesell, L. J., Ramanathapuram, L. V., Arteaga, C. L., and Akporiaye, E. T. (2003). Transforming Growth Factor Beta Inhibits the Antigen-Presenting Functions and Antitumor Activity of Dendritic Cell Vaccines., *Cancer Research*, Vol. 63, No. 8, 1860–4.
146. Suzuki, A., Masuda, A., Nagata, H., Kameoka, S., Kikawada, Y., Yamakawa, M., and Kasajima, T. (2002). Mature Dendritic Cells Make Clusters with T Cells in the Invasive Margin of Colorectal Carcinoma, *The Journal of Pathology*, Vol. 196, No. 1, 37–43. doi:10.1002/path.1018.
147. Gardner, A., and Ruffell, B. (2016). Dendritic Cells and Cancer Immunity, *Trends in Immunology*, Vol. 37, No. 12, 855–865. doi:10.1016/j.it.2016.09.006.
148. Galati, D., Corazzelli, G., De Filippi, R., and Pinto, A. (2016). Dendritic Cells in Hematological Malignancies, *Critical Reviews in Oncology/Hematology*, Vol. 108, 86–96. doi:10.1016/j.critrevonc.2016.10.006.
149. Truxova, I., Kasikova, L., Hensler, M., Skapa, P., Laco, J., Pecen, L., Belicova, L., Praznovec, I., Halaska, M. J., Brtnicky, T., Salkova, E., Rob, L., Kodet, R., Goc, J., Sautes-Fridman, C., Fridman, W. H., Ryska, A., Galluzzi, L., Spisek, R., and Fucikova, J. (2018). Mature Dendritic Cells Correlate with Favorable Immune Infiltrate and Improved Prognosis in Ovarian Carcinoma Patients, *Journal for Immunotherapy of Cancer*, Vol. 6, No. 1, 139. doi:10.1186/s40425-018-0446-3.
150. Wculek, S. K., Cueto, F. J., Mujal, A. M., Melero, I., Krummel, M. F., and Sancho, D. (2020). Dendritic Cells in Cancer Immunology and Immunotherapy, *Nature Reviews Immunology*, Vol. 20, No. 1, 7–24. doi:10.1038/s41577-019-0210-z.
151. Goc, J., Germain, C., Vo-Bourgais, T. K. D., Lupo, A., Klein, C., Knockaert, S., de Chaisemartin, L., Ouakrim, H., Becht, E., Alifano, M., Validire, P., Remark, R., Hammond, S. A., Cremer, I., Damotte, D., Fridman, W.-H., Sautès-Fridman, C., and Dieu-Nosjean, M.-C. (2014). Dendritic Cells in Tumor-Associated Tertiary Lymphoid Structures Signal a Th1 Cytotoxic Immune Contexture and License the Positive Prognostic Value of Infiltrating CD8+ T Cells, *Cancer Research*, Vol. 74, No. 3, 705–715. doi:10.1158/0008-5472.CAN-13-1342.
152. Carenza, C., Calcaterra, F., Oriolo, F., Di Vito, C., Ubezio, M., Della Porta, M. G., Mavilio, D., and Della Bella, S. (2019). Costimulatory Molecules and Immune Checkpoints Are Differentially Expressed on Different Subsets of Dendritic Cells, *Frontiers in Immunology*, Vol. 10. doi:10.3389/fimmu.2019.01325.
153. Kim, M. K., and Kim, J. (2019). Properties of Immature and Mature Dendritic Cells: Phenotype, Morphology, Phagocytosis, and Migration, *RSC Advances*, Vol. 9, No. 20, 11230–11238. doi:10.1039/C9RA00818G.
154. Anderson, D. A., Murphy, K. M., and Briseño, C. G. (2018). Development, Diversity, and Function of Dendritic Cells in Mouse and Human, *Cold Spring Harbor Perspectives in Biology*, Vol. 10, No. 11, a028613. doi:10.1101/cshperspect.a028613.
155. Bandola-Simon, J., and Roche, P. A. (2019). Dysfunction of Antigen Processing and Presentation by Dendritic Cells in Cancer, *Molecular Immunology*, Vol. 113, 31–37. doi:10.1016/j.molimm.2018.03.025.
156. Mitchell, D., Chintala, S., and Dey, M. (2018). Plasmacytoid Dendritic Cell in Immunity and Cancer, *Journal of Neuroimmunology*, Vol. 322, 63–73. doi:10.1016/j.jneuroim.2018.06.012.

157. Murphy, T. L., and Murphy, K. M. (2022). Dendritic Cells in Cancer Immunology, *Cellular & Molecular Immunology*, Vol. 19, No. 1, 3–13. doi:10.1038/s41423-021-00741-5.
158. Del Prete, A., Salvi, V., Soriani, A., Laffranchi, M., Sozio, F., Bosisio, D., and Sozzani, S. (2023). Dendritic Cell Subsets in Cancer Immunity and Tumor Antigen Sensing, *Cellular & Molecular Immunology*, Vol. 20, No. 5, 432–447. doi:10.1038/s41423-023-00990-6.
159. Li, Y.-R., Fang, Y., Lyu, Z., Zhu, Y., and Yang, L. (2023). Exploring the Dynamic Interplay between Cancer Stem Cells and the Tumor Microenvironment: Implications for Novel Therapeutic Strategies, *Journal of Translational Medicine*, Vol. 21, No. 1, 686. doi:10.1186/s12967-023-04575-9.
160. Marciscano, A. E., and Anandasabapathy, N. (2021). The Role of Dendritic Cells in Cancer and Anti-Tumor Immunity, *Seminars in Immunology*, Vol. 52, 101481. doi:10.1016/j.smim.2021.101481.
161. Mildner, A., and Jung, S. (2014). Development and Function of Dendritic Cell Subsets, *Immunity*, Vol. 40, No. 5, 642–656. doi:10.1016/j.immuni.2014.04.016.
162. Fu, C., and Jiang, A. (2018). Dendritic Cells and CD8 T Cell Immunity in Tumor Microenvironment, *Frontiers in Immunology*, Vol. 9. doi:10.3389/fimmu.2018.03059.
163. Roncarolo, M.-G., Leving, M. K., and Traversari, C. (2001). Differentiation of T Regulatory Cells by Immature Dendritic Cells, *The Journal of Experimental Medicine*, Vol. 193, No. 2, F5–F10. doi:10.1084/jem.193.2.F5.
164. Legitimo, A., Consolini, R., Failli, A., Orsini, G., and Spisni, R. (2014). Dendritic Cell Defects in the Colorectal Cancer, *Human Vaccines & Immunotherapeutics*, Vol. 10, No. 11, 3224–3235. doi:10.4161/hv.29857.
165. Sabado, R. L., Balan, S., and Bhardwaj, N. (2017). Dendritic Cell-Based Immunotherapy, *Cell Research*, Vol. 27, No. 1, 74–95. doi:10.1038/cr.2016.157.
166. Orsini, G., Legitimo, A., Failli, A., Ferrari, P., Nicolini, A., Spisni, R., Miccoli, P., and Consolini, R. (2013). Defective Generation and Maturation of Dendritic Cells from Monocytes in Colorectal Cancer Patients during the Course of Disease, *International Journal of Molecular Sciences*, Vol. 14, No. 11, 22022–22041. doi:10.3390/ijms141122022.
167. Li, J., Yang, J., Hua, L., Wang, R., Li, H., Zhang, C., Zhang, H., Li, S., Zhu, L., and Su, H. (2021). ESE-3 Contributes to Colon Cancer Progression by Downregulating EHD2 and Transactivating INPP4B., *American Journal of Cancer Research*, Vol. 11, No. 1, 92–107.
168. Sprater, F., Azeem, W., and Appel, S. (2014). Activation of Peroxisome Proliferator-activated Receptor Gamma Leads to Upregulation of ESE-3 Expression in Human Monocyte-derived Dendritic Cells, *Scandinavian Journal of Immunology*, Vol. 79, No. 1, 20–26. doi:10.1111/sji.12126.
169. Nagorsen, D., Voigt, S., Berg, E., Stein, H., Thiel, E., and Loddenkemper, C. (2007). Tumor-Infiltrating Macrophages and Dendritic Cells in Human Colorectal Cancer: Relation to Local Regulatory T Cells, Systemic T-Cell Response against Tumor-Associated Antigens and Survival, *Journal of Translational Medicine*, Vol. 5, No. 1, 62. doi:10.1186/1479-5876-5-62.
170. Hsu, Y.-L., Chen, Y.-J., Chang, W.-A., Jian, S.-F., Fan, H.-L., Wang, J.-Y., and Kuo, P.-L. (2018). Interaction between Tumor-Associated Dendritic Cells and Colon Cancer Cells Contributes to Tumor Progression via CXCL1, *International Journal of Molecular Sciences*, Vol. 19, No. 8, 2427. doi:10.3390/ijms19082427.
171. Kießler, M., Plesca, I., Sommer, U., Wehner, R., Wilczkowski, F., Müller, L., Tunger, A., Lai, X., Rentsch, A., Peuker, K., Zeissig, S., Seifert, A. M., Seifert, L., Weitz, J., Bachmann, M., Bornhäuser, M., Aust, D., Baretton, G., and Schmitz, M. (2021). Tumor-Infiltrating Plasmacytoid Dendritic Cells Are Associated with Survival in Human Colon Cancer, *Journal for Immunotherapy of Cancer*, Vol. 9, No. 3, e001813. doi:10.1136/jitc-2020-001813.
172. Ho, W. W., Gomes-Santos, I. L., Aoki, S., Datta, M., Kawaguchi, K., Talele, N. P., Roberge, S., Ren, J., Liu, H., Chen, I. X., Andersson, P., Chatterjee, S., Kumar, A. S., Amoozgar, Z., Zhang, Q., Huang, P., Ng, M. R., Chauhan, V. P., Xu, L., Duda, D. G., Clark, J. W., Pittet, M. J., Fukumura, D., and Jain, R. K. (2021). Dendritic Cell Paucity in Mismatch Repair–Proficient Colorectal Cancer Liver Metastases Limits Immune Checkpoint Blockade Efficacy, *Proceedings of the National Academy of Sciences*, Vol. 118, No. 45. doi:10.1073/pnas.2105323118.
173. Baldin, A. V., Savvateeva, L. V., Bazhin, A. V., and Zamyatnin, A. A. (2020). Dendritic Cells in Anticancer Vaccination: Rationale for Ex Vivo Loading or in Vivo Targeting, *Cancers*, Vol. 12, No. 3, 590. doi:10.3390/cancers12030590.
174. Chang, S.-C., Ke, T.-W., Chen, W. T.-L., Shyu, W.-C., and Jeng, L.-B. (2024). Effect of Autologous Dendritic Cell Cytokine-Induced Killer on Refractory Metastatic Colorectal Cancer: A Matched Case–Control Comparative Study, *Frontiers in Immunology*, Vol. 15. doi:10.3389/fimmu.2024.1329615.
175. Fan, T., Zhang, M., Yang, J., Zhu, Z., Cao, W., and Dong, C. (2023). Therapeutic Cancer Vaccines: Advancements, Challenges and Prospects, *Signal Transduction and Targeted Therapy*, Vol. 8, No. 1, 450. doi:10.1038/s41392-023-01674-3.
176. Nava, S., Lisini, D., Frigerio, S., and Bersano, A. (2021). Dendritic Cells and Cancer Immunotherapy: The Adjuvant Effect, *International Journal of Molecular Sciences*, Vol. 22, No. 22, 12339. doi:10.3390/ijms222212339.
177. Zimmermannova, O., Ferreira, A. G., Ascic, E., Velasco Santiago, M., Kurochkin, I., Hansen, M., Met, Ö., Caiado, I., Shapiro, I. E., Michaux, J., Humbert, M., Soto-Cabrera, D., Benonisson, H., Silvério-Alves, R., Gomez-Jimenez, D., Bernardo, C., Bauden, M., Andersson, R., Höglund, M., Miharada, K., Nakamura, Y., Hugues, S., Greiff, L., Lindstedt, M., Rosa, F. F., Pires, C. F., Bassani-Sternberg, M., Svane, I. M., and Pereira, C.-F. (2023). Restoring Tumor Immunogenicity with Dendritic Cell Reprogramming, *Science Immunology*, Vol. 8, No. 85. doi:10.1126/sciimmunol.add4817.
178. Wagner, S., Mullins, C. S., and Linnebacher, M. (2018). Colorectal Cancer Vaccines: Tumor-Associated Antigens vs Neoantigens, *World Journal of Gastroenterology*, Vol. 24, No. 48, 5418–5432. doi:10.3748/wjg.v24.i48.5418.
179. Xiao, Z., Wang, R., Wang, X., Yang, H., Dong, J., He, X., Yang, Y., Guo, J., Cui, J., and Zhou, Z. (2023). Impaired Function of Dendritic Cells within the Tumor Microenvironment, *Frontiers in Immunology*, Vol. 14. doi:10.3389/fimmu.2023.1213629.
180. Lin, K. X., Istl, A. C., Quan, D., Skaro, A., Tang, E., and Zheng, X. (2023). PD-1 and PD-L1 Inhibitors in Cold Colorectal Cancer: Challenges and Strategies, *Cancer Immunology, Immunotherapy*, Vol. 72, No. 12, 3875–3893. doi:10.1007/s00262-023-03520-5.
181. Seliger, B. (2019). Combinatorial Approaches with Checkpoint Inhibitors to Enhance Anti-Tumor Immunity, *Frontiers in Immunology*, Vol. 10. doi:10.3389/fimmu.2019.00999.
182. Subtirelu, R. C., Teichner, E. M., Ashok, A., Parikh, C., Talasila, S., Matache, I.-M., Alnemri, A. G., Anderson, V., Shahid, O., Mannam, S., Lee, A., Werner, T., Revheim, M.-E., and Alavi, A. (2023). Advancements in Dendritic Cell Vaccination: Enhancing Efficacy and Optimizing Combinatorial Strategies for the Treatment of Glioblastoma, *Frontiers in Neurology*, Vol. 14. doi:10.3389/fneur.2023.1271822.
183. van de Laar, L., Coffey, P. J., and Woltman, A. M. (2012). Regulation of Dendritic Cell Development by GM-CSF: Molecular

- Control and Implications for Immune Homeostasis and Therapy, *Blood*, Vol. 119, No. 15, 3383–3393. doi:10.1182/blood-2011-11-370130.
184. Bhattacharya, P., Budnick, I., Singh, M., Thirupathi, M., Alharshaw, K., Elshabrawy, H., Holterman, M. J., and Prabhakar, B. S. (2015). Dual Role of GM-CSF as a pro-Inflammatory and a Regulatory Cytokine: Implications for Immune Therapy, *Journal of Interferon & Cytokine Research*, Vol. 35, No. 8, 585–599. doi:10.1089/jir.2014.0149.
185. Qin, D., Zhang, Y., Shu, P., Lei, Y., Li, X., and Wang, Y. (2024). Targeting Tumor-Infiltrating Tregs for Improved Antitumor Responses, *Frontiers in Immunology*, Vol. 15. doi:10.3389/fimmu.2024.1325946.
186. Singer, M., Zhang, Z., Dayyani, F., Zhang, Z., Yaghmai, V., Choi, A., Valerin, J., Imagawa, D., and Abi-Jaoudeh, N. (2024). Modulation of Tumor-Associated Macrophages to Overcome Immune Suppression in the Hepatocellular Carcinoma Microenvironment, *Cancers*, Vol. 17, No. 1, 66. doi:10.3390/cancers17010066.
187. den Brok, M. H. M. G. M., Suttmuller, R. P. M., Nierkens, S., Bennink, E. J., Frielink, C., Toonen, L. W. J., Boerman, O. C., Figdor, C. G., Ruers, T. J. M., and Adema, G. J. (2006). Efficient Loading of Dendritic Cells Following Cryo and Radiofrequency Ablation in Combination with Immune Modulation Induces Anti-Tumour Immunity, *British Journal of Cancer*, Vol. 95, No. 7, 896–905. doi:10.1038/sj.bjc.6603341.
188. Ellis, S., Lin, E. J., and Tartar, D. (2018). Immunology of Wound Healing, *Current Dermatology Reports*, Vol. 7, No. 4, 350–358. doi:10.1007/s13671-018-0234-9.
189. Mazaki, J., Katsumata, K., Kasahara, K., Tago, T., Wada, T., Kuwabara, H., Enomoto, M., Ishizaki, T., Nagakawa, Y., and Tsuchida, A. (2020). Neutrophil-to-Lymphocyte Ratio Is a Prognostic Factor for Colon Cancer: A Propensity Score Analysis, *BMC Cancer*, Vol. 20, No. 1, 922. doi:10.1186/s12885-020-07429-5.
190. Triner, D., Devenport, S. N., Ramakrishnan, S. K., Ma, X., Frieler, R. A., Greenson, J. K., Inohara, N., Nunez, G., Colacino, J. A., Mortensen, R. M., and Shah, Y. M. (2019). Neutrophils Restrict Tumor-Associated Microbiota to Reduce Growth and Invasion of Colon Tumors in Mice, *Gastroenterology*, Vol. 156, No. 5, 1467–1482. doi:10.1053/j.gastro.2018.12.003.
191. Yamamoto, M., Kikuchi, H., Ohta, M., Kawabata, T., Hiramatsu, Y., Kondo, K., Baba, M., Kamiya, K., Tanaka, T., Kitagawa, M., and Konno, H. (2008). TSU68 Prevents Liver Metastasis of Colon Cancer Xenografts by Modulating the Premetastatic Niche, *Cancer Research*, Vol. 68, No. 23, 9754–9762. doi:10.1158/0008-5472.CAN-08-1748.
192. Hirai, H., Fujishita, T., Kurimoto, K., Miyachi, H., Kitano, S., Inamoto, S., Itatani, Y., Saitou, M., Maekawa, T., and Taketo, M. M. (2014). CCR1-Mediated Accumulation of Myeloid Cells in the Liver Microenvironment Promoting Mouse Colon Cancer Metastasis, *Clinical & Experimental Metastasis*, Vol. 31, No. 8, 977–989. doi:10.1007/s10585-014-9684-z.
193. Battle, E., and Massagué, J. (2019). Transforming Growth Factor- β Signaling in Immunity and Cancer, *Immunity*, Vol. 50, No. 4, 924–940. doi:10.1016/j.immuni.2019.03.024.
194. Lindau, D., Gielen, P., Kroesen, M., Wesseling, P., and Adema, G. J. (2013). The Immunosuppressive Tumour Network: Myeloid-derived Suppressor Cells, Regulatory T Cells and Natural Killer T Cells, *Immunology*, Vol. 138, No. 2, 105–115. doi:10.1111/imm.12036.
195. Fridlender, Z. G., Sun, J., Kim, S., Kapoor, V., Cheng, G., Ling, L., Worthen, G. S., and Albelda, S. M. (2009). Polarization of Tumor-Associated Neutrophil Phenotype by TGF- β : “N1” versus “N2” TAN, *Cancer Cell*, Vol. 16, No. 3, 183–194. doi:10.1016/j.ccr.2009.06.017.
196. Uribe-Querol, E., and Rosales, C. (2015). Neutrophils in Cancer: Two Sides of the Same Coin, *Journal of Immunology Research*, Vol. 2015, 1–21. doi:10.1155/2015/983698.
197. Qin, F., Liu, X., Chen, J., Huang, S., Wei, W., Zou, Y., Liu, X., Deng, K., Mo, S., Chen, J., Chen, X., Huang, Y., and Liang, W. (2020). Anti-TGF- β Attenuates Tumor Growth via Polarization of Tumor Associated Neutrophils towards an Anti-Tumor Phenotype in Colorectal Cancer, *Journal of Cancer*, Vol. 11, No. 9, 2580–2592. doi:10.7150/jca.38179.
198. Ganguly, D., Chandra, R., Karalis, J., Teke, M., Aguilera, T., Maddipati, R., Wachsmann, M. B., Ghersi, D., Siravegna, G., Zeh, H. J., Brekken, R., Ting, D. T., and Ligorio, M. (2020). Cancer-Associated Fibroblasts: Versatile Players in the Tumor Microenvironment, *Cancers*, Vol. 12, No. 9, 2652. doi:10.3390/cancers12092652.
199. Garvey, C. M., Lau, R., Sanchez, A., Sun, R. X., Fong, E. J., Doche, M. E., Chen, O., Jusuf, A., Lenz, H.-J., Larson, B., and Mumenthaler, S. M. (2020). Anti-EGFR Therapy Induces EGF Secretion by Cancer-Associated Fibroblasts to Confer Colorectal Cancer Chemoresistance, *Cancers*, Vol. 12, No. 6, 1393. doi:10.3390/cancers12061393.
200. Bhome, R., Bullock, M. D., Al Saihati, H. A., Goh, R. W., Primrose, J. N., Sayan, A. E., and Mirnezami, A. H. (2015). A Top-down View of the Tumor Microenvironment: Structure, Cells and Signaling, *Frontiers in Cell and Developmental Biology*, Vol. 3. doi:10.3389/fcell.2015.00033.
201. Sahai, E., Astsaturov, I., Cukierman, E., DeNardo, D. G., Egeblad, M., Evans, R. M., Fearon, D., Greten, F. R., Hingorani, S. R., Hunter, T., Hynes, R. O., Jain, R. K., Janowitz, T., Jorgensen, C., Kimmelman, A. C., Kolonin, M. G., Maki, R. G., Powers, R. S., Puré, E., Ramirez, D. C., Scherz-Shouval, R., Sherman, M. H., Stewart, S., Tlsty, T. D., Tuveson, D. A., Watt, F. M., Weaver, V., Weeraratna, A. T., and Werb, Z. (2020). A Framework for Advancing Our Understanding of Cancer-Associated Fibroblasts, *Nature Reviews Cancer*, Vol. 20, No. 3, 174–186. doi:10.1038/s41568-019-0238-1.
202. Erdogan, B., Ao, M., White, L. M., Means, A. L., Brewer, B. M., Yang, L., Washington, M. K., Shi, C., Franco, O. E., Weaver, A. M., Hayward, S. W., Li, D., and Webb, D. J. (2017). Cancer-Associated Fibroblasts Promote Directional Cancer Cell Migration by Aligning Fibronectin, *Journal of Cell Biology*, Vol. 216, No. 11, 3799–3816. doi:10.1083/jcb.201704053.
203. Walker, C., Mojares, E., and Del Río Hernández, A. (2018). Role of Extracellular Matrix in Development and Cancer Progression, *International Journal of Molecular Sciences*, Vol. 19, No. 10, 3028. doi:10.3390/ijms19103028.
204. Bauer, J., Emon, M. A. B., Staudacher, J. J., Thomas, A. L., Zessner-Spitzenberg, J., Mancinelli, G., Krett, N., Saif, M. T., and Jung, B. (2020). Increased Stiffness of the Tumor Microenvironment in Colon Cancer Stimulates Cancer Associated Fibroblast-Mediated Prometastatic Activin A Signaling, *Scientific Reports*, Vol. 10, No. 1, 50. doi:10.1038/s41598-019-55687-6.
205. Herrera, M., Llorens, C., Rodríguez, M., Herrera, A., Ramos, R., Gil, B., Candia, A., Larriba, M. J., Garre, P., Earl, J., Rodríguez-Garrote, M., Caldés, T., Bonilla, F., Carrato, A., García-Barberán, V., and Peña, C. (2018). Differential Distribution and Enrichment of Non-Coding RNAs in Exosomes from Normal and Cancer-Associated Fibroblasts in Colorectal Cancer, *Molecular Cancer*, Vol. 17, No. 1, 114. doi:10.1186/s12943-018-0863-4.
206. Chan, T.-S., Shaked, Y., and Tsai, K. K. (2019). Targeting the Interplay between Cancer Fibroblasts, Mesenchymal Stem Cells, and Cancer Stem Cells in Desmoplastic Cancers, *Frontiers in Oncology*, Vol. 9. doi:10.3389/fonc.2019.00688.

207. Jung, Y., Kim, J. K., Shiozawa, Y., Wang, J., Mishra, A., Joseph, J., Berry, J. E., McGee, S., Lee, E., Sun, H., Wang, J., Jin, T., Zhang, H., Dai, J., Krebsbach, P. H., Keller, E. T., Pienta, K. J., and Taichman, R. S. (2013). Recruitment of Mesenchymal Stem Cells into Prostate Tumours Promotes Metastasis, *Nature Communications*, Vol. 4, No. 1, 1795. doi:10.1038/ncomms2766.
208. Mishra, P. J., Mishra, P. J., Humeniuk, R., Medina, D. J., Alexe, G., Mesirov, J. P., Ganesan, S., Glod, J. W., and Banerjee, D. (2008). Carcinoma-Associated Fibroblast-like Differentiation of Human Mesenchymal Stem Cells, *Cancer Research*, Vol. 68, No. 11, 4331–4339. doi:10.1158/0008-5472.CAN-08-0943.
209. Gaggioli, C., Hooper, S., Hidalgo-Carcedo, C., Grosse, R., Marshall, J. F., Harrington, K., and Sahai, E. (2007). Fibroblast-Led Collective Invasion of Carcinoma Cells with Differing Roles for RhoGTPases in Leading and Following Cells, *Nature Cell Biology*, Vol. 9, No. 12, 1392–1400. doi:10.1038/ncb1658.
210. Otomo, R., Otsubo, C., Matsushima-Hibiya, Y., Miyazaki, M., Tashiro, F., Ichikawa, H., Kohno, T., Ochiya, T., Yokota, J., Nakagama, H., Taya, Y., and Enari, M. (2014). TSPAN12 Is a Critical Factor for Cancer-Fibroblast Cell Contact-Mediated Cancer Invasion, *Proceedings of the National Academy of Sciences*, Vol. 111, No. 52, 18691–18696. doi:10.1073/pnas.1412062112.
211. Semba, S., Kodama, Y., Ohnuma, K., Mizuuchi, E., Masuda, R., Yashiro, M., Hirakawa, K., and Yokozaki, H. (2009). Direct Cancer-Stromal Interaction Increases Fibroblast Proliferation and Enhances Invasive Properties of Scirrhous-Type Gastric Carcinoma Cells, *British Journal of Cancer*, Vol. 101, No. 8, 1365–1373. doi:10.1038/sj.bjc.6605309.
212. Rynne-Vidal, A., Jiménez-Heffernan, J., Fernández-Chacón, C., López-Cabrera, M., and Sandoval, P. (2015). The Mesothelial Origin of Carcinoma Associated-Fibroblasts in Peritoneal Metastasis, *Cancers*, Vol. 7, No. 4, 1994–2011. doi:10.3390/cancers7040872.
213. Germann, M., Zangger, N., Sauvain, M., Sempoux, C., Bowler, A. D., Wirapati, P., Kandalaf, L. E., Delorenzi, M., Tejpar, S., Coukos, G., and Radtke, F. (2020). Neutrophils Suppress Tumor-infiltrating T Cells in Colon Cancer via Matrix Metalloproteinase-mediated Activation of TGF β , *EMBO Molecular Medicine*, Vol. 12, No. 1. doi:10.15252/emmm.201910681.
214. Wang, L., Yang, J., Huang, J., Wen, Z.-Q., Xu, N., Liu, X., Zhang, J.-H., and Li, W.-L. (2020). MiRNA Expression Profile in the N2 Phenotype Neutrophils of Colorectal Cancer and Screen of Putative Key MiRNAs, *Cancer Management and Research*, Vol. Volume 12, 5491–5503. doi:10.2147/CMAR.S251427.
215. Tauriello, D. V. F., Palomo-Ponce, S., Stork, D., Berenguer-Llargo, A., Badia-Ramentol, J., Iglesias, M., Sevillano, M., Ibiza, S., Cañellas, A., Hernando-Mombolona, X., Byrom, D., Matarin, J. A., Calon, A., Rivas, E. I., Nebreda, A. R., Riera, A., Attolini, C. S.-O., and Batlle, E. (2018). TGF β Drives Immune Evasion in Genetically Reconstituted Colon Cancer Metastasis, *Nature*, Vol. 554, No. 7693, 538–543. doi:10.1038/nature25492.
216. Zhang, R., Qi, F., Zhao, F., Li, G., Shao, S., Zhang, X., Yuan, L., and Feng, Y. (2019). Cancer-Associated Fibroblasts Enhance Tumor-Associated Macrophages Enrichment and Suppress NK Cells Function in Colorectal Cancer, *Cell Death & Disease*, Vol. 10, No. 4, 273. doi:10.1038/s41419-019-1435-2.
217. Schellerer, V. S., Langheinrich, M., Hohenberger, W., Croner, R. S., Merkel, S., Rau, T. T., Sturzl, M., and Naschberger, E. (2014). Tumor-Associated Fibroblasts Isolated from Colorectal Cancer Tissues Exhibit Increased ICAM-1 Expression and Affinity for Monocytes, *Oncology Reports*, Vol. 31, No. 1, 255–261. doi:10.3892/or.2013.2860.
218. Zhang, D., Bi, J., Liang, Q., Wang, S., Zhang, L., Han, F., Li, S., Qiu, B., Fan, X., Chen, W., Jiao, H., Ye, Y., and Ding, Y. (2020). VCAM1 Promotes Tumor Cell Invasion and Metastasis by Inducing EMT and Transendothelial Migration in Colorectal Cancer, *Frontiers in Oncology*, Vol. 10. doi:10.3389/fonc.2020.01066.
219. Nagasaki, T., Hara, M., Nakanishi, H., Takahashi, H., Sato, M., and Takeyama, H. (2014). Interleukin-6 Released by Colon Cancer-Associated Fibroblasts Is Critical for Tumor Angiogenesis: Anti-Interleukin-6 Receptor Antibody Suppressed Angiogenesis and Inhibited Tumor-Stroma Interaction, *British Journal of Cancer*, Vol. 110, No. 2, 469–478. doi:10.1038/bjc.2013.748.
220. Mai, Z., Lin, Y., Lin, P., Zhao, X., and Cui, L. (2024). Modulating Extracellular Matrix Stiffness: A Strategic Approach to Boost Cancer Immunotherapy, *Cell Death & Disease*, Vol. 15, No. 5, 307. doi:10.1038/s41419-024-06697-4.
221. Chen, W.-Z., Jiang, J.-X., Yu, X.-Y., Xia, W.-J., Yu, P.-X., Wang, K., Zhao, Z.-Y., and Chen, Z.-G. (2019). Endothelial Cells in Colorectal Cancer, *World Journal of Gastrointestinal Oncology*, Vol. 11, No. 11, 946–956. doi:10.4251/wjgo.v11.i11.946.
222. Quiroz-Reyes, A. G., Islas, J. F., Delgado-Gonzalez, P., Franco-Villarreal, H., and Garza-Treviño, E. N. (2021). Therapeutic Approaches for Metastases from Colorectal Cancer and Pancreatic Ductal Carcinoma, *Pharmaceutics*, Vol. 13, No. 1, 103. doi:10.3390/pharmaceutics13010103.
223. Hida, K., Maishi, N., Annan, D., and Hida, Y. (2018). Contribution of Tumor Endothelial Cells in Cancer Progression, *International Journal of Molecular Sciences*, Vol. 19, No. 5, 1272. doi:10.3390/ijms19051272.
224. Nishida, N., Yano, H., Nishida, T., Kamura, T., and Kojiro, M. (2006). Angiogenesis in Cancer, *Vascular Health and Risk Management*, Vol. 2, No. 3, 213–219. doi:10.2147/vhrm.2006.2.3.213.
225. Zetter, B. R. (1998). Angiogenesis and Tumor Metastasis, *Annual Review of Medicine*, Vol. 49, No. 1, 407–424. doi:10.1146/annurev.med.49.1.407.
226. Croix, B. St., Rago, C., Velculescu, V., Traverso, G., Romans, K. E., Montgomery, E., Lal, A., Riggins, G. J., Lengauer, C., Vogelstein, B., and Kinzler, K. W. (2000). Genes Expressed in Human Tumor Endothelium, *Science*, Vol. 289, No. 5482, 1197–1202. doi:10.1126/science.289.5482.1197.
227. Lu, J., Ye, X., Fan, F., Xia, L., Bhattacharya, R., Bellister, S., Tozzi, F., Sceusi, E., Zhou, Y., Tachibana, I., Maru, D. M., Hawke, D. H., Rak, J., Mani, S. A., Zweidler-McKay, P., and Ellis, L. M. (2013). Endothelial Cells Promote the Colorectal Cancer Stem Cell Phenotype through a Soluble Form of Jagged-1, *Cancer Cell*, Vol. 23, No. 2, 171–185. doi:10.1016/j.ccr.2012.12.021.
228. Nübel, T., Dippold, W., Kleinert, H., Kaina, B., and Fritz, G. (2004). Lovastatin Inhibits Rho-regulated Expression of E-selectin by TNF- α and Attenuates Tumor Cell Adhesion, *The FASEB Journal*, Vol. 18, No. 1, 140–142. doi:10.1096/fj.03-0261fje.
229. Motz, G. T., Santoro, S. P., Wang, L.-P., Garrabrant, T., Lastra, R. R., Hagemann, I. S., Lal, P., Feldman, M. D., Benencia, F., and Coukos, G. (2014). Tumor Endothelium FasL Establishes a Selective Immune Barrier Promoting Tolerance in Tumors, *Nature Medicine*, Vol. 20, No. 6, 607–615. doi:10.1038/nm.3541.
230. Wang, J., Uddin, Md. N., Akter, R., and Wu, Y. (2021). Contribution of Endothelial Cell-Derived Transcriptomes to the Colon Cancer Based on Bioinformatics Analysis, *Mathematical Biosciences and Engineering*, Vol. 18, No. 6, 7280–7300. doi:10.3934/mbe.2021360.
231. Hu, L., Hayashi, Y., Kidoya, H., and Takakura, N. (2021). Endothelial Cell-Derived Apelin Inhibits Tumor Growth by Altering Immune Cell Localization, *Scientific Reports*, Vol. 11, No. 1, 14047. doi:10.1038/s41598-021-93619-5.
232. Naschberger, E., Liebel, A., Schellerer, V. S., Schütz, M., Britzen-Laurent, N., Köbel, P., Schaal, U., Haep, L., Regensburger, D., Wittmann, T., Klein-Hitpass, L., Rau, T. T., Diel, B., Méniel, V. S.,

- Clarke, A. R., Merkel, S., Croner, R. S., Hohenberger, W., and Stürzl, M. (2016). Matricellular Protein SPARCL1 Regulates Tumor Microenvironment-Dependent Endothelial Cell Heterogeneity in Colorectal Carcinoma, *Journal of Clinical Investigation*, Vol. 126, No. 11, 4187–4204. doi:10.1172/JCI78260.
233. Allen, E., Jabouille, A., Rivera, L. B., Lodewijckx, I., Missiaen, R., Steri, V., Feyen, K., Tawney, J., Hanahan, D., Michael, I. P., and Bergers, G. (2017). Combined Antiangiogenic and Anti-PD-L1 Therapy Stimulates Tumor Immunity through HEV Formation, *Science Translational Medicine*, Vol. 9, No. 385. doi:10.1126/scitranslmed.aak9679.
234. Li, Z.-L., Wang, Z.-J., Wei, G.-H., Yong, Y., and Wang, X.-W. (2020). Changes in Extracellular Matrix in Different Stages of Colorectal Cancer and Their Effects on Proliferation of Cancer Cells, *World Journal of Gastrointestinal Oncology*, Vol. 12, No. 3, 267–275. doi:10.4251/wjgo.v12.i3.267.
235. Najafi, M., Farhood, B., and Mortezaee, K. (2019). Extracellular Matrix (ECM) Stiffness and Degradation as Cancer Drivers, *Journal of Cellular Biochemistry*, Vol. 120, No. 3, 2782–2790. doi:10.1002/jcb.27681.
236. Tan, F., Huang, Y., Pei, Q., Liu, H., Pei, H., and Zhu, H. (2019). Matrix Stiffness Mediates Stemness Characteristics via Activating the Yes-associated Protein in Colorectal Cancer Cells, *Journal of Cellular Biochemistry*, Vol. 120, No. 2, 2213–2225. doi:10.1002/jcb.27532.
237. Liu, C., Pei, H., and Tan, F. (2020). Matrix Stiffness and Colorectal Cancer, *Oncotargets and Therapy*, Vol. Volume 13, 2747–2755. doi:10.2147/OTT.S231010.
238. Wu, X., Cai, J., Zuo, Z., and Li, J. (2019). Collagen Facilitates the Colorectal Cancer Stemness and Metastasis through an Integrin/PI3K/AKT/Snail Signaling Pathway, *Biomedicine & Pharmacotherapy*, Vol. 114, 108708. doi:10.1016/j.biopha.2019.108708.
239. Yi, W., Xiao, E., Ding, R., Luo, P., and Yang, Y. (2016). High Expression of Fibronectin Is Associated with Poor Prognosis, Cell Proliferation and Malignancy via the NF-KB/P53-Apoptosis Signaling Pathway in Colorectal Cancer, *Oncology Reports*, Vol. 36, No. 6, 3145–3153. doi:10.3892/or.2016.5177.
240. Ye, Y., Zhang, R., and Feng, H. (2020). Fibronectin Promotes Tumor Cells Growth and Drugs Resistance through a CDC42-YAP-dependent Signaling Pathway in Colorectal Cancer, *Cell Biology International*, Vol. 44, No. 9, 1840–1849. doi:10.1002/cbin.11390.
241. Kuermanbayi, S., Yang, Y., Zhao, Y., Li, Y., Wang, L., Yang, J., Zhou, Y., Xu, F., and Li, F. (2022). In Situ Monitoring of Functional Activity of Extracellular Matrix Stiffness-Dependent Multidrug Resistance Protein 1 Using Scanning Electrochemical Microscopy, *Chemical Science*, Vol. 13, No. 35, 10349–10360. doi:10.1039/D2SC02708A.
242. Muñoz, N. M., Williams, M., Dixon, K., Dupuis, C., McWatters, A., Avritscher, R., Manrique, S. Z., McHugh, K., Murthy, R., Tam, A., Naing, A., Patel, S. P., Leach, D., Hartgerink, J. D., Young, S., Prakash, P., Hwu, P., and Sheth, R. A. (2021). Influence of Injection Technique, Drug Formulation and Tumor Microenvironment on Intratumoral Immunotherapy Delivery and Efficacy, *Journal for Immunotherapy of Cancer*, Vol. 9, No. 2, e001800. doi:10.1136/jitc-2020-001800.
243. Ikeda, K., Iyama, K., Ishikawa, N., Egami, H., Nakao, M., Sado, Y., Ninomiya, Y., and Baba, H. (2006). Loss of Expression of Type IV Collagen A5 and A6 Chains in Colorectal Cancer Associated with the Hypermethylation of Their Promoter Region, *The American Journal of Pathology*, Vol. 168, No. 3, 856–865. doi:10.2353/ajpath.2006.050384.
244. Sala, M., Ros, M., and Saltel, F. (2020). A Complex and Evolutive Character: Two Face Aspects of ECM in Tumor Progression, *Frontiers in Oncology*, Vol. 10. doi:10.3389/fonc.2020.01620.
245. Yang, B., Tang, F., Zhang, B., Zhao, Y., Feng, J., and Rao, Z. (2014). Matrix Metalloproteinase-9 Overexpression Is Closely Related to Poor Prognosis in Patients with Colon Cancer, *World Journal of Surgical Oncology*, Vol. 12, No. 1, 24. doi:10.1186/1477-7819-12-24.
246. Dong, W., Li, H., Zhang, Y., Yang, H., Guo, M., Li, L., and Liu, T. (2011). Matrix Metalloproteinase 2 Promotes Cell Growth and Invasion in Colorectal Cancer, *Acta Biochimica et Biophysica Sinica*, Vol. 43, No. 11, 840–848. doi:10.1093/abbs/gmr085.
247. Sis, B., Sağol, Ö., Küpelioglu, A., Sokmen, S., Terzi, C., Fuzun, M., Özer, E., and Bishop, P. (2004). Prognostic Significance of Matrix Metalloproteinase-2, Cathepsin D, and Tenascin-C Expression in Colorectal Carcinoma, *Pathology - Research and Practice*, Vol. 200, No. 5, 379–387. doi:10.1016/j.prp.2004.02.012.
248. Zeng, Z.-S., Cohen, A. M., and Guillem, J. G. (1999). Loss of Basement Membrane Type IV Collagen Is Associated with Increased Expression of Metalloproteinases 2 and 9 (MMP-2 and MMP-9) during Human Colorectal Tumorigenesis, *Carcinogenesis*, Vol. 20, No. 5, 749–755. doi:10.1093/carcin/20.5.749.
249. Sand, J. M., Larsen, L., Hogaboam, C., Martinez, F., Han, M., Røssel Larsen, M., Nawrocki, A., Zheng, Q., Asser Karsdal, M., and Leeming, D. J. (2013). MMP Mediated Degradation of Type IV Collagen Alpha 1 and Alpha 3 Chains Reflects Basement Membrane Remodeling in Experimental and Clinical Fibrosis – Validation of Two Novel Biomarker Assays, *PLoS ONE*, Vol. 8, No. 12, e84934. doi:10.1371/journal.pone.0084934.
250. Hu, Y., Lin, L., Chen, J., Maruyama, A., Tian, H., and Chen, X. (2020). Synergistic Tumor Immunological Strategy by Combining Tumor Nanovaccine with Gene-Mediated Extracellular Matrix Scavenger, *Biomaterials*, Vol. 252, 120114. doi:10.1016/j.biomaterials.2020.120114.
251. Yasunaga, M., Manabe, S., Tarin, D., and Matsumura, Y. (2011). Cancer-Stroma Targeting Therapy by Cytotoxic Immunoconjugate Bound to the Collagen 4 Network in the Tumor Tissue, *Bioconjugate Chemistry*, Vol. 22, No. 9, 1776–1783. doi:10.1021/bc200158j.
252. Huijbers, E. J. M., Ringvall, M., Femel, J., Kalamajski, S., Lukinius, A., Åbrink, M., Hellman, L., and Olsson, A. (2010). Vaccination against the Extra Domain-B of Fibronectin as a Novel Tumor Therapy, *The FASEB Journal*, Vol. 24, No. 11, 4535–4544. doi:10.1096/fj.10-163022.
253. Scott, A. M., Wiseman, G., Welt, S., Adjei, A., Lee, F.-T., Hopkins, W., Divgi, C. R., Hanson, L. H., Mitchell, P., Gansen, D. N., Larson, S. M., Ingle, J. N., Hoffman, E. W., Tanswell, P., Ritter, G., Cohen, L. S., Bette, P., Arvey, L., Amelsberg, A., Vlock, D., Rettig, W. J., and Old, L. J. (2003). A Phase I Dose-Escalation Study of Sibrotuzumab in Patients with Advanced or Metastatic Fibroblast Activation Protein-Positive Cancer., *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*, Vol. 9, No. 5, 1639–47.
254. Biragyn, A., Schiavo, R., Olkhanud, P., Sumitomo, K., King, A., McCain, M., Indig, F. E., Almanzar, G., and Baatar, D. (2007). Tumor-Associated Embryonic Antigen-Expressing Vaccines That Target CCR6 Elicit Potent CD8+ T Cell-Mediated Protective and Therapeutic Antitumor Immunity, *The Journal of Immunology*, Vol. 179, No. 2, 1381–1388. doi:10.4049/jimmunol.179.2.1381.
255. Nicolas-Boluda, A., Vaquero, J., Vimeux, L., Guilbert, T., Barrin, S., Kantari-Mimoun, C., Ponzo, M., Renault, G., Deptula, P., Pogoda, K., Bucki, R., Cascone, I., Courty, J., Fouassier, L., Gazeau, F., and Donnadieu, E. (2021). Tumor Stiffening Reversion through Collagen Crosslinking Inhibition Improves T

- Cell Migration and Anti-PD-1 Treatment, *ELife*, Vol. 10. doi:10.7554/eLife.58688.
256. Ye, Y., Kuang, X., Xie, Z., Liang, L., Zhang, Z., Zhang, Y., Ma, F., Gao, Q., Chang, R., Lee, H.-H., Zhao, S., Su, J., Li, H., Peng, J., Chen, H., Yin, M., Peng, C., Yang, N., Wang, J., Liu, J., Liu, H., Han, L., and Chen, X. (2020). Small-Molecule MMP2/MMP9 Inhibitor SB-3CT Modulates Tumor Immune Surveillance by Regulating PD-L1, *Genome Medicine*, Vol. 12, No. 1, 83. doi:10.1186/s13073-020-00780-z.
257. Kaviyarasan, V., Das, A., Deka, D., Saha, B., Banerjee, A., Sharma, N. R., Duttaroy, A. K., and Pathak, S. (2024). Advancements in Immunotherapy for Colorectal Cancer Treatment: A Comprehensive Review of Strategies, Challenges, and Future Prospective, *International Journal of Colorectal Disease*, Vol. 40, No. 1, 1. doi:10.1007/s00384-024-04790-w.
258. Westcott, P. M. K., Sacks, N. J., Schenkel, J. M., Ely, Z. A., Smith, O., Hauck, H., Jaeger, A. M., Zhang, D., Backlund, C. M., Beytagh, M. C., Patten, J. J., Elbashir, R., Eng, G., Irvine, D. J., Yilmaz, O. H., and Jacks, T. (2021). Low Neoantigen Expression and Poor T-Cell Priming Underlie Early Immune Escape in Colorectal Cancer, *Nature Cancer*, Vol. 2, No. 10, 1071–1085. doi:10.1038/s43018-021-00247-z.
259. Zhang, Y., Rajput, A., Jin, N., and Wang, J. (2020). Mechanisms of Immunosuppression in Colorectal Cancer, *Cancers*, Vol. 12, No. 12, 3850. doi:10.3390/cancers12123850.
260. Han, J., Khatwani, N., Searles, T. G., Turk, M. J., and Angeles, C. V. (2020). Memory CD8+ T Cell Responses to Cancer, *Seminars in Immunology*, Vol. 49, 101435. doi:10.1016/j.smim.2020.101435.
261. Mizukoshi, E., Nakagawa, H., Tamai, T., Kitahara, M., Fushimi, K., Nio, K., Terashima, T., Iida, N., Arai, K., Yamashita, T., Yamashita, T., Sakai, Y., Honda, M., and Kaneko, S. (2022). Peptide Vaccine-Treated, Long-Term Surviving Cancer Patients Harbor Self-Renewing Tumor-Specific CD8+ T Cells, *Nature Communications*, Vol. 13, No. 1, 3123. doi:10.1038/s41467-022-30861-z.
262. Finn, O. J. (2018). The Dawn of Vaccines for Cancer Prevention, *Nature Reviews Immunology*, Vol. 18, No. 3, 183–194. doi:10.1038/nri.2017.140.
263. Middelburg, J., Schaap, G., Sluijter, M., Lloyd, K., Ovcinnikovs, V., Schuurman, J., van der Burg, S. H., Kemper, K., and van Hall, T. (2025). Cancer Vaccines Compensate for the Insufficient Induction of Protective Tumor-Specific Immunity of CD3 Bispecific Antibody Therapy, *Journal for ImmunoTherapy of Cancer*, Vol. 13, No. 1, e010331. doi:10.1136/jitc-2024-010331.
264. Liu, J., Fu, M., Wang, M., Wan, D., Wei, Y., and Wei, X. (2022). Cancer Vaccines as Promising Immuno-Therapeutics: Platforms and Current Progress, *Journal of Hematology & Oncology*, Vol. 15, No. 1, 28. doi:10.1186/s13045-022-01247-x.
265. Rahma, O. E., Gammoh, E., Simon, R. M., and Khleif, S. N. (2014). Is the “3+3” Dose-Escalation Phase I Clinical Trial Design Suitable for Therapeutic Cancer Vaccine Development? A Recommendation for Alternative Design, *Clinical Cancer Research*, Vol. 20, No. 18, 4758–4767. doi:10.1158/1078-0432.CCR-13-2671.
266. Humphrey, J. H., and Perdue, S. S. (n.d.). Immune System, *Britannica*, from <https://www.britannica.com/science/immune-system>, accessed 11-8-2025.
267. In Brief: The Innate and Adaptive Immune Systems. (2006). *InformedHealth.Org*, Institute for Quality and Efficiency in Health Care (IQWiG), Cologne, Germany.
268. Panawala, L. (2017). *Difference between humoral and cell mediated immunity*.
269. Abbas, A., Lichtman, A., and Pillai, S. (2016). *Basic Immunology: Functions and Disorders of the Immune System* (5th ed.), Elsevier, St. Louis, Missouri.
270. Sherwood, L. (2016). *Human Physiology: From Cells to Systems* (9th ed.), Cengage Learning, Boston.
271. Neefjes, J., Jongstra, M. L. M., Paul, P., and Bakke, O. (2011). Towards a Systems Understanding of MHC Class I and MHC Class II Antigen Presentation, *Nature Reviews Immunology*, Vol. 11, No. 12, 823–836. doi:10.1038/nri3084.
272. Roche, P. A., and Furuta, K. (2015). The Ins and Outs of MHC Class II-Mediated Antigen Processing and Presentation, *Nature Reviews Immunology*, Vol. 15, No. 4, 203–216. doi:10.1038/nri3818.
273. Skwarczynski, M., and Toth, I. (2016). Peptide-Based Synthetic Vaccines, *Chemical Science*, Vol. 7, No. 2, 842–854. doi:10.1039/C5SC03892H.
274. Wooster, A. L., Girgis, L. H., Brazeale, H., Anderson, T. S., Wood, L. M., and Lowe, D. B. (2021). Dendritic Cell Vaccine Therapy for Colorectal Cancer, *Pharmacological Research*, Vol. 164, 105374. doi:10.1016/j.phrs.2020.105374.
275. Li, W.-H., and Li, Y.-M. (2020). Chemical Strategies to Boost Cancer Vaccines, *Chemical Reviews*, Vol. 120, No. 20, 11420–11478. doi:10.1021/acs.chemrev.9b00833.
276. Hu, D., and Irving, A. T. (2023). Massively-Multiplexed Epitope Mapping Techniques for Viral Antigen Discovery, *Frontiers in Immunology*, Vol. 14. doi:10.3389/fimmu.2023.1192385.
277. Sinigaglia, F., and Hammer, J. (1994). Defining Rules for the Peptide-MHC Class II Interaction, *Current Opinion in Immunology*, Vol. 6, No. 1, 52–56. doi:10.1016/0952-7915(94)90033-7.
278. Kotsias, F., Cebrian, I., and Alloati, A. (2019). Antigen Processing and Presentation, 69–121. doi:10.1016/bs.ircmb.2019.07.005.
279. Keshavarz-Fathi, M., and Rezaei, N. (2021). Cancer Immunoprevention: Current Status and Future Directions, *Archivum Immunologiae et Therapiae Experimentalis*, Vol. 69, No. 1, 3. doi:10.1007/s00005-021-00604-x.
280. Sahin, U., and Türeci, Ö. (2018). Personalized Vaccines for Cancer Immunotherapy, *Science*, Vol. 359, No. 6382, 1355–1360. doi:10.1126/science.aar7112.
281. Chen, D. S., and Mellman, I. (2017). Elements of Cancer Immunity and the Cancer-Immune Set Point, *Nature*, Vol. 541, No. 7637, 321–330. doi:10.1038/nature21349.
282. Lohmueller, J., and Finn, O. J. (2017). Current Modalities in Cancer Immunotherapy: Immunomodulatory Antibodies, CARs and Vaccines, *Pharmacology & Therapeutics*, Vol. 178, 31–47. doi:10.1016/j.pharmthera.2017.03.008.
283. Spellberg, B., and Edwards, J. E. (2001). Type 1/Type 2 Immunity in Infectious Diseases, *Clinical Infectious Diseases*, Vol. 32, No. 1, 76–102. doi:10.1086/317537.
284. Berger, A. (2000). Science Commentary: Th1 and Th2 Responses: What Are They?, *BMJ*, Vol. 321, No. 7258, 424–424. doi:10.1136/bmj.321.7258.424.
285. Mendonça Gorgulho, C., Krishnamurthy, A., Lanzi, A., Galon, J., Housseau, F., Kaneno, R., and Lotze, M. T. (2021). Gutting It out: Developing Effective Immunotherapies for Patients with Colorectal Cancer, *Journal of Immunotherapy*, Vol. 44, No. 2, 49–62. doi:10.1097/CJI.0000000000000357.
286. Jia, W., Zhang, T., Huang, H., Feng, H., Wang, S., Guo, Z., Luo, Z., Ji, X., Cheng, X., and Zhao, R. (2022). Colorectal Cancer Vaccines: The Current Scenario and Future Prospects, *Frontiers in Immunology*, Vol. 13. doi:10.3389/fimmu.2022.942235.
287. Roberts, E. W., Broz, M. L., Binnewies, M., Headley, M. B., Nelson, A. E., Wolf, D. M., Kaisho, T., Bogunovic, D., Bhardwaj, N., and Krummel, M. F. (2016). Critical Role for CD103+/CD141+ Dendritic Cells Bearing CCR7 for Tumor Antigen Trafficking and

- Priming of T Cell Immunity in Melanoma, *Cancer Cell*, Vol. 30, No. 2, 324–336. doi:10.1016/j.ccell.2016.06.003.
288. Kaech, S. M., Wherry, E. J., and Ahmed, R. (2002). Effector and Memory T-Cell Differentiation: Implications for Vaccine Development, *Nature Reviews Immunology*, Vol. 2, No. 4, 251–262. doi:10.1038/nri778.
289. Wculek, S. K., Cueto, F. J., Mujal, A. M., Melero, I., Krummel, M. F., and Sancho, D. (2020). Dendritic Cells in Cancer Immunology and Immunotherapy, *Nature Reviews Immunology*, Vol. 20, No. 1, 7–24. doi:10.1038/s41577-019-0210-z.
290. Mitchell, D. M., Ravkov, E. V., and Williams, M. A. (2010). Distinct Roles for IL-2 and IL-15 in the Differentiation and Survival of CD8+ Effector and Memory T Cells, *The Journal of Immunology*, Vol. 184, No. 12, 6719–6730. doi:10.4049/jimmunol.0904089.
291. Carrio, R., Bathe, O. F., and Malek, T. R. (2004). Initial Antigen Encounter Programs CD8+ T Cells Competent to Develop into Memory Cells That Are Activated in an Antigen-Free, IL-7- and IL-15-Rich Environment, *The Journal of Immunology*, Vol. 172, No. 12, 7315–7323. doi:10.4049/jimmunol.172.12.7315.
292. Zangemeister-Wittke, U., Kyewski, B., and Schirrmacher, V. (1989). Recruitment and Activation of Tumor-Specific Immune T Cells in Situ. CD8+ Cells Predominate the Secondary Response in Sponge Matrices and Exert Both Delayed-Type Hypersensitivity-like and Cytotoxic T Lymphocyte Activity., *Journal of Immunology (Baltimore, Md. : 1950)*, Vol. 143, No. 1, 379–85.
293. Tuttle, T. M., Inge, T. H., Lind, D. S., and Bear, H. D. (1992). Adoptive Transfer of Bryostatin 1-Activated T Cells Provides Long-Term Protection from Tumour Metastases, *Surgical Oncology*, Vol. 1, No. 4, 299–307. doi:10.1016/0960-7404(92)90091-X.
294. Crosby, E. J., Hobeika, A. C., Niedzwiecki, D., Rushing, C., Hsu, D., Berglund, P., Smith, J., Osada, T., Gwin III, W. R., Hartman, Z. C., Morse, M. A., and Lyster, H. K. (2020). Long-Term Survival of Patients with Stage III Colon Cancer Treated with VRP-CEA(6D), an Alphavirus Vector That Increases the CD8+ Effector Memory T Cell to Treg Ratio, *Journal for ImmunoTherapy of Cancer*, Vol. 8, No. 2, e001662. doi:10.1136/jitc-2020-001662.
295. Snook, A. E., Baybutt, T. R., Xiang, B., Abraham, T. S., Flickinger, J. C., Hyslop, T., Zhan, T., Kraft, W. K., Sato, T., and Waldman, S. A. (2019). Split Tolerance Permits Safe Ad5-GUCY2C-PADRE Vaccine-Induced T-Cell Responses in Colon Cancer Patients, *Journal for ImmunoTherapy of Cancer*, Vol. 7, No. 1, 104. doi:10.1186/s40425-019-0576-2.
296. Farber, D. L., Yudanin, N. A., and Restifo, N. P. (2014). Human Memory T Cells: Generation, Compartmentalization and Homeostasis, *Nature Reviews Immunology*, Vol. 14, No. 1, 24–35. doi:10.1038/nri3567.
297. Martin, M. D., and Badovinac, V. P. (2018). Defining Memory CD8 T Cell, *Frontiers in Immunology*, Vol. 9. doi:10.3389/fimmu.2018.02692.
298. Gattinoni, L., Speiser, D. E., Lichterfeld, M., and Bonini, C. (2017). T Memory Stem Cells in Health and Disease, *Nature Medicine*, Vol. 23, No. 1, 18–27. doi:10.1038/nm.4241.
299. Schenkel, J. M., and Masopust, D. (2014). Tissue-Resident Memory T Cells, *Immunity*, Vol. 41, No. 6, 886–897. doi:10.1016/j.immuni.2014.12.007.
300. Pedicord, V. A., Montalvo, W., Leiner, I. M., and Allison, J. P. (2011). Single Dose of Anti-CTLA-4 Enhances CD8 + T-Cell Memory Formation, Function, and Maintenance, *Proceedings of the National Academy of Sciences*, Vol. 108, No. 1, 266–271. doi:10.1073/pnas.1016791108.
301. Fonseca, R., Beura, L. K., Quarnstrom, C. F., Ghoneim, H. E., Fan, Y., Zebley, C. C., Scott, M. C., Fares-Frederickson, N. J., Wijeyesinghe, S., Thompson, E. A., Borges da Silva, H., Vezys, V., Youngblood, B., and Masopust, D. (2020). Developmental Plasticity Allows Outside-in Immune Responses by Resident Memory T Cells, *Nature Immunology*, Vol. 21, No. 4, 412–421. doi:10.1038/s41590-020-0607-7.
302. Xu, L., Ye, L., and Huang, Q. (2025). Tissue-resident Memory CD8+ T Cells: Differentiation, Phenotypic Heterogeneity, Biological Function, Disease, and Therapy, *MedComm*, Vol. 6, No. 3. doi:10.1002/mco2.70132.
303. Cornista, A. M., Giolito, M. V., Baker, K., Hazime, H., Dufait, I., Datta, J., Khumukcham, S. S., De Ridder, M., Roper, J., Abreu, M. T., Breckpot, K., and Van der Jeught, K. (2023). Colorectal Cancer Immunotherapy: State of the Art and Future Directions, *Gastro Hep Advances*, Vol. 2, No. 8, 1103–1119. doi:10.1016/j.gastha.2023.09.007.
304. Binnewies, M., Roberts, E. W., Kersten, K., Chan, V., Fearon, D. F., Merad, M., Coussens, L. M., Gabrilovich, D. I., Ostrand-Rosenberg, S., Hedrick, C. C., Vonderheide, R. H., Pittet, M. J., Jain, R. K., Zou, W., Howcroft, T. K., Woodhouse, E. C., Weinberg, R. A., and Krummel, M. F. (2018). Understanding the Tumor Immune Microenvironment (TIME) for Effective Therapy, *Nature Medicine*, Vol. 24, No. 5, 541–550. doi:10.1038/s41591-018-0014-x.
305. Yu, H., Yang, R., Li, M., Li, D., and Xu, Y. (2025). The Role of Treg Cells in Colorectal Cancer and the Immunotherapy Targeting Treg Cells, *Frontiers in Immunology*, Vol. 16. doi:10.3389/fimmu.2025.1574327.
306. Pitt, J. M., Marabelle, A., Eggermont, A., Soria, J.-C., Kroemer, G., and Zitvogel, L. (2016). Targeting the Tumor Microenvironment: Removing Obstruction to Anticancer Immune Responses and Immunotherapy, *Annals of Oncology*, Vol. 27, No. 8, 1482–1492. doi:10.1093/annonc/mdw168.
307. Mishra, J., Drummond, J., Quazi, S. H., Karanki, S. S., Shaw, J. J., Chen, B., and Kumar, N. (2013). Prospective of Colon Cancer Treatments and Scope for Combinatorial Approach to Enhanced Cancer Cell Apoptosis, *Critical Reviews in Oncology/Hematology*, Vol. 86, No. 3, 232–250. doi:10.1016/j.critrevonc.2012.09.014.
308. Collins, J. M., Redman, J. M., and Gulley, J. L. (2018). Combining Vaccines and Immune Checkpoint Inhibitors to Prime, Expand, and Facilitate Effective Tumor Immunotherapy, *Expert Review of Vaccines*, Vol. 17, No. 8, 697–705. doi:10.1080/14760584.2018.1506332.
309. Melero, I., Berman, D. M., Aznar, M. A., Korman, A. J., Gracia, J. L. P., and Haanen, J. (2015). Evolving Synergistic Combinations of Targeted Immunotherapies to Combat Cancer, *Nature Reviews Cancer*, Vol. 15, No. 8, 457–472. doi:10.1038/nrc3973.
310. Cassidy, S., and Syed, B. A. (2017). Colorectal Cancer Drugs Market, *Nature Reviews Drug Discovery*, Vol. 16, No. 8, 525–526. doi:10.1038/nrd.2017.59.
311. Shin, M. H., Kim, J., Lim, S. A., Kim, J., and Lee, K.-M. (2020). Current Insights into Combination Therapies with MAPK Inhibitors and Immune Checkpoint Blockade, *International Journal of Molecular Sciences*, Vol. 21, No. 7, 2531. doi:10.3390/ijms21072531.
312. Goronzy, J. J., and Weyand, C. M. (2013). Understanding Immunosenescence to Improve Responses to Vaccines, *Nature Immunology*, Vol. 14, No. 5, 428–436. doi:10.1038/ni.2588.
313. Linton, P. J., and Dorshkind, K. (2004). Age-Related Changes in Lymphocyte Development and Function, *Nature Immunology*, Vol. 5, No. 2, 133–139. doi:10.1038/ni1033.
314. Blackman, M. A., and Woodland, D. L. (2011). The Narrowing of the CD8 T Cell Repertoire in Old Age, *Current Opinion in*

- Immunology*, Vol. 23, No. 4, 537–542. doi:10.1016/j.coi.2011.05.005.
315. Denis, F., Mounier, M., Hessel, L., Michel, J. P., Gualde, N., Dubois, F., Barin, F., and Goudeau, A. (1984). Hepatitis-B Vaccination in the Elderly, *Journal of Infectious Diseases*, Vol. 149, No. 6, 1019–1019. doi:10.1093/infdis/149.6.1019.
316. Pang, W. W., Price, E. A., Sahoo, D., Beerman, I., Maloney, W. J., Rossi, D. J., Schrier, S. L., and Weissman, I. L. (2011). Human Bone Marrow Hematopoietic Stem Cells Are Increased in Frequency and Myeloid-Biased with Age, *Proceedings of the National Academy of Sciences*, Vol. 108, No. 50, 20012–20017. doi:10.1073/pnas.1116110108.
317. Beerman, I., Bhattacharya, D., Zandi, S., Sigvardsson, M., Weissman, I. L., Bryder, D., and Rossi, D. J. (2010). Functionally Distinct Hematopoietic Stem Cells Modulate Hematopoietic Lineage Potential during Aging by a Mechanism of Clonal Expansion, *Proceedings of the National Academy of Sciences*, Vol. 107, No. 12, 5465–5470. doi:10.1073/pnas.1000834107.
318. Wang, J., Sun, Q., Morita, Y., Jiang, H., Groß, A., Lechel, A., Hildner, K., Guachalla, L. M., Gompf, A., Hartmann, D., Schambach, A., Wuestefeld, T., Dauch, D., Schrezenmeier, H., Hofmann, W.-K., Nakauchi, H., Ju, Z., Kestler, H. A., Zender, L., and Rudolph, K. L. (2012). A Differentiation Checkpoint Limits Hematopoietic Stem Cell Self-Renewal in Response to DNA Damage, *Cell*, Vol. 148, No. 5, 1001–1014. doi:10.1016/j.cell.2012.01.040.
319. Mandal, P. K., and Rossi, D. J. (2012). DNA-Damage-Induced Differentiation in Hematopoietic Stem Cells, *Cell*, Vol. 148, No. 5, 847–848. doi:10.1016/j.cell.2012.02.011.
320. Colmegna, I., Diaz-Borjon, A., Fujii, H., Schaefer, L., Goronzy, J. J., and Weyand, C. M. (2008). Defective Proliferative Capacity and Accelerated Telomeric Loss of Hematopoietic Progenitor Cells in Rheumatoid Arthritis, *Arthritis & Rheumatism*, Vol. 58, No. 4, 990–1000. doi:10.1002/art.23287.
321. Ademokun, A., Wu, Y.-C., and Dunn-Walters, D. (2010). The Ageing B Cell Population: Composition and Function, *Biogerontology*, Vol. 11, No. 2, 125–137. doi:10.1007/s10522-009-9256-9.
322. Haynes, L., Linton, P.-J., Eaton, S. M., Tonkonogy, S. L., and Swain, S. L. (1999). Interleukin 2, but Not Other Common γ Chain-Binding Cytokines, Can Reverse the Defect in Generation of Cd4 Effector T Cells from Naive T Cells of Aged Mice, *The Journal of Experimental Medicine*, Vol. 190, No. 7, 1013–1024. doi:10.1084/jem.190.7.1013.
323. Coffman, R. L., Sher, A., and Seder, R. A. (2010). Vaccine Adjuvants: Putting Innate Immunity to Work, *Immunity*, Vol. 33, No. 4, 492–503. doi:10.1016/j.immuni.2010.10.002.
324. Goronzy, J. J., Li, G., Yu, M., and Weyand, C. M. (2012). Signaling Pathways in Aged T Cells – a Reflection of T Cell Differentiation, Cell Senescence and Host Environment, *Seminars in Immunology*, Vol. 24, No. 5, 365–372. doi:10.1016/j.smim.2012.04.003.
325. Nikolich-Zugich, J., Li, G., Uhrlaub, J. L., Renkema, K. R., and Smithey, M. J. (2012). Age-Related Changes in CD8 T Cell Homeostasis and Immunity to Infection, *Seminars in Immunology*, Vol. 24, No. 5, 356–364. doi:10.1016/j.smim.2012.04.009.
326. Corulli, L. R., Cecil, D. L., Gad, E., Koehnlein, M., Coveler, A. L., Childs, J. S., Lubet, R. A., and Disis, M. L. (2021). Multi-Epitope-Based Vaccines for Colon Cancer Treatment and Prevention, *Frontiers in Immunology*, Vol. 12. doi:10.3389/fimmu.2021.729809.
327. Goodell, V., Waisman, J., Salazar, L. G., dela Rosa, C., Link, J., Coveler, A. L., Childs, J. S., Fintak, P. A., Higgins, D. M., and Disis, M. L. (2008). Level of HER-2/Neu Protein Expression in Breast Cancer May Affect the Development of Endogenous HER-2/Neu-Specific Immunity, *Molecular Cancer Therapeutics*, Vol. 7, No. 3, 449–454. doi:10.1158/1535-7163.MCT-07-0386.
328. Mineev, B., and Salgaller, M. L. (2004). Vaccines for the Immunotherapy of Prostate Cancer, *Handbook of Cancer Vaccines*, Humana Press, Totowa, NJ, 451–464. doi:10.1007/978-1-59259-680-5_29.
329. Kankanala, V., Zubair, M., and Mukkamla, S. (n.d.). *Carcinoembryonic Antigen*, StatPearls Publishing, Treasure Island (FL).
330. Fletcher, R. H. (1986). Carcinoembryonic Antigen, *Annals of Internal Medicine*, Vol. 104, No. 1, 66–73. doi:10.7326/0003-4819-104-1-66.
331. Gold, P., and Freedman, S. O. (1965). Demonstration of Tumor-Specific Antigens in Human Colonic Carcinomata by Immunologic Tolerance and Absorption Techniques, *The Journal of Experimental Medicine*, Vol. 121, No. 3, 439–462. doi:10.1084/jem.121.3.439.
332. Ohlsson, L., Israelsson, A., Öberg, Å., Palmqvist, R., Stenlund, H., Hammarström, M., Hammarström, S., and Lindmark, G. (2012). Lymph Node CEA and MUC2 mRNA as Useful Predictors of Outcome in Colorectal Cancer, *International Journal of Cancer*, Vol. 130, No. 8, 1833–1843. doi:10.1002/ijc.26182.
333. Wahren, B., and Harmenberg, U. (1991). Tumour Markers in Gastrointestinal Cancer, *Scandinavian Journal of Clinical and Laboratory Investigation*, Vol. 51, No. sup206, 21–27. doi:10.3109/0036519109107722.
334. Hammarström, S., and Baranov, V. (2001). Is There a Role for CEA in Innate Immunity in the Colon?, *Trends in Microbiology*, Vol. 9, No. 3, 119–125. doi:10.1016/S0966-842X(01)01952-7.
335. Taheri, M., Saragovi, U., Fuks, A., Makkerh, J., Mort, J., and Stanners, C. P. (2000). Self Recognition in the Ig Superfamily: Identification of Precise Subdomains in Carcinoembryonic Antigen Required for Intercellular Adhesion, *Journal of Biological Chemistry*, Vol. 275, No. 35, 26935–26943. doi:10.1016/S0021-9258(19)61463-8.
336. Yu, Z., Feng, J., Zhu, Y., Xie, X., Huang, H., Li, Y., Lu, Q., Jiang, J., and Wang, H. (2021). Clinicopathological and Prognostic Significance of TM4SF5 in Colorectal Cancer, *Research Square*. doi:10.21203/rs.3.rs-153931/v1.
337. Haeuw, J.-F., Goetsch, L., Bailly, C., and Corvaia, N. (2011). Tetraspanin CD151 as a Target for Antibody-Based Cancer Immunotherapy, *Biochemical Society Transactions*, Vol. 39, No. 2, 553–558. doi:10.1042/BST0390553.
338. Nair, S. K., Hull, S., Coleman, D., Gilboa, E., Lysterly, H. K., and Morse, M. A. (1999). Induction of Carcinoembryonic Antigen (CEA)-Specific Cytotoxic t-Lymphocyte Responses In Vitro Using Autologous Dendritic Cells Loaded with CEA Peptide or CEA RNA in Patients with Metastatic Malignancies Expressing CEA, *International Journal of Cancer*, Vol. 82, No. 1, 121–124. doi:10.1002/(SICI)1097-0215(19990702)82:1<121::AID-IJC20>3.0.CO;2-X.
339. Zaremba, S., Barzaga, E., Zhu, M., Soares, N., Tsang, K. Y., and Schlom, J. (1997). Identification of an Enhancer Agonist Cytotoxic T Lymphocyte Peptide from Human Carcinoembryonic Antigen, *Cancer Research*, Vol. 57, No. 20, 4570–7.
340. Park, J.-S., Kim, H.-S., Park, H.-M., Kim, C.-H., and Kim, T.-G. (2011). Efficient Induction of Anti-Tumor Immunity by a TAT-CEA Fusion Protein Vaccine with Poly(I:C) in a Murine Colorectal Tumor Model, *Vaccine*, Vol. 29, No. 47, 8642–8648. doi:10.1016/j.vaccine.2011.09.052.
341. Salucci, V., Mennuni, C., Calvaruso, F., Cerino, R., Neuner, P., Ciliberto, G., La Monica, N., and Scarselli, E. (2006). CD8+ T-cell

- Tolerance Can Be Broken by an Adenoviral Vaccine While CD4+ T-cell Tolerance Is Broken by Additional Co-administration of a Toll-like Receptor Ligand, *Scandinavian Journal of Immunology*, Vol. 63, No. 1, 35–41. doi:10.1111/j.1365-3083.2006.01706.x.
342. Gemei, M., Mirabelli, P., Di Noto, R., Corbo, C., Iaccarino, A., Zamboli, A., Troncone, G., Galizia, G., Lieto, E., Del Vecchio, L., and Salvatore, F. (2013). CD66c Is a Novel Marker for Colorectal Cancer Stem Cell Isolation, and Its Silencing Halts Tumor Growth in Vivo, *Cancer*, Vol. 119, No. 4, 729–738. doi:10.1002/cncr.27794.
343. Schölzel, S., Zimmermann, W., Schwarzkopf, G., Grunert, F., Rogaczewski, B., and Thompson, J. (2000). Carcinoembryonic Antigen Family Members CEACAM6 and CEACAM7 Are Differentially Expressed in Normal Tissues and Oppositely Deregulated in Hyperplastic Colorectal Polyps and Early Adenomas, *The American Journal of Pathology*, Vol. 156, No. 2, 595–605. doi:10.1016/S0002-9440(10)64764-5.
344. Jantschkeff, P., Terracciano, L., Lowy, A., Glatz-Krieger, K., Grunert, F., Micheel, B., Brümmer, J., Laffer, U., Metzger, U., Herrmann, R., and Rochlitz, C. (2003). Expression of CEACAM6 in Resectable Colorectal Cancer: A Factor of Independent Prognostic Significance, *Journal of Clinical Oncology*, Vol. 21, No. 19, 3638–3646. doi:10.1200/JCO.2003.55.135.
345. Blumenthal, R. D., Leon, E., Hansen, H. J., and Goldenberg, D. M. (2007). Expression Patterns of CEACAM5 and CEACAM6 in Primary and Metastatic Cancers, *BMC Cancer*, Vol. 7, No. 1, 2. doi:10.1186/1471-2407-7-2.
346. Kim, K. S., Kim, J.-T., Lee, S.-J., Kang, M. A., Choe, I. S., Kang, Y. H., Kim, S.-Y., Yeom, Y. Il, Lee, Y.-H., Kim, J. H., Kim, K. H., Kim, C. N., Kim, J. W., Nam, M.-S., and Lee, H. G. (2013). Overexpression and Clinical Significance of Carcinoembryonic Antigen-Related Cell Adhesion Molecule 6 in Colorectal Cancer, *Clinica Chimica Acta*, Vol. 415, 12–19. doi:10.1016/j.cca.2012.09.003.
347. Chen, J., Li, Q., An, Y., Lv, N., Xue, X., Wei, J., Jiang, K., Wu, J., Gao, W., Qian, Z., Dai, C., Xu, Z., and Miao, Y. (2013). CEACAM6 Induces Epithelial-Mesenchymal Transition and Mediates Invasion and Metastasis in Pancreatic Cancer, *International Journal of Oncology*, Vol. 43, No. 3, 877–885. doi:10.3892/ijco.2013.2015.
348. Thomas, P., Forse, R. A., and Bajenova, O. (2011). Carcinoembryonic Antigen (CEA) and Its Receptor HnRNP M Are Mediators of Metastasis and the Inflammatory Response in the Liver, *Clinical & Experimental Metastasis*, Vol. 28, No. 8, 923–932. doi:10.1007/s10585-011-9419-3.
349. Apostolopoulos, V., Stojanovska, L., and Gargosky, S. E. (2015). MUC1 (CD227): A Multi-Tasked Molecule, *Cellular and Molecular Life Sciences*, Vol. 72, No. 23, 4475–4500. doi:10.1007/s00018-015-2014-z.
350. Vlad, A. M., Kettel, J. C., Alajez, N. M., Carlos, C. A., and Finn, O. J. (2004). MUC1 Immunobiology: From Discovery to Clinical Applications, 249–293. doi:10.1016/S0065-2776(04)82006-6.
351. Ajioka, Y., Watanabe, H., and Jass, J. R. (1997). MUC1 and MUC2 Mucins in Flat and Polypoid Colorectal Adenomas., *Journal of Clinical Pathology*, Vol. 50, No. 5, 417–421. doi:10.1136/jcp.50.5.417.
352. Turner, M. S., McKolanis, J. R., Ramanathan, R. K., Whitcomb, D. C., and Finn, O. J. (2003). Mucins in Gastrointestinal Cancers, 259–274. doi:10.1016/S0921-4410(03)21012-7.
353. Guo, M., You, C., and Dou, J. (2018). Role of Transmembrane Glycoprotein Mucin 1 (MUC1) in Various Types of Colorectal Cancer and Therapies: Current Research Status and Updates, *Biomedicine & Pharmacotherapy*, Vol. 107, 1318–1325. doi:10.1016/j.biopha.2018.08.109.
354. Agrawal, B., Krantz, M. J., Reddish, M. A., and Longenecker, B. M. (1998). Cancer-Associated MUC1 Mucin Inhibits Human T-Cell Proliferation, Which Is Reversible by IL-2, *Nature Medicine*, Vol. 4, No. 1, 43–49. doi:10.1038/nm0198-043.
355. Zhang, Y., Dong, X., Bai, L., Shang, X., and Zeng, Y. (2020). MUC1-induced Immunosuppression in Colon Cancer Can Be Reversed by Blocking the PD1/PDL1 Signaling Pathway, *Oncology Letters*, Vol. 20, No. 6, 1–1. doi:10.3892/ol.2020.12180.
356. Biemer-Hüttmann, A.-E., Walsh, M. D., McGuckin, M. A., Ajioka, Y., Watanabe, H., Leggett, B. A., and Jass, J. R. (1999). Immunohistochemical Staining Patterns of MUC1, MUC2, MUC4, and MUC5AC Mucins in Hyperplastic Polyps, Serrated Adenomas, and Traditional Adenomas of the Colorectum, *Journal of Histochemistry & Cytochemistry*, Vol. 47, No. 8, 1039–1048. doi:10.1177/002215549904700808.
357. Mukherjee, P., Pathangey, L. B., Bradley, J. B., Tinder, T. L., Basu, G. D., Akporiaye, E. T., and Gendler, S. J. (2007). MUC1-Specific Immune Therapy Generates a Strong Anti-Tumor Response in a MUC1-Tolerant Colon Cancer Model, *Vaccine*, Vol. 25, No. 9, 1607–1618. doi:10.1016/j.vaccine.2006.11.007.
358. Guo, M., You, C., Dong, W., Luo, B., Wu, Y., Chen, Y., Li, J., Pan, M., Li, M., Zhao, F., and Dou, J. (2020). The Surface Dominant Antigen MUC1 Is Required for Colorectal Cancer Stem Cell Vaccine to Exert Anti-Tumor Efficacy, *Biomedicine & Pharmacotherapy*, Vol. 132, 110804. doi:10.1016/j.biopha.2020.110804.
359. Guo, M., Luo, B., Pan, M., Li, M., Xu, H., Zhao, F., and Dou, J. (2020). Colorectal Cancer Stem Cell Vaccine with High Expression of MUC1 Serves as a Novel Prophylactic Vaccine for Colorectal Cancer, *International Immunopharmacology*, Vol. 88, 106850. doi:10.1016/j.intimp.2020.106850.
360. Kimura, T., McKolanis, J. R., Dzubinski, L. A., Islam, K., Potter, D. M., Salazar, A. M., Schoen, R. E., and Finn, O. J. (2013). MUC1 Vaccine for Individuals with Advanced Adenoma of the Colon: A Cancer Immunoprevention Feasibility Study, *Cancer Prevention Research*, Vol. 6, No. 1, 18–26. doi:10.1158/1940-6207.CAPR-12-0275.
361. Schoen, R. E., Boardman, L. A., Cruz-Correa, M., Bansal, A., Kastenberger, D., Hur, C., Dzubinski, L., Kaufman, S. F., Rodriguez, L. M., Richmond, E., Umar, A., Szabo, E., Salazar, A., McKolanis, J., Beatty, P., Pai, R. K., Singhi, A. D., Jacqueline, C. M., Bao, R., Diergaarde, B., McMurray, R. P., Strand, C., Foster, N. R., Zahrieh, D. M., Limburg, P. J., and Finn, O. J. (2023). Randomized, Double-Blind, Placebo-Controlled Trial of MUC1 Peptide Vaccine for Prevention of Recurrent Colorectal Adenoma, *Clinical Cancer Research*, Vol. 29, No. 9, 1678–1688. doi:10.1158/1078-0432.CCR-22-3168.
362. Arteaga, C. L. (2002). Overview of Epidermal Growth Factor Receptor Biology and Its Role as a Therapeutic Target in Human Neoplasia, *Seminars in Oncology*, Vol. 29, No. 5 Suppl 14, asonc02905o0003. doi:10.1053/sonc.2002.35642.
363. Boerner, J. (2003). Ligand-Independent Oncogenic Signaling by the Epidermal Growth Factor Receptor: V-ErbB as a Paradigm, *Experimental Cell Research*, Vol. 284, No. 1, 111–121. doi:10.1016/S0014-4827(02)00096-4.
364. Foy, K. C., Wygle, R. M., Miller, M. J., Overholser, J. P., Bekaii-Saab, T., and Kaumaya, P. T. P. (2013). Peptide Vaccines and Peptidomimetics of EGFR (HER-1) Ligand Binding Domain Inhibit Cancer Cell Growth In Vitro and In Vivo, *The Journal of Immunology*, Vol. 191, No. 1, 217–227. doi:10.4049/jimmunol.1300231.
365. Brabender, J., Danenberg, K. D., Metzger, R., Schneider, P. M., Park, J., Salonga, D., Hölscher, A. H., and Danenberg, P. V. (2001). Epidermal Growth Factor Receptor and HER2-Neu mRNA Expression in Non-Small Cell Lung Cancer Is Correlated with Survival., *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*, Vol. 7, No. 7, 1850–5.

366. de Almagro, M. C., and Vucic, D. (2012). The Inhibitor of Apoptosis (IAP) Proteins Are Critical Regulators of Signaling Pathways and Targets for Anti-Cancer Therapy., *Experimental Oncology*, Vol. 34, No. 3, 200–211.
367. Mehrotra, S., Languino, L. R., Raskett, C. M., Mercurio, A. M., Dohi, T., and Altieri, D. C. (2010). IAP Regulation of Metastasis, *Cancer Cell*, Vol. 17, No. 1, 53–64.
368. Ambrosini, G., Adida, C., and Altieri, D. C. (1997). A Novel Anti-Apoptosis Gene, Survivin, Expressed in Cancer and Lymphoma, *Nature Medicine*, Vol. 3, No. 8, 917–921. doi:10.1038/nm0897-917.
369. Velculescu, V. E., Madden, S. L., Zhang, L., Lash, A. E., Yu, J., Rago, C., Lal, A., Wang, C. J., Beaudry, G. A., Ciriello, K. M., Cook, B. P., Dufault, M. R., Ferguson, A. T., Gao, Y., He, T.-C., Hermeking, H., Hiraldo, S. K., Hwang, P. M., Lopez, M. A., Luderer, H. F., Mathews, B., Petroziello, J. M., Polyak, K., Zawel, L., Zhang, W., Zhang, X., Zhou, W., Haluska, F. G., Jen, J., Sukumar, S., Landes, G. M., Riggins, G. J., Vogelstein, B., and Kinzler, K. W. (1999). Analysis of Human Transcriptomes, *Nature Genetics*, Vol. 23, No. 4, 387–388. doi:10.1038/70487.
370. Sarela, A. I. (2000). Expression of the Antiapoptosis Gene, Survivin, Predicts Death from Recurrent Colorectal Carcinoma, *Gut*, Vol. 46, No. 5, 645–650. doi:10.1136/gut.46.5.645.
371. Krieg, A., Werner, T. A., Verde, P. E., Stoecklein, N. H., and Knoefel, W. T. (2013). Prognostic and Clinicopathological Significance of Survivin in Colorectal Cancer: A Meta-Analysis, *PLoS ONE*, Vol. 8, No. 6, e65338. doi:10.1371/journal.pone.0065338.
372. Idenoue, S., Hirohashi, Y., Torigoe, T., Sato, Y., Tamura, Y., Hariu, H., Yamamoto, M., Kurotaki, T., Tsuruma, T., Asanuma, H., Kanaseki, T., Ikeda, H., Kashiwagi, K., Okazaki, M., Sasaki, K., Sato, T., Ohmura, T., Hata, F., Yamaguchi, K., Hirata, K., and Sato, N. (2005). A Potent Immunogenic General Cancer Vaccine That Targets Survivin, an Inhibitor of Apoptosis Proteins, *Clinical Cancer Research*, Vol. 11, No. 4, 1474–1482. doi:10.1158/1078-0432.CCR-03-0817.
373. Tsuruma, T., Hata, F., Torigoe, T., Furuhashi, T., Idenoue, S., Kurotaki, T., Yamamoto, M., Yagihashi, A., Ohmura, T., Yamaguchi, K., Katsuramaki, T., Yasoshima, T., Sasaki, K., Mizushima, Y., Minamida, H., Kimura, H., Akiyama, M., Hirohashi, Y., Asanuma, H., Tamura, Y., Shimozaawa, K., Sato, N., and Hirata, K. (2004). Phase I Clinical Study of Anti-Apoptosis Protein, Survivin-Derived Peptide Vaccine Therapy for Patients with Advanced or Recurrent Colorectal Cancer, *Journal of Translational Medicine*, Vol. 2, No. 1, 19. doi:10.1186/1479-5876-2-19.
374. Nupur, F. P., Khatun, J., Pervin, S., Hossain, S., Wahid, S. R., Akter, J., Shirin, S., and Snigdha, S. S. (2025). Expression of WT1 in Colorectal Adenocarcinoma and Its Association with Histological Grade and Stage, *World Journal of Internal Medicine and Surgery*, Vol. 2, No. 2, 15–20.
375. Van Driessche, A., Berneman, Z. N., and Van Tendeloo, V. F. I. (2012). Active Specific Immunotherapy Targeting the Wilms' Tumor Protein 1 (WT1) for Patients with Hematological Malignancies and Solid Tumors: Lessons from Early Clinical Trials, *The Oncologist*, Vol. 17, No. 2, 250–259. doi:10.1634/theoncologist.2011-0240.
376. Shimodaira, S., Sano, K., Hirabayashi, K., Koya, T., Higuchi, Y., Mizuno, Y., Yamaoka, N., Yuzawa, M., Kobayashi, T., Ito, K., and Koizumi, T. (2015). Dendritic Cell-Based Adjuvant Vaccination Targeting Wilms' Tumor 1 in Patients with Advanced Colorectal Cancer, *Vaccines*, Vol. 3, No. 4, 1004–1018. doi:10.3390/vaccines3041004.
377. Lee, J. W. (2014). TM4SF5-Mediated Protein-Protein Networks and Tumorigenic Roles, *BMB Reports*, Vol. 47, No. 9, 483–487. doi:10.5483/BMBRep.2014.47.9.146.
378. Richardson, M. M., Jennings, L. K., and Zhang, X. A. (2011). Tetraspanins and Tumor Progression, *Clinical & Experimental Metastasis*, Vol. 28, No. 3, 261–270. doi:10.1007/s10585-010-9365-5.
379. Anderson, K. R., Singer, R. A., Balderes, D. A., Hernandez-Lagunas, L., Johnson, C. W., Artinger, K. B., and Sussel, L. (2011). The L6 Domain Tetraspanin Tm4sf4 Regulates Endocrine Pancreas Differentiation and Directed Cell Migration, *Development*, Vol. 138, No. 15, 3213–3224. doi:10.1242/dev.058693.
380. Lee, S.-A., Lee, S.-Y., Cho, I.-H., Oh, M.-A., Kang, E.-S., Kim, Y.-B., Seo, W. D., Choi, S., Nam, J.-O., Tamamori-Adachi, M., Kitajima, S., Ye, S.-K., Kim, S., Hwang, Y.-J., Kim, I.-S., Park, K. H., and Lee, J. W. (2008). Tetraspanin TM4SF5 Mediates Loss of Contact Inhibition through Epithelial-Mesenchymal Transition in Human Hepatocarcinoma, *Journal of Clinical Investigation*, Vol. 118, No. 4, 1354–1366. doi:10.1172/JCI33768.
381. Kwon, S., Kim, Y.-E., Park, J.-A., Kim, D.-S., Kwon, H.-J., and Lee, Y. (2014). Therapeutic Effect of a TM4SF5-Specific Peptide Vaccine against Colon Cancer in a Mouse Model, *BMB Reports*, Vol. 47, No. 4, 215–220. doi:10.5483/BMBRep.2014.47.4.157.
382. Berger, H., Breuer, M., Peradziry, H., Podleschny, M., Jacob, R., and Borchers, A. (2017). PTK7 Localization and Protein Stability Is Affected by Canonical Wnt Ligands, *Journal of Cell Science*, Vol. 130, No. 11, 1890–1903. doi:10.1242/jcs.198580.
383. Bin-Nun, N., Lichtig, H., Malyarova, A., Levy, M., Elias, S., and Frank, D. (2014). PTK7 Modulates Wnt Signaling Activity via LRP6, *Development*, Vol. 141, No. 2, 410–421. doi:10.1242/dev.095984.
384. Martinez, S., Scerbo, P., Giordano, M., Daulat, A. M., Lhoumeau, A.-C., Thomé, V., Kodjabachian, L., and Borg, J.-P. (2015). The PTK7 and ROR2 Protein Receptors Interact in the Vertebrate WNT/Planar Cell Polarity (PCP) Pathway, *Journal of Biological Chemistry*, Vol. 290, No. 51, 30562–30572. doi:10.1074/jbc.M115.697615.
385. Peradziry, H., Kaplan, N. A., Podleschny, M., Liu, X., Wehner, P., Borchers, A., and Tolwinski, N. S. (2011). PTK7/Otk Interacts with Wnts and Inhibits Canonical Wnt Signalling, *The EMBO Journal*, Vol. 30, No. 18, 3729–3740. doi:10.1038/emboj.2011.236.
386. Berger, H., Wodarz, A., and Borchers, A. (2017). PTK7 Faces the Wnt in Development and Disease, *Frontiers in Cell and Developmental Biology*, Vol. 5, 31. doi:10.3389/fcell.2017.00031.
387. Klaus, A., and Birchmeier, W. (2008). Wnt Signalling and Its Impact on Development and Cancer, *Nature Reviews Cancer*, Vol. 8, No. 5, 387–398. doi:10.1038/nrc2389.
388. Espada, J., Calvo, M. B., Diaz-Prado, S., and Medina, V. (2009). Wnt Signalling and Cancer Stem Cells, *Clinical and Translational Oncology*, Vol. 11, No. 7, 411–427. doi:10.1007/s12094-009-0380-4.
389. Patel, S. A., Nilsson, M. B., Le, X., Cascone, T., Jain, R. K., and Heymach, J. V. (2023). Molecular Mechanisms and Future Implications of VEGF/VEGFR in Cancer Therapy, *Clinical Cancer Research*, Vol. 29, No. 1, 30–39. doi:10.1158/1078-0432.CCR-22-1366.
390. Bu, M. T., Chandrasekhar, P., Ding, L., and Hugo, W. (2022). The Roles of TGF- β and VEGF Pathways in the Suppression of Antitumor Immunity in Melanoma and Other Solid Tumors, *Pharmacology & Therapeutics*, Vol. 240, 108211. doi:10.1016/j.pharmthera.2022.108211.
391. Yang, Y., and Cao, Y. (2022). The Impact of VEGF on Cancer Metastasis and Systemic Disease, *Seminars in Cancer Biology*, Vol. 86, 251–261. doi:10.1016/j.semcancer.2022.03.011.

392. Apte, R. S., Chen, D. S., and Ferrara, N. (2019). VEGF in Signaling and Disease: Beyond Discovery and Development, *Cell*, Vol. 176, No. 6, 1248–1264. doi:10.1016/j.cell.2019.01.021.
393. Cao, Y. (2014). VEGF-Targeted Cancer Therapeutics—Paradoxical Effects in Endocrine Organs, *Nature Reviews Endocrinology*, Vol. 10, No. 9, 530–539. doi:10.1038/nrendo.2014.114.
394. Chen, R., Khatri, P., Mazur, P. K., Polin, M., Zheng, Y., Vaka, D., Hoang, C. D., Shrager, J., Xu, Y., Vicent, S., Butte, A. J., and Sweet-Cordero, E. A. (2014). A Meta-Analysis of Lung Cancer Gene Expression Identifies PTK7 as a Survival Gene in Lung Adenocarcinoma, *Cancer Research*, Vol. 74, No. 10, 2892–2902. doi:10.1158/0008-5472.CAN-13-2775.
395. Ataseven, B., Angerer, R., Kates, R., Gunesch, A., Knyazev, P., Högel, B., Becker, C., Eiermann, W., and Harbeck, N. (2013). PTK7 Expression in Triple-Negative Breast Cancer., *Anticancer Research*, Vol. 33, No. 9, 3759–3763.
396. Gärtner, S., Gunesch, A., Knyazeva, T., Wolf, P., Högel, B., Eiermann, W., Ullrich, A., Knyazev, P., and Ataseven, B. (2014). PTK 7 Is a Transforming Gene and Prognostic Marker for Breast Cancer and Nodal Metastasis Involvement, *PLoS ONE*, Vol. 9, No. 1, e84472. doi:10.1371/journal.pone.0084472.
397. Shin, W.-S., Gim, J., Won, S., and Lee, S.-T. (2018). Biphasic Regulation of Tumorigenesis by PTK7 Expression Level in Esophageal Squamous Cell Carcinoma, *Scientific Reports*, Vol. 8, No. 1, 8519. doi:10.1038/s41598-018-26957-6.
398. Lhoumeau, A.-C., Martinez, S., Boher, J.-M., Monges, G., Castellano, R., Goubard, A., Doremus, M., Poizat, F., Lelong, B., de Chaisemartin, C., Bardin, F., Viens, P., Raoul, J.-L., Prebet, T., Aurrand-Lions, M., Borg, J.-P., and Gonçalves, A. (2015). Overexpression of the Promigratory and Prometastatic PTK7 Receptor Is Associated with an Adverse Clinical Outcome in Colorectal Cancer, *PLOS ONE*, Vol. 10, No. 5, e0123768. doi:10.1371/journal.pone.0123768.
399. Tian, X., Yan, L., Zhang, D., Guan, X., Dong, B., Zhao, M., and Hao, C. (2016). PTK7 Overexpression in Colorectal Tumors: Clinicopathological Correlation and Prognosis Relevance, *Oncology Reports*, Vol. 36, No. 4, 1829–1836. doi:10.3892/or.2016.4983.
400. Pirro, M., Rombouts, Y., Stella, A., Neyrolles, O., Bulet-Schiltz, O., van Vliet, S. J., de Ru, A. H., Mohammed, Y., Wührer, M., van Veelen, P. A., and Hensbergen, P. J. (2020). Characterization of Macrophage Galactose-Type Lectin (MGL) Ligands in Colorectal Cancer Cell Lines, *Biochimica et Biophysica Acta (BBA) - General Subjects*, Vol. 1864, No. 4, 129513. doi:10.1016/j.bbagen.2020.129513.
401. Kanchan, R. K., Doss, D., Khan, P., Nasser, Mohd. W., and Mahapatra, S. (2022). To Kill a Cancer: Targeting the Immune Inhibitory Checkpoint Molecule, B7-H3, *Biochimica et Biophysica Acta (BBA) - Reviews on Cancer*, Vol. 1877, No. 5, 188783. doi:10.1016/j.bbcan.2022.188783.
402. Wang, L., Kang, F., and Shan, B. (2014). B7-H3-mediated Tumor Immunology: Friend or Foe?, *International Journal of Cancer*, Vol. 134, No. 12, 2764–2771. doi:10.1002/ijc.28474.
403. Zang, X., Thompson, R. H., Al-Ahmadie, H. A., Serio, A. M., Reuter, V. E., Eastham, J. A., Scardino, P. T., Sharma, P., and Allison, J. P. (2007). B7-H3 and B7x Are Highly Expressed in Human Prostate Cancer and Associated with Disease Spread and Poor Outcome, *Proceedings of the National Academy of Sciences*, Vol. 104, No. 49, 19458–19463. doi:10.1073/pnas.0709802104.
404. Crispin, P. L., Sheinin, Y., Roth, T. J., Lohse, C. M., Kuntz, S. M., Frigola, X., Thompson, R. H., Boorjian, S. A., Dong, H., Leibovich, B. C., Blute, M. L., and Kwon, E. D. (2008). Tumor Cell and Tumor Vasculature Expression of B7-H3 Predict Survival in Clear Cell Renal Cell Carcinoma, *Clinical Cancer Research*, Vol. 14, No. 16, 5150–5157. doi:10.1158/1078-0432.CCR-08-0536.
405. Kanchan, R. K., Perumal, N., Atri, P., Chirravuri Venkata, R., Thapa, I., Klinkebiel, D. L., Donson, A. M., Perry, D., Punsoni, M., Talmon, G. A., Coulter, D. W., Boue, D. R., Snuderl, M., Nasser, Mohd. W., Batra, S. K., Vibhakar, R., and Mahapatra, S. (2020). MiR-1253 Exerts Tumor-suppressive Effects in Medulloblastoma via Inhibition of CDK6 and CD276 (B7-H3), *Brain Pathology*, Vol. 30, No. 4, 732–745. doi:10.1111/bpa.12829.
406. Sun, Y., Wang, Y., Zhao, J., Gu, M., Giscombe, R., Lefvert, A. K., and Wang, X. (2006). B7-H3 and B7-H4 Expression in Non-Small-Cell Lung Cancer, *Lung Cancer*, Vol. 53, No. 2, 143–151. doi:10.1016/j.lungcan.2006.05.012.
407. Yamato, I., Sho, M., Nomi, T., Akahori, T., Shimada, K., Hotta, K., Kanehiro, H., Konishi, N., Yagita, H., and Nakajima, Y. (2009). Clinical Importance of B7-H3 Expression in Human Pancreatic Cancer, *British Journal of Cancer*, Vol. 101, No. 10, 1709–1716. doi:10.1038/sj.bjc.6605375.
408. Wang, J., Chong, K. K., Nakamura, Y., Nguyen, L., Huang, S. K., Kuo, C., Zhang, W., Yu, H., Morton, D. L., and Hoon, D. S. B. (2013). B7-H3 Associated with Tumor Progression and Epigenetic Regulatory Activity in Cutaneous Melanoma, *Journal of Investigative Dermatology*, Vol. 133, No. 8, 2050–2058. doi:10.1038/jid.2013.114.
409. Zhou, Z., Luther, N., Ibrahim, G. M., Hawkins, C., Vibhakar, R., Handler, M. H., and Souweidane, M. M. (2013). B7-H3, a Potential Therapeutic Target, Is Expressed in Diffuse Intrinsic Pontine Glioma, *Journal of Neuro-Oncology*, Vol. 111, No. 3, 257–264. doi:10.1007/s11060-012-1021-2.
410. Flies, D. B., Han, X., Higuchi, T., Zheng, L., Sun, J., Ye, J. J., and Chen, L. (2014). Coinhibitory Receptor PD-1H Preferentially Suppresses CD4+ T Cell-Mediated Immunity, *Journal of Clinical Investigation*, Vol. 124, No. 5, 1966–1975. doi:10.1172/JCI74589.
411. Wang, G., Wu, Z., Wang, Y., Li, X., Zhang, G., and Hou, J. (2016). Therapy to Target Renal Cell Carcinoma Using 131I-Labeled B7-H3 Monoclonal Antibody, *Oncotarget*, Vol. 7, No. 17, 24888–24898. doi:10.18632/oncotarget.8550.
412. Fauci, J. M., Straughn, J. M., Ferrone, S., and Buchsbaum, D. J. (2012). A Review of B7-H3 and B7-H4 Immune Molecules and Their Role in Ovarian Cancer, *Gynecologic Oncology*, Vol. 127, No. 2, 420–425. doi:10.1016/j.ygyno.2012.08.017.
413. Brunner, A., Hinterholzer, S., Riss, P., Heinze, G., and Brustmann, H. (2012). Immunoexpression of B7-H3 in Endometrial Cancer: Relation to Tumor T-Cell Infiltration and Prognosis, *Gynecologic Oncology*, Vol. 124, No. 1, 105–111. doi:10.1016/j.ygyno.2011.09.012.
414. Liu, T., Huo, Y., Li, G., Yu, G., and Luan, X. (2014). A Negative Correlation between B7-H3 Expression and the Number of CD8+ T Cell Infiltration in Primary Hepatocellular Carcinoma Tissues, *Chinese Journal of Cellular and Molecular Immunology*, Vol. 30, No. 12, 1291–4.
415. Ingebrigtsen, V. A., Boye, K., Tekle, C., Nesland, J. M., Flatmark, K., and Fodstad, Ø. (2012). B7-H3 Expression in Colorectal Cancer: Nuclear Localization Strongly Predicts Poor Outcome in Colon Cancer, *International Journal of Cancer*, Vol. 131, No. 11, 2528–2536. doi:10.1002/ijc.27566.
416. Lee, H., Kim, J. H., Yang, S. Y., Kong, J., Oh, M., Jeong, D. H., Chung, J., Bae, K. B., Shin, J. Y., Hong, K. H., and Choi, I. (2010). Peripheral Blood Gene Expression of B7 and CD28 Family Members Associated with Tumor Progression and Microscopic Lymphovascular Invasion in Colon Cancer Patients, *Journal of Cancer Research and Clinical Oncology*, Vol. 136, No. 9, 1445–1452. doi:10.1007/s00432-010-0800-4.

417. Leitner, J., Klauser, C., Pickl, W. F., Stöckl, J., Majdic, O., Bardet, A. F., Kreil, D. P., Dong, C., Yamazaki, T., Zlabinger, G., Pfistershammer, K., and Steinberger, P. (2009). B7-H3 Is a Potent Inhibitor of Human T-cell Activation: No Evidence for B7-H3 and TREM2 Interaction, *European Journal of Immunology*, Vol. 39, No. 7, 1754–1764. doi:10.1002/eji.200839028.
418. Hofmeyer, K. A., Ray, A., and Zang, X. (2008). The Contrasting Role of B7-H3, *Proceedings of the National Academy of Sciences*, Vol. 105, No. 30, 10277–10278. doi:10.1073/pnas.0805458105.
419. Zhang, T., Jiang, B., Zou, S.-T., Liu, F., and Hua, D. (2015). Overexpression of B7-H3 Augments Anti-Apoptosis of Colorectal Cancer Cells by Jak2-STAT3, *World Journal of Gastroenterology*, Vol. 21, No. 6, 1804. doi:10.3748/wjg.v21.i6.1804.
420. Teng, Y., Ross, J. L., and Cowell, J. K. (2014). The Involvement of JAK-STAT3 in Cell Motility, Invasion, and Metastasis, *JAK-STAT*, Vol. 3, No. 1, e28086. doi:10.4161/jkst.28086.
421. Zang, X., and Allison, J. P. (2007). The B7 Family and Cancer Therapy: Costimulation and Coinhibition, *Clinical Cancer Research*, Vol. 13, No. 18, 5271–5279. doi:10.1158/1078-0432.CCR-07-1030.
422. Prasad, D. V. R., Nguyen, T., Li, Z., Yang, Y., Duong, J., Wang, Y., and Dong, C. (2004). Murine B7-H3 Is a Negative Regulator of T Cells, *The Journal of Immunology*, Vol. 173, No. 4, 2500–2506. doi:10.4049/jimmunol.173.4.2500.
423. Sun, J., Chen, L., Zhang, G., Jiang, J., Zhu, M., Tan, Y., Wang, H., Lu, B., and Zhang, X. (2010). Clinical Significance and Regulation of the Costimulatory Molecule B7-H3 in Human Colorectal Carcinoma, *Cancer Immunology, Immunotherapy*, Vol. 59, No. 8, 1163–1171. doi:10.1007/s00262-010-0841-1.
424. Karapetis, C. S., Khambata-Ford, S., Jonker, D. J., O'Callaghan, C. J., Tu, D., Tebbutt, N. C., Simes, R. J., Chalchal, H., Shapiro, J. D., Robitaille, S., Price, T. J., Shepherd, L., Au, H.-J., Langer, C., Moore, M. J., and Zalcberg, J. R. (2008). *K-Ras* Mutations and Benefit from Cetuximab in Advanced Colorectal Cancer, *New England Journal of Medicine*, Vol. 359, No. 17, 1757–1765. doi:10.1056/NEJMoa0804385.
425. Vaughn, C. P., ZoBell, S. D., Furtado, L. V., Baker, C. L., and Samowitz, W. S. (2011). Frequency of *KRAS*, *BRAF*, and *NRAS* Mutations in Colorectal Cancer, *Genes, Chromosomes and Cancer*, Vol. 50, No. 5, 307–312. doi:10.1002/gcc.20854.
426. Gedde-Dahl, T., Fossum, B., Eriksen, J. A., Thorsby, E., and Gaudernack, G. (1993). T Cell Clones Specific for P21 Ras-derived Peptides: Characterization of Their Fine Specificity and HLA Restriction, *European Journal of Immunology*, Vol. 23, No. 3, 754–760. doi:10.1002/eji.1830230328.
427. Peace, D. J., Chen, W., Nelson, H., and Cheever, M. A. (1991). T Cell Recognition of Transforming Proteins Encoded by Mutated Ras Proto-Oncogenes., *Journal of Immunology (Baltimore, Md. : 1950)*, Vol. 146, No. 6, 2059–65.
428. Shono, Y., Tanimura, H., Iwahashi, M., Tsunoda, T., Tani, M., Tanaka, H., Matsuda, K., and Yamaue, H. (2003). Specific T-Cell Immunity against Ki-Ras Peptides in Patients with Pancreatic and Colorectal Cancers, *British Journal of Cancer*, Vol. 88, No. 4, 530–536. doi:10.1038/sj.bjc.6600697.
429. Khleif, S. N., Abrams, S. I., Hamilton, J. M., Bergmann-Leitner, E., Chen, A., Bastian, A., Bernstein, S., Chung, Y., Allegra, C. J., and Schlom, J. (1999). A Phase I Vaccine Trial with Peptides Reflecting Ras Oncogene Mutations of Solid Tumors, *Journal of Immunotherapy*, Vol. 22, No. 2, 155–165. doi:10.1097/00002371-199903000-00007.
430. Toubaji, A., Achta, M., Provenzano, M., Herrin, V. E., Behrens, R., Hamilton, M., Bernstein, S., Venzon, D., Gause, B., Marincola, F., and Khleif, S. N. (2008). Pilot Study of Mutant Ras Peptide-Based Vaccine as an Adjuvant Treatment in Pancreatic and Colorectal Cancers, *Cancer Immunology, Immunotherapy*, Vol. 57, No. 9, 1413–1420. doi:10.1007/s00262-008-0477-6.
431. Tran, E., Robbins, P. F., Lu, Y.-C., Prickett, T. D., Gartner, J. J., Jia, L., Pasetto, A., Zheng, Z., Ray, S., Groh, E. M., Kriley, I. R., and Rosenberg, S. A. (2016). T-Cell Transfer Therapy Targeting Mutant KRAS in Cancer, *New England Journal of Medicine*, Vol. 375, No. 23, 2255–2262. doi:10.1056/NEJMoa1609279.
432. Rahma, O. E., Hamilton, J. M., Wojtowicz, M., Dakheel, O., Bernstein, S., Liewehr, D. J., Steinberg, S. M., and Khleif, S. N. (2014). The Immunological and Clinical Effects of Mutated Ras Peptide Vaccine in Combination with IL-2, GM-CSF, or Both in Patients with Solid Tumors, *Journal of Translational Medicine*, Vol. 12, No. 1, 55. doi:10.1186/1479-5876-12-55.
433. Russo, A., Bazan, V., Iacopetta, B., Kerr, D., Soussi, T., and Gebbia, N. (2005). The TP53 Colorectal Cancer International Collaborative Study on the Prognostic and Predictive Significance of P53 Mutation: Influence of Tumor Site, Type of Mutation, and Adjuvant Treatment, *Journal of Clinical Oncology*, Vol. 23, No. 30, 7518–7528. doi:10.1200/JCO.2005.00.471.
434. van der Burg, S. H., Menon, A. G., Redeker, A., Bonnet, M.-C., Drijfhout, J. W., Tollenaar, R. A. E. M., van de Velde, C. J. H., Moingeon, P., Kuppen, P. J. K., Offringa, R., and Melief, C. J. M. (2002). Induction of P53-Specific Immune Responses in Colorectal Cancer Patients Receiving a Recombinant ALVAC-P53 Candidate Vaccine., *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*, Vol. 8, No. 5, 1019–27.
435. Speetjens, F. M., Kuppen, P. J. K., Welters, M. J. P., Essahsah, F., Voet van den Brink, A. M. E. G., Lantrua, M. G. K., Valentijn, A. R. P. M., Oostendorp, J., Fathers, L. M., Nijman, H. W., Drijfhout, J. W., van de Velde, C. J. H., Melief, C. J. M., and van der Burg, S. H. (2009). Induction of P53-Specific Immunity by a P53 Synthetic Long Peptide Vaccine in Patients Treated for Metastatic Colorectal Cancer, *Clinical Cancer Research*, Vol. 15, No. 3, 1086–1095. doi:10.1158/1078-0432.CCR-08-2227.
436. Zeestraten, E. C. M., Speetjens, F. M., Welters, M. J. P., Saadatmand, S., Stynenbosch, L. F. M., Jongen, R., Kapiteijn, E., Gelderblom, H., Nijman, H. W., Valentijn, A. R. P. M., Oostendorp, J., Fathers, L. M., Drijfhout, J. W., van de Velde, C. J. H., Kuppen, P. J. K., van der Burg, S. H., and Melief, C. J. M. (2013). Addition of Interferon- α to the P53-SLP $\text{\textcircled{R}}$ Vaccine Results in Increased Production of Interferon- γ in Vaccinated Colorectal Cancer Patients: A Phase I/II Clinical Trial, *International Journal of Cancer*, Vol. 132, No. 7, 1581–1591. doi:10.1002/ijc.27819.
437. Kerkar, S. P., Wang, Z.-F., Lasota, J., Park, T., Patel, K., Groh, E., Rosenberg, S. A., and Miettinen, M. M. (2016). MAGE-A Is More Highly Expressed than NY-ESO-1 in a Systematic Immunohistochemical Analysis of 3668 Cases, *Journal of Immunotherapy*, Vol. 39, No. 4, 181–187. doi:10.1097/CJI.000000000000119.
438. Choi, J., and Chang, H. (2012). The Expression of MAGE and SSSX, and Correlation of COX2, VEGF, and Survivin in Colorectal Cancer, *Anticancer Research*, Vol. 32, 559–564.
439. Shantha Kumara, H. M. C., Grieco, M. J., Caballero, O. L., Su, T., Ahmed, A., Ritter, E., Gnjjatic, S., Cekic, V., Old, L. J., Simpson, A. J., Cordon-Cardo, C., and Whelan, R. L. (2012). MAGE-A3 Is Highly Expressed in a Subset of Colorectal Cancer Patients., *Cancer Immunity*, Vol. 12, 16.
440. Takahashi, N., Ohkuri, T., Homma, S., Ohtake, J., Wakita, D., Togashi, Y., Kitamura, H., Todo, S., and Nishimura, T. (2012). First Clinical Trial of Cancer Vaccine Therapy with Artificially Synthesized Helper/Killer-hybrid Epitope Long Peptide of MAGE-A4 Cancer Antigen, *Cancer Science*, Vol. 103, No. 1, 150–153. doi:10.1111/j.1349-7006.2011.02106.x.

441. Kavanagh, B., Ko, A., Venook, A., Margolin, K., Zeh, H., Lotze, M., Schillinger, B., Liu, W., Lu, Y., Mitsky, P., Schilling, M., Bercovici, N., Loudovaris, M., Guillermo, R., Lee, S. M., Bender, J., Mills, B., and Fong, L. (2007). Vaccination of Metastatic Colorectal Cancer Patients With Matured Dendritic Cells Loaded With Multiple Major Histocompatibility Complex Class I Peptides, *Journal of Immunotherapy*, Vol. 30, No. 7, 762–772. doi:10.1097/CJI.0b013e318133451c.
442. Jiang, S., Good, D., and Wei, M. Q. (2019). Vaccinations for Colorectal Cancer: Progress, Strategies, and Novel Adjuvants, *International Journal of Molecular Sciences*, Vol. 20, No. 14, 3403. doi:10.3390/ijms20143403.
443. De Vries, N., Swets, M., Vahrmeijer, A., Hokland, M., and Kuppen, P. (2016). The Immunogenicity of Colorectal Cancer in Relation to Tumor Development and Treatment, *International Journal of Molecular Sciences*, Vol. 17, No. 7, 1030. doi:10.3390/ijms17071030.
444. Hu, Z., Ott, P. A., and Wu, C. J. (2018). Towards Personalized, Tumour-Specific, Therapeutic Vaccines for Cancer, *Nature Reviews Immunology*, Vol. 18, No. 3, 168–182. doi:10.1038/nri.2017.131.
445. Shah, B. A., Holden, J. A., Lenzo, J. C., Hadjigol, S., and O'Brien-Simpson, N. M. (2025). Multi-Disciplinary Approaches Paving the Way for Clinically Effective Peptide Vaccines for Cancer, *Npj Vaccines*, Vol. 10, No. 1, 68. doi:10.1038/s41541-025-01118-9.
446. Abd-Aziz, N., and Poh, C. L. (2022). Development of Peptide-Based Vaccines for Cancer, *Journal of Oncology*, Vol. 2022, 1–17. doi:10.1155/2022/9749363.
447. Buonaguro, L., and Tagliamonte, M. (2023). Peptide-Based Vaccine for Cancer Therapies, *Frontiers in Immunology*, Vol. 14. doi:10.3389/fimmu.2023.1210044.
448. Zahedipour, F., Jamialahmadi, K., Zamani, P., and Reza Jaafari, M. (2023). Improving the Efficacy of Peptide Vaccines in Cancer Immunotherapy, *International Immunopharmacology*, Vol. 123, 110721. doi:10.1016/j.intimp.2023.110721.
449. Paston, S. J., Brentville, V. A., Symonds, P., and Durrant, L. G. (2021). Cancer Vaccines, Adjuvants, and Delivery Systems, *Frontiers in Immunology*, Vol. 12. doi:10.3389/fimmu.2021.627932.
450. Potluri, H. K., Ng, T. L., Newton, M. A., and McNeel, D. G. (2022). GM-CSF Elicits Antibodies to Tumor-Associated Proteins When Used as a Prostate Cancer Vaccine Adjuvant, *Cancer Immunology, Immunotherapy*, Vol. 71, No. 9, 2267–2275. doi:10.1007/s00262-022-03150-3.
451. Disis, M. L., Bernhard, H., Shiota, F. M., Hand, S. L., Gralow, J. R., Huseby, E. S., Gillis, S., and Cheever, M. A. (1996). Granulocyte-Macrophage Colony-Stimulating Factor: An Effective Adjuvant for Protein and Peptide-Based Vaccines., *Blood*, Vol. 88, No. 1, 202–10.
452. Taylor, B. C., and Balko, J. M. (2022). Mechanisms of MHC-I Downregulation and Role in Immunotherapy Response, *Frontiers in Immunology*, Vol. 13. doi:10.3389/fimmu.2022.844866.
453. Carretero, F. J., del Campo, A. B., Flores-Martín, J. F., Mendez, R., García-Lopez, C., Cozar, J. M., Adams, V., Ward, S., Cabrera, T., Ruiz-Cabello, F., Garrido, F., and Aptsiauri, N. (2016). Frequent HLA Class I Alterations in Human Prostate Cancer: Molecular Mechanisms and Clinical Relevance, *Cancer Immunology, Immunotherapy*, Vol. 65, No. 1, 47–59. doi:10.1007/s00262-015-1774-5.
454. Li, W., Joshi, M., Singhania, S., Ramsey, K., and Murthy, A. (2014). Peptide Vaccine: Progress and Challenges, *Vaccines*, Vol. 2, No. 3, 515–536. doi:10.3390/vaccines2030515.
455. Hubbard, J. M., Tóke, E. R., Moretto, R., Graham, R. P., Youssoufian, H., Lórinz, O., Molnár, L., Csiszovszki, Z., Mitchell, J. L., Wessling, J., Tóth, J., and Cremolini, C. (2022). Safety and Activity of PolyPEP1018 Combined with Maintenance Therapy in Metastatic Colorectal Cancer: An Open-Label, Multicenter, Phase Ib Study, *Clinical Cancer Research*, Vol. 28, No. 13, 2818–2829. doi:10.1158/1078-0432.CCR-22-0112.
456. Kopetz, S., Prenen, H., Sharma, S., Van Cutsem, E., Mayol, J., Trapani, F., Bogenrieder, T., and Lenz, H. (2021). SO-11 KISIMA-01 Trial: Safety, Tolerability and Immunogenicity of ATP128 with or without Ezabenlimab (BI 754091) in Patients with Stage IV Colorectal Cancer – Preliminary Results from a Phase 1b Study, *Annals of Oncology*, Vol. 32, S206–S207. doi:10.1016/j.annonc.2021.05.035.
457. Shahnazari, M., Samadi, P., Pourjafar, M., and Jalali, A. (2020). Therapeutic Vaccines for Colorectal Cancer: The Progress and Future Prospect, *International Immunopharmacology*, Vol. 88, 106944. doi:10.1016/j.intimp.2020.106944.
458. Shariati, A., Khezrpour, A., Shariati, F., Afkhami, H., Yarahmadi, A., Alavimanesht, S., Kamrani, S., Modarressi, M. H., and Khani, P. (2025). DNA Vaccines as Promising Immuno-Therapeutics against Cancer: A New Insight, *Frontiers in Immunology*, Vol. 15. doi:10.3389/fimmu.2024.1498431.
459. Sobhani, N., Scaggiante, B., Morris, R., Chai, D., Catalano, M., Tardiel-Cyril, D. R., Neeli, P., Roviello, G., Mondani, G., and Li, Y. (2022). Therapeutic Cancer Vaccines: From Biological Mechanisms and Engineering to Ongoing Clinical Trials, *Cancer Treatment Reviews*, Vol. 109, 102429. doi:10.1016/j.ctrv.2022.102429.
460. Ramsay, R. G., and Gonda, T. J. (2008). MYB Function in Normal and Cancer Cells, *Nature Reviews Cancer*, Vol. 8, No. 7, 523–534. doi:10.1038/nrc2439.
461. Cross, R. S., Malaterre, J., Davenport, A. J., Carpinteri, S., Anderson, R. L., Darcy, P. K., and Ramsay, R. G. (2015). Therapeutic DNA Vaccination against Colorectal Cancer by Targeting the MYB Oncoprotein, *Clinical & Translational Immunology*, Vol. 4, No. 1. doi:10.1038/cti.2014.29.
462. Williams, B. B., Wall, M., Miao, R. Y., Williams, B., Bertoncello, I., Kershaw, M. H., Mantamadiotis, T., Haber, M., Norris, M. D., Gautam, A., Darcy, P. K., and Ramsay, R. G. (2008). Induction of T Cell-Mediated Immunity Using a c-Myb DNA Vaccine in a Mouse Model of Colon Cancer, *Cancer Immunology, Immunotherapy*, Vol. 57, No. 11, 1635–1645. doi:10.1007/s00262-008-0497-2.
463. Lu, B., Lim, J. M., Yu, B., Song, S., Neeli, P., Sobhani, N., K, P., Bonam, S. R., Kurapati, R., Zheng, J., and Chai, D. (2024). The Next-Generation DNA Vaccine Platforms and Delivery Systems: Advances, Challenges and Prospects, *Frontiers in Immunology*, Vol. 15. doi:10.3389/fimmu.2024.1332939.
464. Pandya, A., Shah, Y., Kothari, N., Postwala, H., Shah, A., Parekh, P., and Chorawala, M. R. (2023). The Future of Cancer Immunotherapy: DNA Vaccines Leading the Way, *Medical Oncology*, Vol. 40, No. 7, 200. doi:10.1007/s12032-023-02060-3.
465. Pardi, N., Hogan, M. J., Porter, F. W., and Weissman, D. (2018). mRNA Vaccines — a New Era in Vaccinology, *Nature Reviews Drug Discovery*, Vol. 17, No. 4, 261–279. doi:10.1038/nrd.2017.243.
466. Guasp, P., Reiche, C., Sethna, Z., and Balachandran, V. P. (2024). RNA Vaccines for Cancer: Principles to Practice, *Cancer Cell*, Vol. 42, No. 7, 1163–1184. doi:10.1016/j.ccell.2024.05.005.
467. Katopodi, T., Petanidis, S., Grigoriadou, E., Anastakis, D., Charalampidis, C., Chatziprodromidou, I., Floros, G., Eskitzis, P., Zarogoulidis, P., Koulouris, C., Sevva, C., Papadopoulos, K., Roulia, P., Mantalovas, S., Dagher, M., Karakousis, A. V., Varsamis, N., Vlassopoulos, K., Theodorou, V., Mystakidou, C.

- M., Katsios, N. I., Farmakis, K., and Kosmidis, C. (2024). Immune Specific and Tumor-Dependent mRNA Vaccines for Cancer Immunotherapy: Reprogramming Clinical Translation into Tumor Editing Therapy, *Pharmaceutics*, Vol. 16, No. 4, 455. doi:10.3390/pharmaceutics16040455.
468. Cafri, G., Gartner, J. J., Hopson, K., Meehan, R. S., Zaks, T. Z., Robbins, P., and Rosenberg, S. A. (2019). Immunogenicity and Tolerability of Personalized mRNA Vaccine mRNA-4650 Encoding Defined Neoantigens Expressed by the Autologous Cancer, *Journal of Clinical Oncology*, Vol. 37, No. 15_suppl, 2643–2643. doi:10.1200/JCO.2019.37.15_suppl.2643.
469. Burris, H. A., Patel, M. R., Cho, D. C., Clarke, J. M., Gutierrez, M., Zaks, T. Z., Frederick, J., Hopson, K., Mody, K., Binanti-Berube, A., Robert-Tissot, C., Goldstein, B., Breton, B., Sun, J., Zhong, S., Pruitt, S. K., Keating, K., Meehan, R. S., and Gainor, J. F. (2019). A Phase I Multicenter Study to Assess the Safety, Tolerability, and Immunogenicity of mRNA-4157 Alone in Patients with Resected Solid Tumors and in Combination with Pembrolizumab in Patients with Unresectable Solid Tumors, *Journal of Clinical Oncology*, Vol. 37, No. 15_suppl, 2523–2523. doi:10.1200/JCO.2019.37.15_suppl.2523.
470. Asimgil, H., Ertetik, U., Çevik, N. C., Ekizce, M., Doğruöz, A., Gökalp, M., Arik-Sever, E., Istvanffy, R., Friess, H., Ceyhan, G. O., and Demir, I. E. (2022). Targeting the Undruggable Oncogenic KRAS: The Dawn of Hope, *JCI Insight*, Vol. 7, No. 1. doi:10.1172/jci.insight.153688.
471. Youssef, M., Hitti, C., Puppim Chaves Fulber, J., and Kamen, A. A. (2023). Enabling mRNA Therapeutics: Current Landscape and Challenges in Manufacturing, *Biomolecules*, Vol. 13, No. 10, 1497. doi:10.3390/biom13101497.
472. Diao, L., and Liu, M. (2023). Rethinking Antigen Source: Cancer Vaccines Based on Whole Tumor Cell/Tissue Lysate or Whole Tumor Cell, *Advanced Science*, Vol. 10, No. 22. doi:10.1002/advs.202300121.
473. Alzeeb, G., Tortorelli, C., Taleb, J., De Luca, F., Berge, B., Bardet, C., Limagne, E., Brun, M., Chalus, L., Pinteur, B., Bravetti, P., Gongora, C., Apetoh, L., and Ghiringhelli, F. (2024). Efficacy of Novel Allogeneic Cancer Cells Vaccine to Treat Colorectal Cancer, *Frontiers in Oncology*, Vol. 14. doi:10.3389/fonc.2024.1427428.
474. Baars, A., Claessen, A. M. E., Wagstaff, J., Giaccone, G., Scheper, R. J., Meijer, S., Schakel, M. J. A. G., Gall, H. E., Meijer, C. J. L. M., Vermorken, J. B., Pinedo, H. M., and van den Eertwegh, A. J. M. (2002). A Phase II Study of Active Specific Immunotherapy and 5-FU/Leucovorin as Adjuvant Therapy for Stage III Colon Carcinoma, *British Journal of Cancer*, Vol. 86, No. 8, 1230–1234. doi:10.1038/sj.bjc.6600254.
475. Yarchoan, M., Huang, C., Zhu, Q., Ferguson, A. K., Durham, J. N., Anders, R. A., Thompson, E. D., Rozich, N. S., Thomas, D. L., Nauroth, J. M., Rodriguez, C., Osipov, A., De Jesus-Acosta, A., Le, D. T., Murphy, A. G., Laheru, D., Donehower, R. C., Jaffee, E. M., Zheng, L., and Azad, N. S. (2020). A Phase 2 Study of GVAX Colon Vaccine with Cyclophosphamide and Pembrolizumab in Patients with Mismatch Repair Proficient Advanced Colorectal Cancer, *Cancer Medicine*, Vol. 9, No. 4, 1485–1494. doi:10.1002/cam4.2763.
476. Wang, D.-K., Zuo, Q., He, Q.-Y., and Li, B. (2021). Targeted Immunotherapies in Gastrointestinal Cancer: From Molecular Mechanisms to Implications, *Frontiers in Immunology*, Vol. 12. doi:10.3389/fimmu.2021.705999.
477. Rodriguez, J., Castañón, E., Perez-Gracia, J. L., Rodriguez, I., Viudez, A., Alfaro, C., Oñate, C., Perez, G., Rotellar, F., Inogés, S., López-Díaz de Cerio, A., Resano, L., Ponz-Sarvisé, M., Rodríguez-Ruiz, M. E., Chopitea, A., Vera, R., and Melero, I. (2018). A Randomized Phase II Clinical Trial of Dendritic Cell Vaccination Following Complete Resection of Colon Cancer Liver Metastasis, *Journal for ImmunoTherapy of Cancer*, Vol. 6, No. 1, 96. doi:10.1186/s40425-018-0405-z.
478. Toh, H. C., Wang, W.-W., Chia, W. K., Kvistborg, P., Sun, L., Teo, K., Phoon, Y. P., Soe, Y., Tan, S. H., Hee, S. W., Foo, K. F., Ong, S., Koo, W. H., Zocca, M.-B., and Claesson, M. H. (2009). Clinical Benefit of Allogeneic Melanoma Cell Lysate-Pulsed Autologous Dendritic Cell Vaccine in MAGE-Positive Colorectal Cancer Patients, *Clinical Cancer Research*, Vol. 15, No. 24, 7726–7736. doi:10.1158/1078-0432.CCR-09-1537.
479. Muthukutty, P., Woo, H. Y., and Yoo, S. Y. (2025). Therapeutic Colorectal Cancer Vaccines: Emerging Modalities and Translational Opportunities, *Vaccines*, Vol. 13, No. 7, 689. doi:10.3390/vaccines13070689.
480. Nami, S., Mohammadi, R., Vakili, M., Khezripour, K., Mirzaei, H., and Morovati, H. (2019). Fungal Vaccines, Mechanism of Actions and Immunology: A Comprehensive Review, *Biomedicine & Pharmacotherapy*, Vol. 109, 333–344. doi:10.1016/j.biopha.2018.10.075.
481. Morse, M. A., Chaudhry, A., Gabitzsch, E. S., Hobeika, A. C., Osada, T., Clay, T. M., Amalfitano, A., Burnett, B. K., Devi, G. R., Hsu, D. S., Xu, Y., Balcaitis, S., Dua, R., Nguyen, S., Balint, J. P., Jones, F. R., and Lyerly, H. K. (2013). Novel Adenoviral Vector Induces T-Cell Responses despite Anti-Adenoviral Neutralizing Antibodies in Colorectal Cancer Patients, *Cancer Immunology, Immunotherapy*, Vol. 62, No. 8, 1293–1301. doi:10.1007/s00262-013-1400-3.
482. Redman, J. M., Tsai, Y.-T., Weinberg, B. A., Donahue, R. N., Gandhy, S., Gatti-Mays, M. E., Abdul Sater, H., Bilusic, M., Cordes, L. M., Steinberg, S. M., Marte, J. L., Jochems, C., Kim, S. S., Marshall, J. L., McMahon, S., Redmond, E., Schlom, J., Gulley, J. L., and Strauss, J. (2022). A Randomized Phase II Trial of MFOLFOX6 + Bevacizumab Alone or with AdCEA Vaccine + Avelumab Immunotherapy for Untreated Metastatic Colorectal Cancer, *The Oncologist*, Vol. 27, No. 3, 198–209. doi:10.1093/oncolo/oyab046.
483. Youn, J. W., Hur, S.-Y., Woo, J. W., Kim, Y.-M., Lim, M. C., Park, S. Y., Seo, S. S., No, J. H., Kim, B.-G., Lee, J.-K., Shin, S. J., Kim, K., Chaney, M. F., Choi, Y.-J., Suh, Y. S., Park, J. S., and Sung, Y. C. (2020). Pembrolizumab plus GX-188E Therapeutic DNA Vaccine in Patients with HPV-16-Positive or HPV-18-Positive Advanced Cervical Cancer: Interim Results of a Single-Arm, Phase 2 Trial, *The Lancet Oncology*, Vol. 21, No. 12, 1653–1660. doi:10.1016/S1470-2045(20)30486-1.
484. Kantoff, P. W., Higano, C. S., Shore, N. D., Berger, E. R., Small, E. J., Penson, D. F., Redfern, C. H., Ferrari, A. C., Dreicer, R., Sims, R. B., Xu, Y., Frohlich, M. W., and Schellhammer, P. F. (2010). Sipuleucel-T Immunotherapy for Castration-Resistant Prostate Cancer, *New England Journal of Medicine*, Vol. 363, No. 5, 411–422. doi:10.1056/NEJMoa1001294.
485. Schwartzentruber, D. J., Lawson, D. H., Richards, J. M., Conry, R. M., Miller, D. M., Treisman, J., Gailani, F., Riley, L., Conlon, K., Pockaj, B., Kendra, K. L., White, R. L., Gonzalez, R., Kuzel, T. M., Curti, B., Leming, P. D., Whitman, E. D., Balkissoon, J., Reintgen, D. S., Kaufman, H., Marincola, F. M., Merino, M. J., Rosenberg, S. A., Choyke, P., Vena, D., and Hwu, P. (2011). Gp100 Peptide Vaccine and Interleukin-2 in Patients with Advanced Melanoma, *New England Journal of Medicine*, Vol. 364, No. 22, 2119–2127. doi:10.1056/NEJMoa1012863.
486. Schuster, S. J., Neelapu, S. S., Gause, B. L., Janik, J. E., Muggia, F. M., Gockerman, J. P., Winter, J. N., Flowers, C. R., Nikcevich, D. A., Sotomayor, E. M., McGaughey, D. S., Jaffe, E. S., Chong, E. A., Reynolds, C. W., Berry, D. A., Santos, C. F., Popa, M. A., McCord, A. M., and Kwak, L. W. (2011). Vaccination with Patient-Specific Tumor-Derived Antigen in First Remission Improves Disease-Free Survival in Follicular Lymphoma, *Journal of Clinical*

- Oncology*, Vol. 29, No. 20, 2787–2794. doi:10.1200/JCO.2010.33.3005.
487. Verma, C., Pawar, V., Srivastava, S., Tyagi, A., Kaushik, G., Shukla, S., and Kumar, V. (2023). Cancer Vaccines in the Immunotherapy Era: Promise and Potential, *Vaccines*, Vol. 11, No. 12, 1783. doi:10.3390/vaccines11121783.
488. Saxena, M., van der Burg, S. H., Melief, C. J. M., and Bhardwaj, N. (2021). Therapeutic Cancer Vaccines, *Nature Reviews Cancer*, Vol. 21, No. 6, 360–378. doi:10.1038/s41568-021-00346-0.
489. Sellars, M. C., Wu, C. J., and Fritsch, E. F. (2022). Cancer Vaccines: Building a Bridge over Troubled Waters, *Cell*, Vol. 185, No. 15, 2770–2788. doi:10.1016/j.cell.2022.06.035.
490. Pounraj, S., Chen, S., Ma, L., Mazzieri, R., Dolcetti, R., and Rehm, B. H. A. (2024). Targeting Tumor Heterogeneity with Neoantigen-Based Cancer Vaccines, *Cancer Research*, Vol. 84, No. 3, 353–363. doi:10.1158/0008-5472.CAN-23-2042.
491. Liu, K.-J., Wang, C.-C., Chen, L.-T., Cheng, A.-L., Lin, D.-T., Wu, Y.-C., Yu, W.-L., Hung, Y.-M., Yang, H.-Y., Juang, S.-H., and Whang-Peng, J. (2004). Generation of Carcinoembryonic Antigen (CEA)-Specific T-Cell Responses in HLA-A*0201 and HLA-A*2402 Late-Stage Colorectal Cancer Patients after Vaccination with Dendritic Cells Loaded with CEA Peptides, *Clinical Cancer Research*, Vol. 10, No. 8, 2645–2651. doi:10.1158/1078-0432.CCR-03-0430.
492. Zhang, B., Liu, J., Mo, Y., Zhang, K., Huang, B., and Shang, D. (2024). CD8+ T Cell Exhaustion and Its Regulatory Mechanisms in the Tumor Microenvironment: Key to the Success of Immunotherapy, *Frontiers in Immunology*, Vol. 15. doi:10.3389/fimmu.2024.1476904.
493. Jiang, Y., Li, Y., and Zhu, B. (2015). T-Cell Exhaustion in the Tumor Microenvironment, *Cell Death & Disease*, Vol. 6, No. 6, e1792–e1792. doi:10.1038/cddis.2015.162.
494. Stone, L. (2023). Singling out the Immune-Suppressive TME in Prostate Cancer, *Nature Reviews Urology*, Vol. 20, No. 4, 199–199. doi:10.1038/s41585-023-00758-7.
495. Wang, C., Singer, M., and Anderson, A. C. (2017). Molecular Dissection of CD8 + T-Cell Dysfunction, *Trends in Immunology*, Vol. 38, No. 8, 567–576. doi:10.1016/j.it.2017.05.008.
496. Schreiber, R. D., Old, L. J., and Smyth, M. J. (2011). Cancer Immunoediting: Integrating Immunity's Roles in Cancer Suppression and Promotion, *Science*, Vol. 331, No. 6024, 1565–1570. doi:10.1126/science.1203486.
497. Llosa, N. J., Cruise, M., Tam, A., Wicks, E. C., Hechenbleikner, E. M., Taube, J. M., Blosser, R. L., Fan, H., Wang, H., Luber, B. S., Zhang, M., Papadopoulos, N., Kinzler, K. W., Vogelstein, B., Sears, C. L., Anders, R. A., Pardoll, D. M., and Housseau, F. (2015). The Vigorous Immune Microenvironment of Microsatellite Instable Colon Cancer Is Balanced by Multiple Counter-Inhibitory Checkpoints, *Cancer Discovery*, Vol. 5, No. 1, 43–51. doi:10.1158/2159-8290.CD-14-0863.
498. Le, D. T., Uram, J. N., Wang, H., Bartlett, B. R., Kemberling, H., Eyring, A. D., Skora, A. D., Luber, B. S., Azad, N. S., Laheru, D., Biedrzycki, B., Donehower, R. C., Zaheer, A., Fisher, G. A., Crocenzi, T. S., Lee, J. J., Duffy, S. M., Goldberg, R. M., de la Chapelle, A., Koshiji, M., Bhajee, F., Huebner, T., Hruban, R. H., Wood, L. D., Cuka, N., Pardoll, D. M., Papadopoulos, N., Kinzler, K. W., Zhou, S., Cornish, T. C., Taube, J. M., Anders, R. A., Eshleman, J. R., Vogelstein, B., and Diaz, L. A. (2015). PD-1 Blockade in Tumors with Mismatch-Repair Deficiency, *New England Journal of Medicine*, Vol. 372, No. 26, 2509–2520. doi:10.1056/NEJMoa1500596.
499. Ganesh, K., Stadler, Z. K., Cercek, A., Mendelsohn, R. B., Shia, J., Segal, N. H., and Diaz, L. A. (2019). Immunotherapy in Colorectal Cancer: Rationale, Challenges and Potential, *Nature Reviews Gastroenterology & Hepatology*, Vol. 16, No. 6, 361–375. doi:10.1038/s41575-019-0126-x.
500. Martin, J., Petrillo, A., Smyth, E. C., Shaida, N., Khwaja, S., Cheow, H., Duckworth, A., Heister, P., Praseedom, R., Jah, A., Balakrishnan, A., Harper, S., Liao, S., Kosmolaptsis, V., and Huguet, E. (2020). Colorectal Liver Metastases: Current Management and Future Perspectives, *World Journal of Clinical Oncology*, Vol. 11, No. 10, 761–808. doi:10.5306/wjco.v11.i10.761.
501. Yu, J., Green, M. D., Li, S., Sun, Y., Journey, S. N., Choi, J. E., Rizvi, S. M., Qin, A., Waninger, J. J., Lang, X., Chopra, Z., El Naqa, I., Zhou, J., Bian, Y., Jiang, L., Tezel, A., Skvarce, J., Achar, R. K., Sitto, M., Rosen, B. S., Su, F., Narayanan, S. P., Cao, X., Wei, S., Szeliga, W., Vatan, L., Mayo, C., Morgan, M. A., Schonewolf, C. A., Cuneo, K., Kryczek, I., Ma, V. T., Lao, C. D., Lawrence, T. S., Ramnath, N., Wen, F., Chinnaiyan, A. M., Cieslik, M., Alva, A., and Zou, W. (2021). Liver Metastasis Restrains Immunotherapy Efficacy via Macrophage-Mediated T Cell Elimination, *Nature Medicine*, Vol. 27, No. 1, 152–164. doi:10.1038/s41591-020-1131-x.
502. Ringelhan, M., Pfister, D., O'Connor, T., Pikarsky, E., and Heikenwalder, M. (2018). The Immunology of Hepatocellular Carcinoma, *Nature Immunology*, Vol. 19, No. 3, 222–232. doi:10.1038/s41590-018-0044-z.
503. Tume, P. C., Hellmann, M. D., Hamid, O., Tsai, K. K., Loo, K. L., Gubens, M. A., Rosenblum, M., Harview, C. L., Taube, J. M., Handley, N., Khurana, N., Nosrati, A., Krummel, M. F., Tucker, A., Sosa, E. V., Sanchez, P. J., Banayan, N., Osorio, J. C., Nguyen-Kim, D. L., Chang, J., Shintaku, I. P., Boasberg, P. D., Taylor, E. J., Munster, P. N., Algazi, A. P., Chmielowski, B., Dummer, R., Grogan, T. R., Elashoff, D., Hwang, J., Goldinger, S. M., Garon, E. B., Pierce, R. H., and Daud, A. (2017). Liver Metastasis and Treatment Outcome with Anti-PD-1 Monoclonal Antibody in Patients with Melanoma and NSCLC, *Cancer Immunology Research*, Vol. 5, No. 5, 417–424. doi:10.1158/2326-6066.CIR-16-0325.
504. Zitvogel, L., Daillère, R., Roberti, M. P., Routy, B., and Kroemer, G. (2017). Anticancer Effects of the Microbiome and Its Products, *Nature Reviews Microbiology*, Vol. 15, No. 8, 465–478. doi:10.1038/nrmicro.2017.44.
505. Routy, B., Le Chatelier, E., Derosa, L., Duong, C. P. M., Alou, M. T., Daillère, R., Fluckiger, A., Messaoudene, M., Rauber, C., Roberti, M. P., Fidelle, M., Flament, C., Poirier-Colame, V., Opolon, P., Klein, C., Iribarren, K., Mondragón, L., Jacquelot, N., Qu, B., Ferrere, G., Clémenson, C., Mezquita, L., Masip, J. R., Naltet, C., Brosseau, S., Kaderbhai, C., Richard, C., Rizvi, H., Levenez, F., Galleron, N., Quinquis, B., Pons, N., Ryffel, B., Minard-Colin, V., Gonin, P., Soria, J.-C., Deutsch, E., Lloriot, Y., Ghiringhelli, F., Zalcman, G., Goldwasser, F., Escudier, B., Hellmann, M. D., Eggermont, A., Raouf, D., Albiges, L., Kroemer, G., and Zitvogel, L. (2018). Gut Microbiome Influences Efficacy of PD-1–Based Immunotherapy against Epithelial Tumors, *Science*, Vol. 359, No. 6371, 91–97. doi:10.1126/science.aan3706.
506. Gopalakrishnan, V., Helmink, B. A., Spencer, C. N., Reuben, A., and Wargo, J. A. (2018). The Influence of the Gut Microbiome on Cancer, Immunity, and Cancer Immunotherapy, *Cancer Cell*, Vol. 33, No. 4, 570–580. doi:10.1016/j.ccell.2018.03.015.
507. Zhang, W., An, Y., Qin, X., Wu, X., Wang, X., Hou, H., Song, X., Liu, T., Wang, B., Huang, X., and Cao, H. (2021). Gut Microbiota-Derived Metabolites in Colorectal Cancer: The Bad and the Challenges, *Frontiers in Oncology*, Vol. 11. doi:10.3389/fonc.2021.739648.
508. Mowat, A. M., and Agace, W. W. (2014). Regional Specialization within the Intestinal Immune System, *Nature Reviews Immunology*, Vol. 14, No. 10, 667–685. doi:10.1038/nri3738.

509. Hooper, L. V., Littman, D. R., and Macpherson, A. J. (2012). Interactions Between the Microbiota and the Immune System, *Science*, Vol. 336, No. 6086, 1268–1273. doi:10.1126/science.1223490.
510. Van Cutsem, E., Cervantes, A., Adam, R., Sobrero, A., Van Krieken, J. H., Aderka, D., Aranda Aguilar, E., Bardelli, A., Benson, A., Bodoky, G., Ciardiello, F., D'Hoore, A., Diaz-Rubio, E., Douillard, J.-Y., Ducreux, M., Falcone, A., Grothey, A., Gruenberger, T., Haustermans, K., Heinemann, V., Hoff, P., Köhne, C.-H., Labianca, R., Laurent-Puig, P., Ma, B., Maughan, T., Muro, K., Normanno, N., Österlund, P., Oyen, W. J. G., Papamichael, D., Pentheroudakis, G., Pfeiffer, P., Price, T. J., Punt, C., Ricke, J., Roth, A., Salazar, R., Scheithauer, W., Schmoll, H. J., Taberero, J., Taïeb, J., Tejpar, S., Wasan, H., Yoshino, T., Zaanan, A., and Arnold, D. (2016). ESMO Consensus Guidelines for the Management of Patients with Metastatic Colorectal Cancer, *Annals of Oncology*, Vol. 27, No. 8, 1386–1422. doi:10.1093/annonc/mdw235.
511. Benson, A. B., Venook, A. P., Al-Hawary, M. M., Arain, M. A., Chen, Y.-J., Ciombor, K. K., Cohen, S., Cooper, H. S., Deming, D., Farkas, L., Garrido-Laguna, I., Grem, J. L., Gunn, A., Hecht, J. R., Hoffe, S., Hubbard, J., Hunt, S., Johung, K. L., Kirilcuk, N., Krishnamurthi, S., Messersmith, W. A., Meyerhardt, J., Miller, E. D., Mulcahy, M. F., Nurkin, S., Overman, M. J., Parikh, A., Patel, H., Pedersen, K., Saltz, L., Schneider, C., Shibata, D., Skibber, J. M., Sofocleous, C. T., Stoffel, E. M., Stotsky-Himelfarb, E., Willett, C. G., Gregory, K. M., and Gurski, L. A. (2021). Colon Cancer, Version 2.2021, NCCN Clinical Practice Guidelines in Oncology, *Journal of the National Comprehensive Cancer Network*, Vol. 19, No. 3, 329–359. doi:10.6004/jnccn.2021.0012.
512. Li, L., Goedegebuure, S. P., and Gillanders, W. E. (2017). Preclinical and Clinical Development of Neoantigen Vaccines, *Annals of Oncology*, Vol. 28, xii11–xii17. doi:10.1093/annonc/mdx681.
513. Jaroslowski, S., Caban, A., and Toumi, M. (2015). Sipuleucel-T (Provenge®): Autopsy of an Innovative Change of Paradigm in Cancer Treatment, *Value in Health*, Vol. 18, No. 7, A479. doi:10.1016/j.jval.2015.09.1294.
514. Baghdadi, R. (2010). Dendreon vs CMS: Why the Provenge Coverage Controversy Is Bigger than Just One Product, *Oncology*, Vol. 24, No. 10.
515. Madan, R. A., and Gulley, J. L. (2011). Sipuleucel-T: Harbinger of a New Age of Therapeutics for Prostate Cancer, *Expert Review of Vaccines*, Vol. 10, No. 2, 141–150. doi:10.1586/erv.10.173.
516. Andrews, A. (2015). Treating with Checkpoint Inhibitors-Figure \$1 Million per Patient, *American Health & Drug Benefits*, Vol. 8, No. Spec Issue, 9.
517. Paul, J., Mitchell, A. P., Kesselheim, A. S., and Rome, B. N. (2024). Trends in Prices of Checkpoint Inhibitors in the US, 2016–2023., *Journal of Clinical Oncology*, Vol. 42, No. 16_suppl, 11075–11075. doi:10.1200/JCO.2024.42.16_suppl.11075.
518. Zhang, H., Patenaude, B., Ma, C., and Fang, H. (2023). Vaccine Pricing Strategies in China, *BMJ Global Health*, Vol. 8, No. 7, e011405. doi:10.1136/bmjgh-2022-011405.
519. Soria-Guerra, R. E., Nieto-Gomez, R., Govea-Alonso, D. O., and Rosales-Mendoza, S. (2015). An Overview of Bioinformatics Tools for Epitope Prediction: Implications on Vaccine Development, *Journal of Biomedical Informatics*, Vol. 53, 405–414. doi:10.1016/j.jbi.2014.11.003.
520. María, R. R., Arturo, C. J., Alicia, J. A., Paulina, M. G., and Gerardo, A. O. (2017). The Impact of Bioinformatics on Vaccine Design and Development, *Vaccines*, Intech. doi:10.5772/intechopen.69273.
521. Bayat, A. (2002). Science, Medicine, and the Future: Bioinformatics, *BMJ*, Vol. 324, No. 7344, 1018–1022. doi:10.1136/bmj.324.7344.1018.
522. Bioinformatics. (n.d.). *IPB University*, from [https://www.ipb.ac.id/page/bioinformatics/#:~:text=Bioinformatic%20is%20concerned%20with%20the,health%20\(one%2Dhealth\)](https://www.ipb.ac.id/page/bioinformatics/#:~:text=Bioinformatic%20is%20concerned%20with%20the,health%20(one%2Dhealth),), accessed 11-8-2025.
523. Mitra, D., Mitra, D., Sabri Bensaad, M., Sinha, S., Pant, K., Pant, M., Priyadarshini, A., Singh, P., Dassamiour, S., Hambaba, L., Panneerselvam, P., and Das Mohapatra, P. K. (2022). Evolution of Bioinformatics and Its Impact on Modern Bio-Science in the Twenty-First Century: Special Attention to Pharmacology, Plant Science and Drug Discovery, *Computational Toxicology*, Vol. 24, 100248. doi:10.1016/j.comtox.2022.100248.
524. Urrutia-Baca, V. H., Gomez-flores, R., De La Garza-Ramos, M. A., Tamez-guerra, P., Lucio-sauceda, D. G., and Rodríguez-padilla, M. C. (2019). Immunoinformatics Approach to Design a Novel Epitope-Based Oral Vaccine against Helicobacter Pylori, *Journal of Computational Biology*, Vol. 26, No. 10, 1177–1190. doi:10.1089/cmb.2019.0062.
525. Ali, S. A., Almofti, Y. A., and Abd-elrahman, K. A. (2019). Immunoinformatics Approach for Multiepitopes Vaccine Prediction against Glycoprotein B of Avian Infectious Laryngotracheitis Virus, *Advances in Bioinformatics*, Vol. 2019, 1–23. doi:10.1155/2019/1270485.
526. Dhama, K., Sharun, K., Tiwari, R., Dadar, M., Malik, Y. S., Singh, K. P., and Chaicumpa, W. (2020). COVID-19, an Emerging Coronavirus Infection: Advances and Prospects in Designing and Developing Vaccines, Immunotherapeutics, and Therapeutics, *Human Vaccines & Immunotherapeutics*, Vol. 16, No. 6, 1232–1238. doi:10.1080/21645515.2020.1735227.
527. Schaap-Johansen, A.-L., Vujović, M., Borch, A., Hadrup, S. R., and Marcatili, P. (2021). T Cell Epitope Prediction and Its Application to Immunotherapy, *Frontiers in Immunology*, Vol. 12. doi:10.3389/fimmu.2021.712488.
528. Calvo-Calle, J. M., Oliveira, G. A., Watta, C. O., Soverow, J., Parra-Lopez, C., and Nardin, E. H. (2006). A Linear Peptide Containing Minimal T- and B-Cell Epitopes of Plasmodium Falciparum Circumsporozoite Protein Elicits Protection against Transgenic Sporozoite Challenge., *Infection and Immunity*, Vol. 74, No. 12, 6929–39. doi:10.1128/IAI.01151-06.
529. Krishnarjuna, B., Andrew, D., MacRaild, C. A., Morales, R. A. V., Beeson, J. G., Anders, R. F., Richards, J. S., and Norton, R. S. (2016). Strain-Transcending Immune Response Generated by Chimeras of the Malaria Vaccine Candidate Merozoite Surface Protein 2, *Scientific Reports*, Vol. 6, No. 1, 20613. doi:10.1038/srep20613.
530. Peng, M., Mo, Y., Wang, Y., Wu, P., Zhang, Y., Xiong, F., Guo, C., Wu, X., Li, Y., Li, X., Li, G., Xiong, W., and Zeng, Z. (2019). Neoantigen Vaccine: An Emerging Tumor Immunotherapy, *Molecular Cancer*, Vol. 18, No. 1, 128. doi:10.1186/s12943-019-1055-6.
531. Kaliamurthi, S., Selvaraj, G., Kaushik, A., Gu, K., and Wei, D. (2018). Designing of CD8+ and CD8+-Overlapped CD4+ Epitope Vaccine by Targeting Late and Early Proteins of Human Papillomavirus, *Biologics: Targets and Therapy*, Vol. Volume 12, 107–125. doi:10.2147/BTT.S177901.
532. Kanampalliar, A. M. (2020). Reverse Vaccinology and Its Applications, *Methods Mol Biol*, 1–16. doi:10.1007/978-1-0716-0389-5_1.
533. Bui, H.-H., Sidney, J., Dinh, K., Southwood, S., Newman, M. J., and Sette, A. (2006). Predicting Population Coverage of T-Cell Epitope-Based Diagnostics and Vaccines, *BMC Bioinformatics*, Vol. 7, No. 1, 153. doi:10.1186/1471-2105-7-153.

534. Hackl, H., Charoentong, P., Finotello, F., and Trajanoski, Z. (2016). Computational Genomics Tools for Dissecting Tumour-Immune Cell Interactions, *Nature Reviews Genetics*, Vol. 17, No. 8, 441–458. doi:10.1038/nrg.2016.67.
535. Schumacher, T. N., and Schreiber, R. D. (2015). Neoantigens in Cancer Immunotherapy, *Science*, Vol. 348, No. 6230, 69–74. doi:10.1126/science.aaa4971.
536. Gubin, M. M., Zhang, X., Schuster, H., Caron, E., Ward, J. P., Noguchi, T., Ivanova, Y., Hundal, J., Arthur, C. D., Krebber, W.-J., Mulder, G. E., Toebes, M., Vesely, M. D., Lam, S. S. K., Korman, A. J., Allison, J. P., Freeman, G. J., Sharpe, A. H., Pearce, E. L., Schumacher, T. N., Abersold, R., Rammensee, H.-G., Melief, C. J. M., Mardis, E. R., Gillanders, W. E., Artyomov, M. N., and Schreiber, R. D. (2014). Checkpoint Blockade Cancer Immunotherapy Targets Tumour-Specific Mutant Antigens, *Nature*, Vol. 515, No. 7528, 577–581. doi:10.1038/nature13988.
537. Sahin, U., Derhovanessian, E., Miller, M., Kloke, B.-P., Simon, P., Löwer, M., Bukur, V., Tadmor, A. D., Luxemburger, U., Schrörs, B., Omokoko, T., Vormehr, M., Albrecht, C., Paruzynski, A., Kuhn, A. N., Buck, J., Heesch, S., Schreeb, K. H., Müller, F., Ortseifer, I., Vogler, I., Godehardt, E., Attig, S., Rae, R., Breitkreuz, A., Tolliver, C., Suchan, M., Martic, G., Hohberger, A., Sorn, P., Diekmann, J., Ciesla, J., Waksman, O., Brück, A.-K., Witt, M., Zillgen, M., Rothermel, A., Kasemann, B., Langer, D., Bolte, S., Diken, M., Kreiter, S., Nemecek, R., Gebhardt, C., Grabbe, S., Höller, C., Utikal, J., Huber, C., Loquai, C., and Türeci, Ö. (2017). Personalized RNA Mutanome Vaccines Mobilize Poly-Specific Therapeutic Immunity against Cancer, *Nature*, Vol. 547, No. 7662, 222–226. doi:10.1038/nature23003.
538. Ott, P. A., Hu, Z., Keskin, D. B., Shukla, S. A., Sun, J., Bozym, D. J., Zhang, W., Luoma, A., Giobbie-Hurder, A., Peter, L., Chen, C., Olive, O., Carter, T. A., Li, S., Lieb, D. J., Eisenhaure, T., Gjini, E., Stevens, J., Lane, W. J., Javeri, I., Nellaippan, K., Salazar, A. M., Daley, H., Seaman, M., Buchbinder, E. I., Yoon, C. H., Harden, M., Lennon, N., Gabriel, S., Rodig, S. J., Barouch, D. H., Aster, J. C., Getz, G., Wucherpfennig, K., Neuberger, D., Ritz, J., Lander, E. S., Fritsch, E. F., Hacohen, N., and Wu, C. J. (2017). An Immunogenic Personal Neoantigen Vaccine for Patients with Melanoma, *Nature*, Vol. 547, No. 7662, 217–221. doi:10.1038/nature22991.
539. Hundal, J., Kiwala, S., McMichael, J., Miller, C. A., Xia, H., Wollam, A. T., Liu, C. J., Zhao, S., Feng, Y.-Y., Graubert, A. P., Wollam, A. Z., Neichin, J., Neveau, M., Walker, J., Gillanders, W. E., Mardis, E. R., Griffith, O. L., and Griffith, M. (2020). PVACTools: A Computational Toolkit to Identify and Visualize Cancer Neoantigens, *Cancer Immunology Research*, Vol. 8, No. 3, 409–420. doi:10.1158/2326-6066.CIR-19-0401.
540. Bjerregaard, A.-M., Nielsen, M., Hadrup, S. R., Szallasi, Z., and Eklund, A. C. (2017). MuPeXI: Prediction of Neo-Epitopes from Tumor Sequencing Data, *Cancer Immunology, Immunotherapy*, Vol. 66, No. 9, 1123–1130. doi:10.1007/s00262-017-2001-3.
541. Schenck, R. O., Lakatos, E., Gatenbee, C., Graham, T. A., and Anderson, A. R. A. (2019). NeoPredPipe: High-Throughput Neoantigen Prediction and Recognition Potential Pipeline, *BMC Bioinformatics*, Vol. 20, No. 1, 264. doi:10.1186/s12859-019-2876-4.
542. Reynisson, B., Alvarez, B., Paul, S., Peters, B., and Nielsen, M. (2020). NetMHCpan-4.1 and NetMHCIIpan-4.0: Improved Predictions of MHC Antigen Presentation by Concurrent Motif Deconvolution and Integration of MS MHC Eluted Ligand Data, *Nucleic Acids Research*, Vol. 48, No. W1, W449–W454. doi:10.1093/nar/gkaa379.543. O'Donnell, T. J., Rubinsteyn, A., Bonsack, M., Riemer, A. B., Laserson, U., and Hammerbacher, J. (2018). MHCflurry: Open-Source Class I MHC Binding Affinity Prediction, *Cell Systems*, Vol. 7, No. 1, 129–132.e4. doi:10.1016/j.cels.2018.05.014.
544. Bassani-Sternberg, M., Bräunlein, E., Klar, R., Engleitner, T., Sinitcyn, P., Audehm, S., Straub, M., Weber, J., Slotta-Huspenina, J., Specht, K., Martignoni, M. E., Werner, A., Hein, R., H. Busch, D., Peschel, C., Rad, R., Cox, J., Mann, M., and Krackhardt, A. M. (2016). Direct Identification of Clinically Relevant Neopeptides Presented on Native Human Melanoma Tissue by Mass Spectrometry, *Nature Communications*, Vol. 7, No. 1, 13404. doi:10.1038/ncomms13404.
545. Abelin, J. G., Harjanto, D., Malloy, M., Suri, P., Colson, T., Goulding, S. P., Creech, A. L., Serrano, L. R., Nasir, G., Nasrullah, Y., McGann, C. D., Velez, D., Ting, Y. S., Poran, A., Rothenberg, D. A., Chhangawala, S., Rubinsteyn, A., Hammerbacher, J., Gaynor, R. B., Fritsch, E. F., Greshock, J., Oslund, R. C., Barthelme, D., Addona, T. A., Arieta, C. M., and Rooney, M. S. (2019). Defining HLA-II Ligand Processing and Binding Rules with Mass Spectrometry Enhances Cancer Epitope Prediction, *Immunity*, Vol. 51, No. 4, 766–779.e17. doi:10.1016/j.immuni.2019.08.012.
546. Chong, C., Coukos, G., and Bassani-Sternberg, M. (2022). Identification of Tumor Antigens with Immunopeptidomics, *Nature Biotechnology*, Vol. 40, No. 2, 175–188. doi:10.1038/s41587-021-01038-8.
547. Yadav, M., Jhunjhunwala, S., Phung, Q. T., Lupardus, P., Tanguay, J., Bumbaca, S., Franci, C., Cheung, T. K., Fritsche, J., Weinschenk, T., Modrusan, Z., Mellman, I., Lill, J. R., and Delamarre, L. (2014). Predicting Immunogenic Tumour Mutations by Combining Mass Spectrometry and Exome Sequencing, *Nature*, Vol. 515, No. 7528, 572–576. doi:10.1038/nature14001.
548. Keskin, D. B., Anandappa, A. J., Sun, J., Tirosh, I., Mathewson, N. D., Li, S., Oliveira, G., Giobbie-Hurder, A., Felt, K., Gjini, E., Shukla, S. A., Hu, Z., Li, L., Le, P. M., Allesøe, R. L., Richman, A. R., Kowalczyk, M. S., Abdelrahman, S., Geduldig, J. E., Charbonneau, S., Pelton, K., Iorgulescu, J. B., Elagina, L., Zhang, W., Olive, O., McCluskey, C., Olsen, L. R., Stevens, J., Lane, W. J., Salazar, A. M., Daley, H., Wen, P. Y., Chiocca, E. A., Harden, M., Lennon, N. J., Gabriel, S., Getz, G., Lander, E. S., Regev, A., Ritz, J., Neuberger, D., Rodig, S. J., Ligon, K. L., Suvà, M. L., Wucherpfennig, K. W., Hacohen, N., Fritsch, E. F., Livak, K. J., Ott, P. A., Wu, C. J., and Reardon, D. A. (2019). Neoantigen Vaccine Generates Intratumoral T Cell Responses in Phase Ib Glioblastoma Trial, *Nature*, Vol. 565, No. 7738, 234–239. doi:10.1038/s41586-018-0792-9.